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Moringa olifera leaf extract increases physio-biochemical properties, growth and yield of *Pisum sativum* grown under salinity stress

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Abstract

The soil salinity is a dominant abiotic problem in arid and semi-arid region of the world. In these areas it occurs natural due low rainfall to leech salts from soil, aided by human activities due to excessive irrigations, over-use of fertilizers and poor drainage. The purpose of this research was to elaborate the toxic effect of salt stress to pea plants and to determine mitigating role of exogenously applied *Moringa olifera* leaf extract (MLE) under salinity stress. The variety of pea known as "Pea-2009" was put under salt stress at concentrations of 0 and 100 mM NaCl and treated with MLE extract at concentrations of 0 and 3% as a priming and foliar approach. Under salinity stress, the results of this study showed a decrease in plant growth (fresh and dry biomass of shoot and root s), leaf relative water contents, chlorophyll a and b, carotenoid, TFAAs, proline, K^+, and antioxidants (POD, APX, and CAT), whereas the application of MLE either (seed priming/foliar spray) enhanced all studied attributes under both checked and control conditions. On the other hand, with salinity stress, the levels of Na^+, MDA, and H_2O_2 rose, but the MLE treatment decreased the levels of Na^+, MDA, and H_2O_2 particularly under salt stress. Salt treatment resulted in a substantial decrease in yield (number of pods plant-1, pod weight, and seed weight), while MLE fertigation increased the production of pea plants. Therefore, exogenous application of *MLE* potentially enhance growth of yield of economically important crops grown under stressful environmental conditions like salinity by maintaining better physio-biochemical indices.

Keywords: Moringa leaf extract, superoxide dismutase, total soluble protein, Proline, seed weight

1. Introduction

The world population has plaaced increased demand on resources including food, water and energy. It is predicted that human population will mount to 9.7 billion by 2050 which is a serious problem to food security (Nadathur et al. 2017). There must be a major concern of all

nations to develop proper strategy to cope with upcoming food insecurity through better use of agricultural soils which are becoming barren due to increase in the intensity of environmental threats like drought and salinity. Soil salinity is the major environmental stress that is spreading speedly due to excessive use of saline water that converts valuable agricultural arable land into a salinized wasteland (Kaya et al. 2020).

Plants grown under saline environment are forced to accumulate salts to toxic levels into their cellular system which is the prime cause of oxidative stress leading to accumulation of reactive oxygen species (ROS) to toxic levels, which hampers plants' normal growth by upsetting physio-biochemical processes including photosynthesis (Mansoor et al., 2022). The plants under saline conditions experience physiological drought that cause; i) water shortage in rhizophere resulting in lowered root water potential; ii) phytotoxicity by excessive ions i.e., Na^+ and Cl^- and iii) nutritional imbalance through hindring in the uptake of other nutrients *i.e.* K^+, Ca^{++}, N, P *etc* (Akhter et al. 2021), thus hampering plant normal vegetative and reproductive growth.

The synthesis of ROS is natural process in plants however, under adverse environmental conditions like salinity the production of ROS like superoxide radicle, singlet oxygen, H_2O_2 and OH^- radicle is augumented (Mansoor et al., 2022). These increased ROS level stimulate the phytotoxic reaction in plants such as protein degradation, membrane depolarization, lipid peroxidation, DNA mutation, and protein degradation (Mansoor et al. 2022). Plants under such adversities has develop a counter defense mechanism through enhanced antioxidant activity and organic osmolyte accumulation (Ahanger et al., 2020)

including catalase (CAT), peroxidases (POD), superoxide dismutase (SOD), ascorbate peroxidase (APX) and proline to quench excessive ROS (Mansoor et al., 2022). Similarly, amino acids, proteins and sugars also supports the plants for efficient detoxification of salinity-induced ROS (Ahanger et al., 2020).

Fertigation with plant extract can efficiently improve plant growth and yield, however their stimulatory or inhibitory effect is totally based on source, type of applied concentration, dose and stage at which the extract is applied to of plant. Moringa (*Moringa oleifera*) leaf extract is rich source of vitamins *i.e*. A and C, phenolics, calcium, iron and carotenes (Ademiluyi et al. 2018). Despite having high level of antioxidants, total free amino acids (TFAAs), soluble sugars and sufficient amount of phytohormones like cytokinins (Nouman et al. 2016), but there are scanty reports which focusing on amelioration of salt stress by combined application (foliar and seed priming) of moringa extract in vegetable crops.

Pisum sativum L., the common pea (locally known as garden or field pea), is an herbaceous annual and cool season crop in the Fabaceae (formerly Leguminosae) family native to Mediterranean basin and near east (Macák et al. 2020). In Pakistan, pea important vegetable and was cultivated at 10479 hectors yielding upto 71793 tons with an average of 6.9 tons/hec. In Pakistan pea is ranked as 3rd familiar crop and Punjab is the major contributor (71%) of the total production (Aslam et al. 2000; Ullah et al. 2020). The current study was aimed to focus on: (1) impact of salinity stress on growth attributes, mineral uptake, photosynthetic pigments, compatible solutes, antioxidant level and relative water contents (RWC) of pea plants, (2) role

of *MLE* in amelioration of salinity stress in pea (3) to compare the response of *MLE* (foliar applied and seed soaking) to salt stressed pea.

2. Materials and Methods

The seeds of Pea (*Pisum sativum* var. Pea-2009) were collected from Ayub Agriculture Research Institute (AARI), Faisalabad and experiment was carried out at IP and AB Bahauddin Zakariya University Multan, Pakistan in grwoing season October – December 2019-20. The average growing conditions (October – December 2019-20) were recorded as rainfall 10 mm, wind 8 Kmph, relative humidity 30 %, clouds 9 %, minimum temperature 19°C and maximum temperature 30°C. The plastic pots were arrangened used in this experiment is given in Fig. A. The pots were filled with equil wight 8kg river sand (double washed with distilled water) and holes with diameter of 1.5 cm were made at the bottem of each pot in order to remove extra water. The healthy seeds were surface seterlized with sodium hypochlorite for 15 minutes follwed by drying in the shade. Seven seeds were sown in each pot during winter season on December 2019. After complete germination, seedlings were thinned to three plants per pot. The plants were irrigated with full strength Hoagland and Arnon (1950) nutrient solution in order to ensure the availability of all the nutrients required for optimum plant growth.

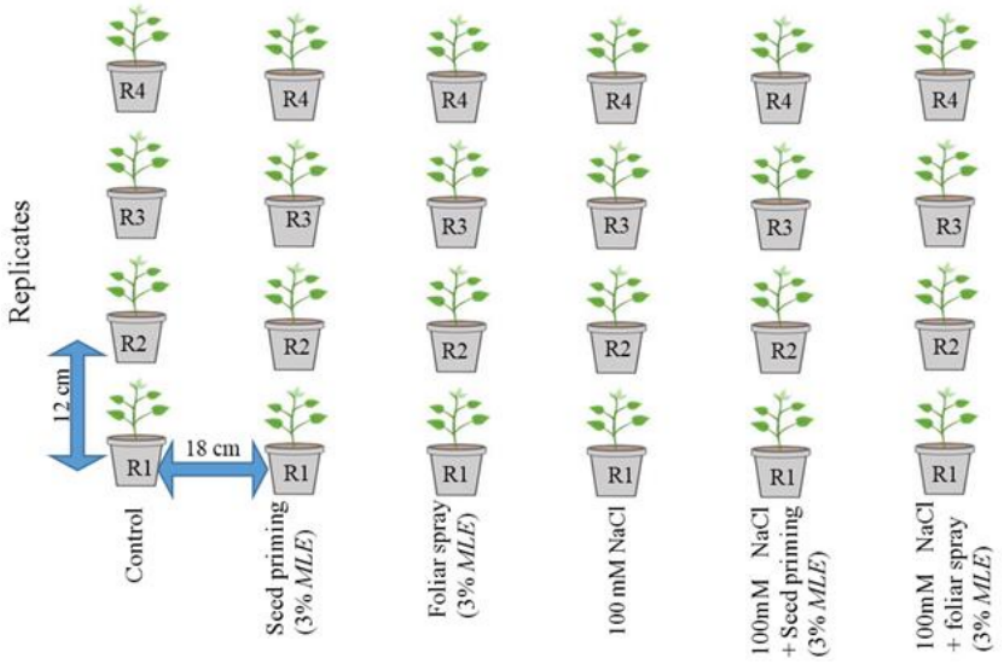

Fig. A The arrangement of pots used in this stud. The distance of pots between each treatment was 18 cm while among treatment 12 cm.

2.2 *Moringa Oleifera* leaf (*MLE*) extraction method

Leaves of *Moringa* from tender branches (18 g) were collected, washed with tap water and dried at room temperature. The leaves were grinded in pestle and motor by adding 5ml distilled water to make a paste. Filtered the paste via Whatman No.1 filter paper to obtain a clear *MLE* extract. Finally, 30 ml *MLE* was added to 970 ml distilled (1:30) to make ready to use *MLE*. To ensure optimal penetration in leaf tissues during foliar spray, surfactant tween-20 was used @ 100ml/ 1000ml *MLE* solution.

2.2 Pre-sowing Seed priming with *MLE*

MLE extract (3ml) was diluted with addition of 97 ml distilled water in a beaker to make a final volume of 1L. Seeds were dipped in beaker containing 3% *MLE* solution, covered with aluminium foil and placed overnight. Soaked seeds were sown on next day after treatment of 24h.

2.3 Foliar application with *MLE*

Solution prepared (3ml *MLE* / 97 ml distilled water) was foliarly applied to young plants at 3rd week of sowing. To ensure an equal level of foliar spray (10 ml for each seedling), a plant sprayer with an equal sized nozzle tip was used.

2.4 Application of salinity stress

The salt treatment was applied to plant after 21 days of sowing. The salinity stress was developed by adding weighed quanitity of NaCl (National Refined) to measured volume of tap water (For 100 mM salt stress 5.84g of NaCl was added to 1 litter water).

Following data was collected during the course of experimentation.

2.5 Morphological attributes

When the plants were of 48 days, a plant was carefuuly uprooted from each pot to record shoot and root length and fresh weights. The dry biomass of shoot and root was recorded after oven drying of these samples.

2.6 Leaf relative water contents (LRWC)

A mature leaf from each replicate was trimmed and fresh weights (FW) were recorded immediately. The leaf samples were placed in distilled water from their cut side and turgid weights (TW) were recorded after 10 hours. These leaves were then oven dried, and dry weights

(DW) were recorded Smart and Bingham (1974) method. Following formula was used for measuring LRWC.

$$LRWC = [(FW-DW)/(TW-DW)] \times 100$$

2.7 Photosynthetic pigments

Acetone based extraction; detection at 480, 645 and 663 nm using spectrophotometer and calculating with equations devised by Lichtenthaler (Lichtenthaler and Wellburn 1983) was used for estimation of concentrations of photosynthetic pigments.

2.8 Total soluble protein (TSP)

The fresh flag leaf samples (0.5 g) from ach replicate were homogenized in sodium phosphate buffer (8 mL, pH 7.8) and centrifuged at 15000 rpm for 12 minutes at 4 °C. The 0.1 mL of supernatant thus removed was poured in test tubes containing 5 mL Bradford reagent. The reading was taken at 595 nm using spectrophotometer after 15 minutes of incubation at room temperature (Bradford 1976). TSP contents were determined using the standard curve

2.9 Proline

For proline estimation, plant samples (0.5 g) were grinded in 10 mL 3% sulfosalicylic acid solution and were filtration with Whatman filter paper. 2 mLof extract sample was added to 2 mL ninhidrin and 2 mL of glacial acetic acid solution in test tubes which were water bathed at 100 °C for 60 minutes followed by immediate cooling in ice. After cooling, 4.0 mL of toluene was poured in these test tubes, mixed vigorously and kept at room temperature till two layers were formed. The absorbance of upper colored layer was taken at 520 nm (Bates et al. 1973).

2.10 Hydrogen peroxide (H_2O_2) contents

To assay H_2O_2 contents plant material (0.25 g) was homogenized in 5 mL TCA (0.1%) solution and centrifuged for 15 minutes at 12,000 rpm. The supernatant (0.5 mL) was mixed with 0.5 mL sodium phosphate buffer and 1 mL of potassium iodide (KI) solution in test tubes. Test tubes were vortexed and absorbance was read at 390 nm (Velikova et al. 2000).

2.11 Malondialdehyde (MDA) contents

The MDA contents were estimated using Heath and Packer (Cakmak and Horst 1991) methodology. 1 mL of supernatant (same as used in protein estimation) was mixed to 1 mL TBA (0.5%) solution prepared in 20% TCA solution in test tubes and were water bathed for 30 minutes at 95 °C. The tubes were then ice bathed for 5 minutes followed centrifugation at 6,000 rpm. The absorbance was recorded at 532 nm and 600 nm. The extension coefficient (155 mM^{-1} cm^{-1}) was used for MDA contents calculation.

2.12 Antioxidants

The SOD activities in leaves and roots was determined through quantifying the inhibition in photo reduction of nitrobluetetrazolium (NBT), the protocol devised by Beauchamp and Fridovich (Beauchamp and Fridovich 1971). The reaction solution for POD contained 100 μL 30 mM H_2O_2, 100 μL guaiacol and 100 μl of enzyme extract (supernatant) into 2.7 mL sodium phosphate buffer (Chance and Maehly 1955). For estimation of CAT activity same reaction solution as for POD (except guaiacol) was used. The absorbance of POD and CAT samples was observed on time scan (0-60 second) at 470 and 240 nm respectively using spectrophotometer, CAT (Aebi 1984). The activity of APX was determined using Nakano and Asada (Nakano and Asada 1981) methodology. The reaction solution contained 100 μL

ascorbate solution (10 mM), 100 μl H_2O_2 (30%) and 100 μl enzyme extract (supernatant) into 2.7 mL of sodium phosphate buffer. After a gentle shake, the absorbance was read at 290 nm with on time scan (0-60 second) using spectrophotometer.

2.13 ionic (Na^+ and K^+) estimation

Plant samples (shoots and roots) were taken in digestion flasks containing 2 mL of digestion mixture and were kept overnight at 25 ºC. The flasks were then transferred to hot plate set at 250 ºC. Samples were heated till fume formation. At this point 0.5 ml of $HClO_4$ was added in each flask and again transferred these flasks to hot plate at 250 ºC for almost 2 hours until discoloration of samples. After digestion the samplrs were filtred and volume was raised to 50ml with dH2O to be used for estimation of ionic concentration (Allen et al. 1985). The Na^+ and K^+ concentrations in samples were estimated using flame photometer (Jenway-PFP7, United Kingdom).

2.14 Yield attributes

At maturity all the pods from each plant / pot were removed and the data for number of pods per plant, weight of pods per plant, number of seed per plant and weight of seeds per plant were recorded.

2.15 Statistical analysis

The statistical analysis (Two way ANOVA) was performed using SPSS-20 (SPSS Inc. Chicago, IL, USA). To conclude the influence of various treatments, R-sutudio v 4.0.4 was employed. The correlation matrix was designed for estimation of overall relationship of different traits of pea plants under different regimes. Moreover, the data was subjected to

principle component analysis (PCA) to decipher influence of different treatments on various plant parameters using Origin (v.2021). The clustered heatmaps between growth, yield and various plant physiological parameters were then constructed to assess the association among studied traits.

3. Results

3.1. Growth parameters

ANOVA for biomass accumulation (shoot and root fresh and dry biomass) of pea displayed a significant ($P < 0.001$) reduction in these parameters under salt stress, the effect of *MLE* was also highly significant ($P<0.001$) on growth of pea plants (Table 1). Salinity stress reduced shoot fresh biomass (33%), shoot dry biomass 52%), root fresh biomass (35%) and root dry biomass (37%) when compared to non-saline pea plants. However, the application of *MLE* either as seed priming or foliar application enhanced growth under control and saline conditions. Under NaCl stressed conditions, the foliar application of *MLE* enhanced fresh and dry biomass of shoot (26%. and 67%) and root (21% and 35%) respectively. Similarly, seed priming with *MLE* also enhanced fresh and dry weights of shoot (13% and 47%) and root (14% and 20%) respectively (Table 1). It has been observed that the impact of foliar application of *MLE* was more promising in enhancing growth.

3.2. Mineral elements

A significant ($P < 0.001$) increase in shoot and root Na^+ contents was perceived, while K^+ contents were reduced significantly ($P < 0.0001$). The application of *MLE* either foliar spray

or seed priming reduced Na^+ contents while enhanced K^+ contents in roots and shoots of pea plant (Table 2). Salinity stress enhanced root and shoot Na^+ contents to 42% and 212% respectively, while *MLE* application reduced Na^+ contents to 28% (foliar) and 35% (priming) in roots and 142% (foliar) and 191% (priming) in shoots respectively. The root and shoot K^+ contents were decreased to 62% and 6% respectively with NaCl treatment, on the other hand *MLE* application enhanced K^+ contents to 57% (foliar) and 65% (priming) in roots and 18% (foliar) and 30% (priming) in shoots respectively

3.3. Photosynthetic pigments

The data showed that Chl-a, Chl-b, T. Chl. and carotenoid contents were lowered in pea plants under salinity stress. As compared to control salinity stress lowered Chl.a (28%), T. Chl. (8%) and carotenoids (15%) contents, however Chl.b contents were enhanced (39%) under salinity stress (Fig. 1). The foliar application of *MLE* enhanced Chl.a (22%), Chl.b (23%), T.Chl. (22%) and carotenoids (14%) contents under salinity stress as compared to control. But on the other hand, seed priming with *MLE* caused a considerable reduction. Under salinity Chl.a (13%), T.Chl. (3%) and carotenoids (11%) contents when compare to salinity stressed plants (Fig. 1). The Chl. a/b concentration was remarkably decreased (48% and 27%) under salinity stress and when *MLE* was applied as seed priming respectively, while foliar spray of *MLE* maintained Chl. a/b ratio under control as well as saline environment. The chlorophyll / carotenoid were enhanced to 9%, 7% and 10% with salinity treatment, foliar and seed priming *MLE* under salinity stress respectively (Fig. 1).

3.4. Total soluble protein (TSP) and free amino acid (TFAA) contents

The application of *MLE* caused a remarked enhancement in TSP and TFAA in pea plants. Though, the response of TSP to salt stress was non-significant however, TFAA showed significant effect (Fig. 2). The data revealed that TSP contents were reduced 6% with NaCl application while *MLE* application enhanced these contents to 10% and 4% with foliar and seed priming with *MLE* respectively under salinity stress. However, TFAA contents were declined (53%) due to salinity stress, however, the application of *MLE* either applied foliar or seed priming enhanced these contents to 82% and 97% respectively in pea plants (Fig. 2).

3.5. Antioxidant response

The data depicted that salinity stress significantly ($P<0.001$) effect the accumulation of different antioxidants (CAT, POD, APX, proline) in pea plants. The ameliorative effect of *MLE* was also significant ($P<0.001$) showing a positive enhancement in the accumulation of these antioxidants (Fig. 3). The salinity-induced decrease in CAT (16%), POD (31%), APX (10%) and proline (15%) contents was observed in leaves of pea plants. Under non-saline conditions *MLE* fertigation caused enhancement in CAT (26% (foliar) and 0.5% (seed priming); POD (43% (foliar)}, APX (10% (foliar) and 3% (priming) and proline (29% (foliar) and 28% (priming) contents in leaves of pea plants. On the other hand, *MLE* fertigation either as foliar spray and seeds priming enhanced CAT (35%, 29%), POD (51%, 160%), APX (40%, 57%) and proline (119%, 197%) contents respectively in pea plants under salinity stress (Fig. 2).

3.6. Leaf relative water content (LRWC)

The imposition of salinity to growing media lowered (12%) relative water contents of pea leaves. However, these contents were enhanced (7%) by foliar application of *MLE* while the impact of *MLE* as seed priming was not significant when compared to control plants (Fig.3).

3.7. Malondialdehyde (MDA) contents

The ANOVA for MDA contents revealed that these contents were significantly enhanced under salinity stress but were reduced with fertigation of *MLE* either as foliar spray or seed priming (Fig. 3). MDA contents were enhanced to 10% with NaCl treatment while *MLE* application reduced these contents to 16% and 15% when applied as foliar or seed priming respectively (Fig. 3).

3.8. Hydrogen peroxide (H_2O_2) contents

A pronounced rise in H_2O_2 contents was observed in pea under salinity stress. The increase in H_2O_2 contents was 12%, 30% and 26% under control, foliar and seed priming (*MLE*) conditions respectively (Fig. 4). However, when compared to salinity stress alone the foliar spray caused 16% and seed priming caused (12%) decrease in MDA contents under salinity stress (Figure 3).

3.9. Yield attributes

The statistical analysis revealed that factors, salinity and *MLE* application caused remarked effect on yield (number of pods plant^{-1}; pod weight plant^{-1}, number of seeds plant^{-1} and wt. of seeds plant^{-1}) of pea plants. The effect of salinity × *MLE* showed that *MLE* caused a differential effect of yield of pea plants at salinity levels (Table 1). Highest yield either under

control or saline conditions was produced by foliar application of *MLE* showing that foliar spray posed more promising effect on yield (Table 1).

3.10. Multivariate analysis

3.10.1. Correlation matrix

A correlation matrix among morphological attributes and shoot ionic contents represent significant correlation ($P \leq 0.01$). A strong positive correlation was found between TWP: RFW, TWP: SFW, TWS: SFW, RDW: PH and TNS: TWP, whereas a week associations was seen among R/SNa: TNS, R/SNa: TWP and R/SK: PH. The SK, TNP, RNa and SNa showed negative correlations (Fig. 4A). In case of correlation drawn among physiological attributes, Pro: APX, TChl: Chla, Chla: Chla/C showed positive correlation, while MDA, H_2O_2, POD and RWC represent negative correlation (Fig.4B).

3.10.2. Principal component analysis (PCA-Biplot)

Principal component analysis showed a strong influence of *MLE* foliar application as well as seed soaking under control environment (S1T1 and S1T2) on PH, TFAA and TNS of *Pisum sativum*, while RNa affected H_2O_2 and MDA level of plants subjected to saline condition (S2T0). The SNa imposed strong impact on TNP and Pro content of foliar treated slots (S2T1), while Sk had strong influence on POD, TSP, APX and Chl/C in seed priming slots (S2T2) growing under stressed environment (Fig.5A).

Principal component analysis based on mode of application represented two distinct groups (Fig. 5B). Salinity controlled plants (S-0%) showed association with TFAA, PH, TNS,

R/SNa and RK. Salinity treated plants (S-100%) exhibited close association with SNA, H2O2, RNa and MDA. Traits like POD, SK, TSP, APX, Pro, TNP, Chl/C and CAT were influenced in both control and salinity treated plants.

3.10.3. Clustered heatmaps

A clustered heatmap was established among morphological and shoot ionic contents to assess their responses under different salinity and *Moringa* leaf extract (*MLE*) treatments. The TWP, SFW, SDW, RFW and PH showed positive association and strong clustering in S1T1, RDW and TWS in S1T2, whereas RNa and SNa showed high clustering in S2T0 treatment (Fig. 6A). The clustered heatmap on physiological attributes represented close association of Chla, Chlb, TChl, Caro and TSP in S2T1, APX, Pro and POD in S2T2, while TFAA and RWC in S1T1 treatment (Fig. 6B).

4. Discussion

Increasing soil salinity is creating alarming situation for agricultural system across the world. According to current soil salinization rate, it may reduce up to 50% crop production in coming future (Singh 2022). The current situation can be reduced by applying short term procedure to improve productivity of crop especially under hostile environment (Noreen et al. 2021). The application of bio-stimulants like micronutrients, leaf extracts, hormones, vitamins and osmoprotectants are commonly used to induce abiotic stress tolerance in crop plants (Zouari et al. 2019). The current study revealed that fertigation of pea plants and seeds with Moring leaf extract (*MLE*) as foliar and seed priming mitigated salinity stress. It has been previously studied by several researchers that *MLE* can potentially mitigate salinity stress and

tends to maintain optimum growth and productivity (Arif et al. 2022; Khan et al. 2020; Yasmeen et al. 2013a). *MLE* used as potential bio-stimulator or bio-enhancer in having some important minerals, phenolics, alkaloids, sugars and vitamins that support plants plant growth and developmental process (Arif et al. 2022). The current study was aimed to examine the possible mechanism adopted by pea plants in response to salt stress *via* foliar spray and seed priming with *MLE* in a saline environment.

A major reduction growth (biomass production) of pea (*Pisum sativum* L.) was observed under salinity stress that was linked to reduced yield (number and weight of pods) attributes. The decrease in the activities of cell division and cell elongation due to excessive production of ROS in cellular system is evedent under salinity stress. Application of *MLE* as foliar and seed priming not only lessened the salinity's noxious effects on growth and yield attributes but also enhanced growth rate considerable when applied alone or in combination of salt stress. As it was observed in earlier reports of (Howladar 2014) in *Phaseolus vulgaris* and Yasmeen et al. (2013a) in primed wheat seeds to *Moringa* leaf extract (*MLE*) under saline condition. Plants extracts (fresh and/or dry) also have stimulatory impacts on growth by upregulating enzymes activity, metabolic contents and water use efficiency (Arif et al. 2022).

Sodium content significantly increased whereas K^+ content decreased in root and shoot of pea plants under salinity stress (Table 2). However, application of *MLE* as foliar and seed priming reduced Na^+ content and root/shoot Na^+/K^+ ratio while enhanced K^+ content in root and shoot under stress as compared to control. These results are in line of findings of (Nouman et al. 2012; Ragab et al. 2022; Yasmeen et al. 2013a). The K^+ efflux from plant organs and Na^+

accumulation are consequences of salt prone conditions, which significantly influence the cytosolic ion homeostasis and plant survival, which regarded as fundamental salt tolerance mechanism of plants (Desoky et al. 2019; Fu et al. 2022).

Salinity-induced reduction in photosynthetic pigment synthesis is subjected to several factors including mineral imbalance and reduction in the activity of several photosynthetic enzymes (Ondrasek et al. 2022). However, *MLE* fertigation either as foliar or as seed priming enhanced the chlorophyll (chla, chlb, Tchl, chl a/b and Tchl/caro) and carotenoid contents under both controlled and saline condition which is attributed to better mineral uptake and stabilized activities of photosynthetic enzyme. Generally, foliar application of *MLE* on pea plants was found more effective in comparison of seed priming, and this finding strongly agreed with previous reports of (Attanayaka and Harris 2019) on okra and (Yaseen and Takacs-Hajos 2022) on lettuce plants that *MLE* application as foliar pose positive effect on photosynthesis under both normal as well as saline condition. Moreover, enhanced carotenoid content and Total chlorophyll/ carotenoid ratio due to *MLE* application (foliar and seed priming) again play critical role in protecting macromolecules included DNA, proteins and RNA from free radicals produced in response to osmotic stress (Khan et al. 2020; Nouman et al. 2012).

A substantial increase in hydrogen per oxide (H_2O_2) and Malonaldehyde (MDA) contents was observed under salinity stress in pea plants. Similarly, increased level of total soluble protein (TSP), total free amino acid (TFAA) and proline in response to foliar applied and/or seed priming with *MLE* could be crucial for osmoregulation and membrane stability under salt stress, as reported by (Basu et al. 2022). Osmoprotectant accumulation is considered as initial

protective strategy of plants in order to prevent from high salinity (Kaya et al. 2020). Presence of phenolic, alkaloids and other active ingredients with in *MLE* may additionally prevent from membrane leakage and structures de-stability in consequence of lipid peroxidation as was observed in common beans (Howladar 2014).

Salinity-induced ROS stimulates oxidative damage and lipid peroxidation (Mansoor et al. 2022) in plants which can be radially controlled by increasing activity of antioxidants (enzymetic and non-enzymetic) *via* application of bio-stimulant or growth promoters. Among enzymatic antioxidants CAT, POD, and APX worked as front line defense rescue plants from stressful conditions like salinity and drought (Noreen et al. 2021). In response to salt stress, the removal of extra ROS, through *MLE* application as foliar and/ or seed priming cause further increase in activity of APX, , CAT and POD in order to strength the defense system in plants via removal of reactive ROS (Yasmeen et al. 2013b). Both APX and CATare vital for empowering stress tolerance, as they quickly quench the H_2O_2 and ultimately lead to protection of membrane functioning. So the high level of APX, CATand POD APX in *MLE* treated pea plants (foliar and seed primed) indicated the improved stress tolerance to oxidative damage (Hasanuzzaman and Fujita 2022). Such upraise in enzymetic antioxidant activity by foliar and/or seed priming of *MLE* could be beneficial for growth maintenance through rapid quenching of ROS. In concluding, the application of *MLE* as foliar as well as seed priming further strengthened the both enzymetic and non-enzymetic antioxidant in *Pisum sativum* and similar remarks have also been represented by Zulfiqar et al. (2020) and Merwad (2018) in plant species subjected to salt stress.

5. Conclusions

The seed priming or foliar application of Moringa leaf extract (*MLE*) can potentially develop better tolerance to salinity stress in Pea (*Pisum sativum*) as was observed in this experiment. Although pea is sensitive to salinity stress still the application of *MLE* either as seed priming or foliar spray enhanced growth and yield of pea plants especially under salinity stress. The production of excessive ROS, accelerated lipid peroxidation (MDA), accumulation of Na^+ to toxic levels in plant tissues and disturbance in biochemical (antioxidant) response of pea reduced the photosynthetic ability of plats resulting in reduced growth and development are inevitable events under salinity stress. However, *MLE* balanced ionic contents, regulated antioxidant response and stabilized photosynthetic activity to cope the adversities of salinity induced ROS. Therefore, *MLE* fertigation either as seed priming or foliar spray is recommended as remedy for plants grown under saline conditions.

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Tables and Figures legend

Table 1. Effect of MLE as seed priming and foliar spray on shoot and root fresh and dry weights (g), number of pods / plant, weight of pods / plant (g), number of seed / plant and weight of seeds / plant (g) of Pea 2009 (*Pisum sativum* L.) at 0 and 100 mM NaCl stress.

Table 2. Effect of MLE as seed priming and foliar spray on root and shoot Na^+ and K^+ contents (mg g^{-1} d.wt.) of Pea 2009 (*Pisum sativum* L.) at 0 and 100 mM NaCl stress.

Fig. A The arrangement of pots used in this stud. The distance of pots between each treatment was 18 cm while among treatment 12 cm.

Fig. 1. Effect of MLE as seed priming and foliar spray on chlorophyll-a (A), chlorophyll-b (B), total chlorophyll (C), carotenoid (D) contents (mg g^{-1} fw), chlorophyll a/b (E) and chlorophyll / carotenoid (F) ratio of Pea 2009 (*Pisum sativum* L.) at 0 and 100 mM NaCl stress. ΔNaCl and ΔMLE corresponds to percentage decrease / increase with respect to control after application of NaCl and MLE respectively.

Fig. 2. Effect of MLE as seed priming and foliar spray on total soluble protein (mg g^{-1} fw) (A), total free amino acid (mg g^{-1} fw) (B), CAT (U mg^{-1} protein fw) (C), POD (U mg^{-1} protein fw) (D), APX (U mg^{-1} protein fw) (E) and proline (U mg^{-1} protein fw) (F) of Pea 2009 (*Pisum sativum* L.) at 0 and 100 mM NaCl stress. ΔNaCl and ΔMLE corresponds to percentage decrease / increase with respect to control after application of NaCl and MLE respectively.

Fig. 3. Effect of MLE as seed priming and foliar spray on leaf relative water contents (A), MDA (nmol g^{-1} fw) (B) and H_2O_2 (μmol g^{-1} fw) (C) of Pea 2009 (*Pisum sativum* L.) at 0 and 100 mM NaCl stress. ΔNaCl and ΔMLE corresponds to percentage decrease / increase with respect to control after application of NaCl and MLE respectively.

Fig. 4. Correlation matrix representing relationship of (A) growth, yield and shoot ionic contents (B) physio-biochemical attributes of Pea 2009 (*Pisum Sativum* L.) grown under saline environment (S1-salinity 0mm, S2-salinity 100mm) with *Moringa* leaf extract applications (T0-control, T1-foliar application, T2- seed priming).

Fig. 5. Principle component analysis representing influence of different treatments on morphological and physiological attributes of Pea 2009 (*Pisum sativum* L.): (A) Overall impact of *Moringa* leaf extract (Foliar and seed priming) and salinity on different parameters, (B) Eclipses are formed to group parameters on the basis of salinity and mode of treatments (S-0 mM, S-100 mM).

Fig. 6. Clustered heatmaps representing overall response of (A) growth, yield and shoot ionic contents (B) physiobiochemical attributes of Pea 2009 (Pisum sativum L.) grown under saline environment (S0-salinity 0mm, S1-salinity 100mm) with Moringa leaf extract applications (T0-control, T1-foliar appli-cation, T2- seed priming).

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