

supplementary

by Research Research

Submission date: 15-Nov-2023 02:05AM (UTC-0500)

Submission ID: 2228773892

File name: Anticancer_effects_of_pH.docx (41.5K)

Word count: 4640

Character count: 28475

10

Anticancer effects of pH- sensitive carvacrol zinc oxide quantum dots on DMBA induced mammary carcinoma in female sprague dawley rats

Manoj Kumar Srinivasan¹, Briska Jifrina Premnath¹, Ramya Parimelazhagan², Chandramohan Govindasamy³, Kamala-Kannan Seralathan⁴, Nalini Namasivayam*

^{1,*} Department of Biochemistry and Biotechnology, Faculty of Science, Annamalai University, Chidambaram, Tamilnadu, India.

² Department of Biochemistry, Faculty of Medicine, Sri Lakshmi Narayana Institute of Medical Sciences (SLIMS), Puducherry, India.

³ Department of Community Health Sciences, College of Applied Medical Sciences, King Saud University, P.O. Box 10219, Riyadh 11433, Saudi Arabia.

⁴ Division of Biotechnology, Advanced Institute of Environment and Bioscience, College of Environmental and Bioresource Sciences, Jeonbuk National University, Iksan 54596, South Korea.

17

Correspondence:

Dr. Nalini Namasivayam,

Professor,

Department of Biochemistry and Biotechnology,

Faculty of Science,

Annamalai University.

bionalini@gmail.com

manojkrinivasan1002@gmail.com

Abstract

Objectives: The current research explores the anticancer effects of pH sensitive CVC-ZnO QDs (Carvacrol-loaded Zinc Oxide Quantum Dots) on DMBA induced mammary carcinoma in rats. **Methods:** Female SD rats were used, and mammary cancer was induced by chemical carcinogen via subcutaneous injection near the mammary gland. Different concentrations of CVC-ZnO QDs were orally supplemented to evaluate the optimum dose. We assessed the growth rate, body weight changes, tumor volume, tumor incidence and tumor burden in both the inducer and treatment groups. We also evaluated the biochemical parameters (antioxidant status, lipid peroxidation, detoxification enzymes, and lipid profile) and histopathological changes in the kidney and mammary tissues. **Results:** Our findings indicate that CVC-ZnO QDs treated rats significantly decreased the tumor weight, incidence, burden, lipid peroxidation levels, phase I detoxification enzyme activities and increased the body weight, phase II detoxification enzyme activities, and antioxidant status compared to the DMBA alone treated rats. CVC-ZnO QDs treatment also altered the lipid profile of plasma and mammary tissue. Furthermore, histopathological results confirmed that the CVC-ZnO QDs protect against DMBA-mediated damage to the mammary and kidney. **Conclusion:** The findings indicate that the CVC-ZnO QDs administered at 4 mg/kg b.w exhibited a significant anticancer effect against DMBA-induced mammary cancer.

Keywords: Antioxidant; carvacrol; detoxification enzymes; DMBA; mammary cancer; ZnO QDs.

Highlights:

1. pH-sensitive CVC-ZnO QDs show significant anticancer effects on DMBA-induced mammary carcinoma.
2. They reduce tumor growth, improve antioxidant status, and lower lipid peroxidation.
3. CVC-ZnO QDs modulate biotransformation enzyme activities and positively impact lipid profiles.
4. They prevent DMBA-induced tissue damage in mammary and kidney tissues, indicating its potential as a safe and effective anticancer agent for mammary carcinoma.

1. Introduction

Breast cancer begins in the breast tissue and is the most prevalent cancer impacting women globally. Based on the latest statistics, breast cancer represents around 30% of newly diagnosed cancers among women (Cao et al., 2021). While there have been notable strides in diagnosing and treating breast cancer, it remains a pressing public health issue, underscoring the demand for innovative and efficient methods in both therapy and diagnosis (Anwar et al., 2020).

Quantum dots (QDs) represent nanocrystals possessing exceptional optical and electronic characteristics, rendering them highly prospective for various biomedical purposes such as cancer detection and treatment. QDs are composed of semiconductor materials that emit bright and stable fluorescent signals when excited by light, making them ideal for imaging and sensing applications (He et al., 2019).

Recent investigations have highlighted the potential of QDs for breast cancer therapy and imaging. For instance, QDs have been used for imaging breast cancer cells, which allows the detection of cancerous tissues with high accuracy and specificity (Rajitha et al., 2021). In addition, QDs have also been utilized to deliver therapeutic drugs to breast cancer cells, showing promising results in preclinical studies (Tagde et al., 2020). Edis et al. (2021) reported using QDs to deliver chemotherapeutic agents to mammary cancer cells, showing reduced toxicity and improved efficacy compared to conventional chemotherapy.

DMBA²⁸ is a potent chemical carcinogen used to induce mammary tumors in animal models. The DMBA³⁶ induced breast cancer model in female SD (Sprague Dawley) rats has been widely used to study the molecular and cellular mechanisms underlying the progression and development of mammary cancer (Kerdelhué et al., 2016). This ideal has additionally been employed to assess the effectiveness of different chemotherapy drugs and identify new therapeutic targets for treating breast cancer.

Carvacrol (CVC), a phenolic compound found in oregano, has been shown to possess anticancer properties (Li et al., 2021). Zinc oxide quantum dots (ZnO QDs), on the other hand, are nanomaterials that⁴⁷ have been investigated for their potential use in chemotherapy because of their characteristics, such as optical attributes and a significant (Jiang et al., 2018).

From our initial investigation, it has been noted that nanoformulations containing CVC-ZnO quantum dots exhibit pH-responsive behavior, leading to their release specifically in the

acidic pH conditions found within the tumor microenvironment. Consequently, this targeted drug delivery approach primarily directs therapeutic agents towards cancer cells while having a lesser impact on normal cells.

The aim of this study is to explore the anticancer potential of pH-sensitive CVC-ZnO QDs in female SD rats with DMBA-induced mammary carcinoma. This investigation entails an examination of various parameters, including tumor growth rate, biochemical analysis in both plasma and mammary tissue, as well as histological alterations in the kidney and mammary tissue.

2. Materials and methods

2.1. Chemicals

Carvacrol and DMBA were acquired from Sigma Aldrich Chemical Pvt. Ltd., while all other chemicals utilized in this study were of analytical quality.

Synthesis of CVC-ZnO QDs

ZnO QDs were prepared from zinc acetate dihydrate and sodium hydroxide precursors. They were formed by mixing the precursors in anhydrous ethanol at 60°C for 5 h, followed by centrifugation and dissolution in anhydrous ethanol. CVC was loaded onto the QDs during 12 h of continuous stirring in a colloidal suspension. The prepared CVC-ZnO QDs had spherical shape with an average crystal size of 7.64 nm.

2.2. Animal model

We purchased 36 female SD rats from the Biogen Laboratory Animal Facility in Bangalore, India, registered under CPCSEA (Reg No: 971/PO/RcBiBt/S/2006/CPCSEA). The rats were between 6 and 8 weeks old and weighed 100g and 120g. These rats were housed in the Central Animal House, RMMC (Rajah Muthiah Medical College), Annamalai University. Prior to initiating the experiment, we ensured compliance with the guidelines by securing the necessary approval from the Institutional Animal Ethics Committee for the Supervision and Administration of Experimental Animals (IAEC Proposal No: AU-IAEC/1321/6/22). The rats were acclimated to the laboratory conditions, which included controlled humidity (50±10%), a temperature of 24±2°C and a 12 h (light/dark cycle). They were provided with stranded feed (Pellet diet- cereals, animal and vegetal proteins vitamins & minerals) and water throughout the study.

2.3. Induction mammary carcinoma and CVC-ZnO QDs preparation

Mammary carcinoma was induced in rats by administering a single dose of DMBA (25 mg/kg b.w) through an emulsion of physiological saline (0.25 ml) and sunflower oil (0.75 ml). CVC-ZnO QDs were suspended in 0.1% DMSO.

2.4. Experimental design

Thirty-six female SD rats were randomly divided into 6 groups, each consisting of six. The experimental procedure conducted in this study is illustrated in Figure 1. After a study duration of 13 weeks, all animals were euthanized through cervical dislocation. Several parameters were measured and calculated, including animal body weight, growth rate, tumor weight, and tumor characteristics. Blood was collected for biochemical analysis and subjected to centrifugal force for 15 min at 1000g to separate the plasma. Mammary tissue was removed from each rat, processed by homogenization with a suitable buffer, and then subjected to centrifugation. The resulting supernatant was utilized for biochemical analyses. Additionally, kidney and mammary tissues were preserved (10% formalin) for histological studies.

2.5. Biochemical analysis

The biochemical analysis was performed in the control and experimental rat plasma and mammary tissue. The following table 1 shows the biochemical parameters and their methods.

2.6. Histopathological studies

The control and experimental animal's kidney and mammary tissues were examined histopathologically. After fixation in 10 percent buffered formalin, the tissues were regularly treated, embedded in paraffin, and sliced into 2-3 mm sections using a rotary microtome. The resulting tissue slices were placed on glass slides and subjected to H&E (hematoxylin and eosin) staining.

2.7. Statistical analysis

The mean \pm SD is used to represent experimental values. One-way ANOVA (analysis of variance) was used to compare the mean values across groups, and DMRT was used for multiple comparisons. And the significance was set at $p \leq 0.05$.

3. Results

3.1. Effect of CVC-ZnO QDs on growth rate, body weight, and tumor characteristics

Table 2 represents the growth rate, body weight, and tumor characteristics of the control and experimental rats. The body weight was assessed based on the differences observed in the initial and final stages between the control and experimental rats. Rats treated with DMBA alone exhibited a notable reduction in growth rate and body weight. Nevertheless, administering CVC-ZnO QDs orally at various doses resulted in a substantial enhancement of growth rate and body weight in cancer bringing rats (DMBA-induced). In contrast to the control group of rats, the group administered CVC-ZnO QDs showed no notable variances in their growth rate and body weight.

Tumor burden and volume in rats were typically evaluated by measuring the size of tumors using calipers and calculated the tumor volume using the formula $V = 4/3\pi (D1/2) (D2/2) (D3/2)$. In the DMBA alone-induced rats, 100% tumor development was observed, with tumor burden (200.77 mm³) and tumor volume (33.46 mm³). Oral supplementation with CVC-ZnO QDs to cancer-induced rats markedly decreased tumor volume, burden and incidence. In control and CVC-ZnO QDs alone animals, no tumors were seen. Hence, among the 2, 4, and 8 mg/kg b.w doses, the activity of CVC-ZnO QDs at 4 mg/kg b.w had an impact in the DMBA-induced rats.

3.2. Effect of CVC-ZnO QDs on antioxidant status

The enzymatic and non-enzymatic antioxidant activities of control and experimental rats are shown in Tables 3 and 4. The presence of antioxidant in the plasma and mammary tissues of rats treated with DMBA alone was notably reduced when compared to the control group. The addition of CVC-ZnO QDs at various concentrations notably enhanced antioxidant activities in comparison to rats induced with DMBA alone. Particularly observed outcomes were evident in rats that received supplementation of CVC-ZnO QDs at a dose of 4 mg/kg b.w.

3.3. Effect of CVC-ZnO QDs on lipid peroxidation

The lipid peroxidation levels in the plasma and mammary tissue of both control and experimental rats are detailed in Table 5. The findings suggest that the lipid peroxidation levels were significantly elevated in both the plasma and mammary tissues of rats subjected to DMBA treatment alone when compared to the control group. Oral supplementation of CVC-ZnO QDs to DMBA-treated rats significantly reduced lipid peroxidation levels. In comparison with

control rats, CVC-ZnO QDs alone treated demonstrated no significant variation in lipid peroxidation within the plasma and mammary tissue.

12 3.4. Effect of CVC-ZnO QDs on detoxification enzymes

Table 6 displays the enzymatic detoxification activities observed in the mammary tissues of both control and experimental groups of rats. The rats treated with DMBA alone exhibited significantly higher phase I enzyme activity than the control rats. Supplementation with CVC-ZnO QDs reduced DMBA-metabolizing phase I enzyme activities. Compared to the control rats the rats treated with DMBA alone showed a notable decrease in the enzyme activities of phase II. Conversely, treatment through CVC-ZnO QDs significantly improved the phase II enzymes activities in the DMBA-treated rats. Notably, the medium dose of CVC-ZnO QDs at 4 mg/kg b.w demonstrated a more pronounced effect than the other two.

31 3.5. Effect of CVC-ZnO QDs on lipid profile

Table 7 represents the lipid profile of the control and experimental rats in the plasma and mammary tissues. In the DMBA- alone induced rats, concentrations of PL, FFA, TC, and TG were significantly higher in the plasma compared to the control rats. However, in mammary tissues of DMBA alone rats the PL and FFA levels were decreased and the TC and TG levels were significantly increased. CVC-ZnO QDs administration to DMBA-induced rats brought back the levels to near those of the control values in both the plasma and mammary tissues.

3.6. Histological changes

Figure 2 and 3 shows the histological changes of kidney and mammary tissue of control and experimental rats. Histopathological examination of kidney tissue of DMBA-alone induced rats revealed several features, including tubular dilation, loss of nucleus, tubular atrophy, and glomerular injury. Control and CVC-ZnO QDs alone-rats revealed the normal architecture of the kidney tissue. CVC-ZnO QDs treated DMBA-induced rats showed mild acute tubular injury, inflammatory cell infiltration, and fibrosis. Histology of mammary tissue of DMBA induced rats demonstrated infiltrating malignant tumor. CVC-ZnO QDs treated rats showed mild tumor infiltration and fibrosis with near normal architecture. The control and CVC-ZnO QDs treated alone rats showed normal architecture of the mammary tissues.

4. Discussion

Nanotechnology enhances cancer treatment by delivering targeted therapies at the molecular level, increasing precision and minimizing damage to healthy cells (Yao et al., 2020). QDs are nanocrystals with unique optical properties that can be used as carriers due to their small size and biocompatibility (Badilli et al., 2020). There have been studies on drug-loaded QDs for the chemoprevention of cancerous breast cells. One study used folate receptor-targeted DOX-QD-loaded NPs, to treat female mice with breast cancer induced by a triple-negative cancer cell (Alibolandi et al., 2016). Carvacrol is a natural compound with anticarcinogenic activity (Sivaranjani et al., 2016). The previous study found that CVC caused breast cancer cells to undergo cell death and inhibited the cell proliferation genes involved in cancer (Li et al., 2021). DMBA induces mammary carcinogenesis by acting as a potent chemical carcinogen that causes DNA damage and mutations in mammary epithelial cells. It initiates and promotes the mammary tumors by disrupting the regulatory mechanisms of growth and proliferation of cell (Kerdelhué et al., 2016). The present study investigates the pH-sensitive CVC-ZnO QDs anticancer efficacy in DMBA-induced mammary carcinoma by its ability to scavenge free radicals and detoxify chemical carcinogens.

Antioxidants are essential in protecting cells from the damage caused by ROS (reactive oxygen species), which are highly reactive molecules that can induce oxidative harm to lipids, DNA, and proteins. Lipid peroxidation is a process that occurs when ROS attacks the PUFA (polyunsaturated fatty acids) in cell membranes, leading to the formation of highly reactive lipid radicals that can further damage cell structures (Yaman et al., 2021).

Lipid peroxidation and ROS have been implicated in the progression and development of cancer, affecting the metabolisms that lead to weight loss (Davis et al., 2001). In this research, the growth rate and body weight of DMBA alone rats were reduced because of the changes in biochemical processes during tumor development. In addition, elevated lipid peroxidation levels play a crucial role in tumor formation. CVC-ZnO QDs-supplemented rats show elevated body weight and growth rate, indicating decreased lipid peroxidation levels. Further, CVC-ZnO QDs decreased the quantity of tumors and the volume of tumors in rats induced with DMBA. This may be due to the targeted drug delivery of nanoparticles (Alibolandi et al., 2016).

Evidence suggests high lipid peroxidation levels are associated with cancer progression and development. This is because the oxidative stress caused by free radicals and ROS can damage DNA and other cellular components, leading to mutations and abnormal cell growth

(Jelic et al., 2021). This study increased lipid peroxidation levels (TBARs, LOOH, and CD in DMBA-treated rats. However, CVC-ZnO QDs treated rats show decreased lipid peroxidation levels.

Antioxidants work by neutralizing free radicals, which help to protect cells from damage (Jelic et al., 2021). SOD helps to convert the O_2 (superoxide radical) into H_2O_2 (hydrogen peroxide) and O (oxygen). CAT converts H_2O_2 into H_2O and O , thereby preventing the accumulation of H_2O_2 . GPx uses glutathione as a cofactor to reduce H_2O_2 and lipid peroxides into less harmful molecules. GSH (tripeptide), Vit-C, and Vit-E donate electrons to ROS and free radicals, thereby neutralizing their damaging effects (Goodman et al., 2011). Previous research has shown that antioxidants (enzymatic and non-enzymatic) can reduce the severity and incidence of cancer burden in DMBA-induced animals (Latif et al., 2021). In this study, we observed that antioxidants decreased in the cancer-bearing rats. Vayalakkara et al. (2022) study showed that QDs conjugated with a drug antioxidant could selectively target cancer cells and induce cell death by reducing cellular antioxidant defenses. CVC-ZnO QDs treated DMBA-induced rats showed increased antioxidant levels due to reduced oxidative stress.

Detoxification enzymes play crucial roles in metabolizing and eliminating chemicals from the body. Phase I metabolism involves the oxidation of DMBA by enzymes such as Cyt-b5 and CYP450. Phase II enzymes, including GR and GST, can conjugate these reactive intermediates with glutathione and other molecules to facilitate their excretion from the body, thereby reducing cancer risk (Dhamodharan et al., 2021). According to Latif et al., (2021), rats induced with DMBA, increase the activity of phase I enzymes and decrease the activity of phase II enzymes. In this investigation, we noted an augmentation of phase I enzyme activity and a corresponding decrease in phase II enzyme activity in rats subjected to DMBA treatment. On the other hand, CVC-ZnO QDs treated DMBA-induced rats show significantly altered detoxification enzymes, possibly due to CVC delivery to breast cancer cells, potentially enhancing the bodies to eliminate carcinogens and prevent cancer development. A previous study observed that CVC could increase and decrease in phases I and II enzymes activity in DMH-induced colon carcinogenesis, respectively (Sivaranjani et al., 2016).

Lipid profile typically includes PL, FFA, TC, and TG measurements in the bloodstream. From a study by Dhamodharan et al. (2021), DMBA-induced mammary cancer in rats is associated with significant increases in plasma PL, FFA, TC, and TG levels. High PL, FFA, TC, and TG levels, on the other hand, are associated with increased cancer risk. This may be

because cholesterol is a precursor for estrogen, a hormone that can promote the growth of mammary cancer. In our study, we noticed that lipid profile (PL, FFA, TC, and TG) were increased in the plasma of DMBA-induced rats, while in mammary tissues, the PL and FFA levels were decreased. This decrease may result from increased PL degradation, impairing membrane function. Another possible explanation for the reduced PL concentration could be the decreased FFA levels in the mammary tissues, as suggested by Van Hoesen et al. in 1973. Due to the strong antihyperlipidemic properties of CVC-ZnO QDs, supplemented DMBA-treated rats showed significantly decreased plasma PL, FFA, TC, and TG lipid profiles. In contrast, elevated levels of PL and FFA were observed in the mammary tissues.

Nandakumar et al. (2011) reported that DMBA-treated rats showed renal tubular structure enlargement and loss of architecture. In this present study, DMBA-induced rats show significant kidney damage, including tubular dilation, loss of nucleus, tubular atrophy, and glomerular injury. Nanoparticles can be engineered to selectively target the kidneys, allowing for precise delivery of drugs to the affected area. This can be particularly useful in treating conditions such as kidney inflammation. In this context, Ozturk et al. (2018) suggested that CVC could reduce kidney injury induced by bilateral renal ischemia/reperfusion (I/R) in female SD rats. In our study, treatment with CVC-ZnO QDs at different doses significantly reduced these pathological changes, with the 4 mg /kg b.w. dose showing the greatest protective effect.

Histopathology of mammary tissue of DMBA-induced rats exhibited IMT. These results agree with Latif et al. (2021). On the contrary, rats treated with CVC-ZnO QDs displayed no indications of necrosis or cellular growth in the mammary gland. A more noticeable impact was noted in the rats subjected to a dosage of 4 mg/kg b.w of CVC-ZnO QDs, where the mammary tissue exhibited nearly normal architecture with an increased area of fibrosis representing residual tumor. Therefore, those data suggest that the CVC-ZnO QDs have the capability to provide a secure and efficient approach to treating cancer with significant anticancer activities.

5. Conclusion

Based on the research findings, pH-sensitive CVC-ZnO QDs have a significant anticancer effect against mammary carcinoma induced by DMBA in female SD rats. The results demonstrate that CVC-ZnO QDs significantly reduced tumor growth, improved the antioxidant status, decreased lipid peroxidation, and modulated the biotransformation enzyme activities in both plasma and mammary tissues. CVC-ZnO QDs also exhibited a positive impact on the lipid profile. Furthermore, the histological examination of the mammary and kidney tissue revealed that CVC-

ZnO prevents DMBA-induced tissue damage. These findings suggest that CVC-ZnO QDs can be a safe and effective anticancer agent for treating mammary carcinoma. However, further studies are needed to explore the molecular mechanisms underlying the anticancer properties of CVC-ZnO QDs.

Acknowledgments

Researchers Supporting Project number (RSPD2023R712), the King Saud University in Riyadh, Saudi Arabia, and the RUSA 2.0 Project grant (RUSA-100-E-002) from Annamalai University in Tamilnadu, India, provided support for this project.

Conflict of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability statement

Data will be provided upon request.

Reference

1. Alibolandi, M., Abnous, K., Sadeghi, F., Hosseinkhani, H., Ramezani, M. and Hadizadeh, F., 2016. Folate receptor-targeted multimodal polymersomes for delivery of quantum dots and doxorubicin to breast adenocarcinoma: in vitro and in vivo evaluation. *International Journal of Pharmaceutics*, 500(1-2), pp.162-178. <https://doi.org/10.1016/j.ijpharm.2016.01.040>
2. Anwar, M.M., Shalaby, M., Embaby, A.M., Saeed, H., Agwa, M.M. and Hussein, A., 2020. Prodigiosin/PU-H71 as a novel potential combined therapy for triple negative breast cancer (TNBC): preclinical insights. *Scientific Reports*, 10(1), p.14706. <https://doi.org/10.1038/s41598-020-71157-w>
3. Badilli, U., Mollarasouli, F., Bakirhan, N.K., Ozkan, Y. and Ozkan, S.A., 2020. Role of quantum dots in pharmaceutical and biomedical analysis, and its application in drug delivery. *TrAC Trends in Analytical Chemistry*, 131, p.116013. <https://doi.org/10.1016/j.trac.2020.116013>
4. Beutler, E. and Kelly, B.M., 1963. The effect of sodium nitrite on red cell GSH. *Experientia*, 19(2), pp.96-97. <https://doi.org/10.1007/bf02148042>
5. Cao, W., Chen, H.D., Yu, Y.W., Li, N. and Chen, W.Q., 2021. Changing profiles of cancer burden worldwide and in China: a secondary analysis of the global cancer statistics 2020. *Chinese medical journal*, 134(07), pp.783-791. <https://doi.org/10.1097/cm9.0000000000001474>
6. Carlberg, I. and Mannervik, B., 1985. [59] Glutathione reductase. In *Methods in enzymology* (Vol. 113, pp. 484-490). Academic press. [https://doi.org/10.1016/S0076-6879\(85\)13062-4](https://doi.org/10.1016/S0076-6879(85)13062-4)

7. Davis, L. and Kuttan, G., 2001. Effect of *Withania somnifera* on DMBA induced carcinogenesis. *Journal of ethnopharmacology*, 75(2-3), pp.165-168. [https://doi.org/10.1016/s0378-8741\(00\)00404-9](https://doi.org/10.1016/s0378-8741(00)00404-9)
8. Dhamodharan, K., Vengaimaran, M. and Sankaran, M., 2021. Chemomodulatory Effect of Capsaicin Encapsulated Chitosan Nanoparticles on Lipids, Lipoproteins and Glycoprotein Components in 7, 12-Dimethylbenz [a] anthracene (DMBA) Induced Mammary Carcinogenesis in Sprague-Dawley Rats. *Eurasian Journal of Medicine and Oncology*, 5(4), pp.350-357. <https://dx.doi.org/10.14744/ejmo.2021.61180>
9. Edis, Z., Wang, J., Waqas, M.K., Ijaz, M. and Ijaz, M., 2021. Nanocarriers-mediated drug delivery systems for anticancer agents: an overview and perspectives. *International journal of nanomedicine*, pp.1313-1330. <https://doi.org/10.2147/ijn.s289443>
10. Falholt, K., Lund, B. and Falholt, W., 1973. An easy colorimetric micromethod for routine determination of free fatty acids in plasma. *Clinica chimica acta*, 46(2), pp.105-111. [https://doi.org/10.1016/0009-8981\(73\)90016-8](https://doi.org/10.1016/0009-8981(73)90016-8)
11. Foster, L.B. and Dunn, R.T., 1973. Stable reagents for determination of serum triglycerides by a colorimetric Hantzsch condensation method. *Clinical chemistry*, 19(3), pp.338-340.
12. Goodman, M., Bostick, R.M., Kucuk, O. and Jones, D.P., 2011. Clinical trials of antioxidants as cancer prevention agents: past, present, and future. *Free Radical Biology and Medicine*, 51(5), pp.1068-1084. <https://doi.org/10.1016/j.freeradbiomed.2011.05.018>
13. Habig, W.H., Pabst, M.J. and Jakoby, W.B., 1974. Glutathione S-transferases: the first enzymatic step in mercapturic acid formation. *Journal of biological Chemistry*, 249(22), pp.7130-7139. [https://doi.org/10.1016/S0021-9258\(19\)42083-8](https://doi.org/10.1016/S0021-9258(19)42083-8)
14. He, J., Li, C., Ding, L., Huang, Y., Yin, X., Zhang, J., Zhang, J., Yao, C., Liang, M., Pirraco, R.P. and Chen, J., 2019. Tumor targeting strategies of smart fluorescent nanoparticles and their applications in cancer diagnosis and treatment. *Advanced Materials*, 31(40), p.1902409. <https://doi.org/10.1002/adma.201902409>
15. Jelic, M.D., Mandic, A.D., Maricic, S.M. and Srdjenovic, B.U., 2021. Oxidative stress and its role in cancer. *Journal of cancer research and therapeutics*, 17(1), pp.22-28. https://doi.org/10.4103/jcrt.jcrt_862_16
16. Jiang, J., Pi, J. and Cai, J., 2018. The advancing of zinc oxide nanoparticles for biomedical applications. *Bioinorganic chemistry and applications*, 2018. <https://doi.org/10.1155/2018/1062562>
17. Jiang, Z.Y., Hunt, J.V. and Wolff, S.P., 1992. Ferrous ion oxidation in the presence of xylenol orange for detection of lipid hydroperoxide in low density lipoprotein. *Analytical biochemistry*, 202(2), pp.384-389. [https://doi.org/10.1016/0003-2697\(92\)90122-n](https://doi.org/10.1016/0003-2697(92)90122-n)
18. Kakkar, P., Das, B. and Viswanathan, P.N., 1984. A modified spectrophotometric assay of superoxide dismutase.
19. Kerdelhué, B., Forest, C. and Coumoul, X., 2016. Dimethyl-Benz (a) anthracene: A mammary carcinogen and a neuroendocrine disruptor. *Biochimie open*, 3, pp.49-55. <https://doi.org/10.1016/j.biopen.2016.09.003>

20. Latif, A., Issa Khan, M., Rakha, A. and Ali Khan, J., 2021. Evaluating the therapeutic potential of white button mushroom (*Agaricus bisporus*) against DMBA-induced breast cancer in Sprague Dawley rats. *Journal of Food Biochemistry*, 45(12), p.e13979. <https://doi.org/10.1111/jfbc.13979>
21. Li, L., He, L., Wu, Y. and Zhang, Y., 2021. Carvacrol affects breast cancer cells through TRPM7 mediated cell cycle regulation. *Life sciences*, 266, p.118894. <https://doi.org/10.1016/j.lfs.2020.118894>
22. Nandakumar, N. and Balasubramanian, M.P., 2011. Hesperidin protects renal and hepatic tissues against free radical-mediated oxidative stress during DMBA-induced experimental breast cancer. *Journal of Environmental Pathology, Toxicology and Oncology*, 30(4). <https://doi.org/10.1615/jenvironpatholtoxicoloncol.v30.i4.20>
23. Ohkawa, H., Ohishi, N. and Yagi, K., 1979. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Analytical biochemistry*, 95(2), pp.351-358. [https://doi.org/10.1016/0003-2697\(79\)90738-3](https://doi.org/10.1016/0003-2697(79)90738-3)
24. Omaye, S.T., Turnbull, J.D. and Sauberlich, H.E., 1979. [1] Selected methods for the determination of ascorbic acid in animal cells, tissues, and fluids. In *Methods in enzymology* (Vol. 62, pp. 3-11). Academic press. [https://doi.org/10.1016/0076-6879\(79\)62181-x](https://doi.org/10.1016/0076-6879(79)62181-x)
25. Omura, T. and Sato, R., 1964. The carbon monoxide-binding pigment of liver microsomes. I. Evidence for its hemoprotein nature. *J Biol Chem*, 239(7), pp.2370-2378.
26. Ozturk, H., Cetinkaya, A., Duzcu, S.E., Tekce, B.K. and Ozturk, H., 2018. Carvacrol attenuates histopathologic and functional impairments induced by bilateral renal ischemia/reperfusion in rats. *Biomedicine & Pharmacotherapy*, 98, pp.656-661. <https://doi.org/10.1016/j.biopha.2017.12.060>
27. Palan, P.R., Mikhail, M.S., Basu, J. and Romney, S.L., 1991. Plasma levels of antioxidant β -carotene and α -tocopherol in uterine cervix dysplasias and cancer. <https://doi.org/10.1080/01635589109514106>
28. Rajitha, B., Malla, R.R., Vadde, R., Kasa, P., Prasad, G.L.V., Farran, B., Kumari, S., Pavitra, E., Kamal, M.A., Raju, G.S.R. and Peela, S., 2021, February. Horizons of nanotechnology applications in female specific cancers. In *Seminars in cancer biology* (Vol. 69, pp. 376-390). Academic Press. <https://doi.org/10.1016/j.semcancer.2019.07.005>
29. Rao, K.S. and Recknagel, R.O., 1968. Early onset of lipoperoxidation in rat liver after carbon tetrachloride administration. *Experimental and molecular pathology*, 9(2), pp.271-278. [https://doi.org/10.1016/0014-4800\(68\)90041-5](https://doi.org/10.1016/0014-4800(68)90041-5)
30. Rotruck, J.T., Pope, A.L., Ganther, H.E., Swanson, A.B., Hafeman, D.G. and Hoekstra, W.G., 2011. Selenium: biochemical role as a component of glutathione peroxidase. *Science* 1973; 179: 588-90. *Arh Hig Rada Toksikol*, 62, pp.121-129. <https://doi.org/10.1126/science.179.4073.588>
31. Sinha, A.K., 1972. Colorimetric assay of catalase. *Analytical biochemistry*, 47(2), pp.389-394. [https://doi.org/10.1016/0003-2697\(72\)90132-7](https://doi.org/10.1016/0003-2697(72)90132-7)
32. Sivaranjani, A., Sivagami, G. and Nalini, N., 2016. Chemopreventive effect of carvacrol on 1, 2-dimethylhydrazine induced experimental colon carcinogenesis.

- Journal of Cancer Research and Therapeutics, 12(2), pp.755-762.
<https://doi.org/10.4103/0973-1482.154925>
33. Tagde, P., Kulkarni, G.T., Mishra, D.K. and Kesharwani, P., 2020. Recent advances in folic acid engineered nanocarriers for treatment of breast cancer. *Journal of Drug Delivery Science and Technology*, 56, p.101613.
<https://doi.org/10.1016/j.jddst.2020.101613>
 34. Van Hoeven, R.P. and Emmelot, P., 1973. Plasma membrane lipids of normal and neoplastic tissues. *Tumor Lipids* R. Wood, editor. American Oil Chemists Soc., Champaign, 111, pp.126-138.
 35. Vayalakkara, R.K., Lo, C.L., Chen, H.H., Shen, M.Y., Yang, Y.C., Sabu, A., Huang, Y.F. and Chiu, H.C., 2022. Photothermal/NO combination therapy from plasmonic hybrid nanotherapeutics against breast cancer. *Journal of Controlled Release*, 345, pp.417-432.
<https://doi.org/10.1016/j.jconrel.2022.03.030>
 36. Yagi, K., 1987. Lipid peroxides and human diseases. *Chemistry and physics of lipids*, 45(2-4), pp.337-351. [https://doi.org/10.1016/0009-3084\(87\)90071-5](https://doi.org/10.1016/0009-3084(87)90071-5)
 37. Yaman, S.O. and Ayhanci, A., 2021. Lipid peroxidation. *Accenting Lipid Peroxidation*, pp.1-11. <http://dx.doi.org/10.5772/intechopen.95802>
 38. Yao, Y., Zhou, Y., Liu, L., Xu, Y., Chen, Q., Wang, Y., Wu, S., Deng, Y., Zhang, J. and Shao, A., 2020. Nanoparticle-based drug delivery in cancer therapy and its role in overcoming drug resistance. *Frontiers in molecular biosciences*, 7, p.193.
<https://doi.org/10.3389%2Ffmolb.2020.00193>
 39. Zilversmit, D.B. and Davis, A.K., 1950. Microdetermination of plasma phospholipids by trichloroacetic acid precipitation. *The Journal of Laboratory and Clinical Medicine*, 35(1), pp.155-160.
 40. Zlatkis, A., Zak, B. and Boyle, A.J., 1953. A new method for the direct determination of serum cholesterol. *The Journal of laboratory and clinical medicine*, 41(3), pp.486-492.

supplementary

ORIGINALITY REPORT

18%

SIMILARITY INDEX

15%

INTERNET SOURCES

16%

PUBLICATIONS

4%

STUDENT PAPERS

PRIMARY SOURCES

- 1 Michael Faupel, Felix von Blanckenhagen, Johannes Lückmann, Daniel Ruf, Gisela Wiedemann, Jan-Dieter Ludwigs. "Precision farming and environmental pesticide regulation in the EU—How does it fit together?", *Integrated Environmental Assessment and Management*, 2022
Publication 1%
- 2 Uma Saminathan, Pachiappan Pugalendhi, Suganthi Subramaniyan, Rajendran Jayaganesh. "Biochemical studies evaluating the chemopreventive potential of brucine in chemically induced mammary carcinogenesis of rats", *Toxicology Mechanisms and Methods*, 2019
Publication 1%
- 3 www.tandfonline.com
Internet Source 1%
- 4 Otmane Zouirech, Abdullah A. Alyousef, Azeddin El Barnossi, Abdelfattah El Moussaoui et al. "Phytochemical Analysis and

Antioxidant, Antibacterial, and Antifungal Effects of Essential Oil of Black Caraway (*Nigella sativa* L.) Seeds against Drug-Resistant Clinically Pathogenic Microorganisms", BioMed Research International, 2022

Publication

5

[topics.sciencedirect.com](https://www.topics.sciencedirect.com)

Internet Source

1 %

6

A. Lakshmi, S. Subramanian.

"Chemotherapeutic effect of tangeretin, a polymethoxylated flavone studied in 7, 12-dimethylbenz(a)anthracene induced mammary carcinoma in experimental rats", Biochimie, 2014

Publication

1 %

7

P. Pugalendhi, S. Manoharan.

"Chemopreventive Potential of Genistein and Daidzein in Combination during 7,12-dimethylbenz(a)anthracene (DMBA) Induced Mammary Carcinogenesis in Sprague-Dawley Rats", Pakistan Journal of Biological Sciences, 2010

Publication

1 %

8

pericles.pericles-prod.literatumonline.com

Internet Source

1 %

9

www.journaljpri.com

Internet Source

1 %

10	publications.waset.org Internet Source	1 %
11	Manoharan, S.. "Carnosic acid: A potent chemopreventive agent against oral carcinogenesis", <i>Chemico-Biological Interactions</i> , 20101205 Publication	<1 %
12	informaticsjournals.com Internet Source	<1 %
13	Shuang Hao, Rihao Xu, Dan Li, Zhicheng Zhu, Tiance Wang, Kexiang Liu. "Attenuation of Streptozotocin-Induced Lipid Profile Anomalies in the Heart, Brain, and mRNA Expression of HMG-CoA Reductase by Diosgenin in Rats", <i>Cell Biochemistry and Biophysics</i> , 2015 Publication	<1 %
14	coek.info Internet Source	<1 %
15	H. V. Sudeep, K. Venkatakrishna, Amritharaj, K. Gouthamchandra, B. Reethi, P. Naveen, H. B. Lingaraju, K. Shyamprasad. "A standardized black pepper seed extract containing β -caryophyllene improves cognitive function in scopolamine-induced amnesia model mice via regulation of brain-derived neurotrophic	<1 %

factor and MAPK proteins", Journal of Food Biochemistry, 2021

Publication

16

bmccomplementmedtherapies.biomedcentral.com <1 %

Internet Source

17

phcogres.com <1 %

Internet Source

18

A. Justin Thenmozhi, P. Subramanian. " Antioxidant Potential of in Ammonium Chloride-Induced Hyperammonemic Rats ", Evidence-Based Complementary and Alternative Medicine, 2011

Publication

19

Jayaganesh Rajendran, Pugalendhi Pachaiappan, Suganthi Subramaniyan. "Dose-dependent chemopreventive effects of citronellol in DMBA-induced breast cancer among rats", Drug Development Research, 2019

Publication

20

Nagarethinam Baskaran, Govindan Sadasivam Selvam, Subramani Yuvaraj, Albert Abhishek. "Parthenolide attenuates 7,12-dimethylbenz[a]anthracene induced hamster buccal pouch carcinogenesis", Molecular and Cellular Biochemistry, 2017

Publication

21 Pari, L.. "Cytoprotective and antioxidant role of diallyl tetrasulfide on cadmium induced renal injury: An in vivo and in vitro study", Life Sciences, 20070123
Publication <1 %

22 ijpsr.com
Internet Source <1 %

23 Hasan Akgöz, Hüseyin Fındık, Mehmet Gökhan Aslan. "Evaluation of tear parameters and meibomian gland morphology in keratoconus patients after epithelial-on corneal cross-linking", European Journal of Ophthalmology, 2022
Publication <1 %

24 Subramanian Venkateswaran. "Antioxidant effect of Phaseolus vulgaris in streptozotocin-induced diabetic rats", Asia Pacific Journal of Clinical Nutrition, 9/2002
Publication <1 %

25 pesquisa.bvsalud.org
Internet Source <1 %

26 www.jemds.com
Internet Source <1 %

27 Bhuvaneshwari Venkataesan Kumari, Renuka Mani, Balakrishnan Ramajayam Asokan, Karthikeyan Balakrishnan et al. "Green Synthesised Silver Nanoparticles Using

Anoectochilus elatus Leaf Extract:
Characterisation and Evaluation of
Antioxidant, Anti-Inflammatory, Antidiabetic,
and Antimicrobial Activities", Journal of
Composites Science, 2023

Publication

28

Perumal Subramanian. "Prevention by
melatonin of hepatocarcinogenesis in rats
injected with N-nitrosodiethylamine", Journal
of Pineal Research, 7/19/2007

Publication

29

www.tnsroindia.org.in

Internet Source

30

Kazim Sahin, Resat Ozercan, Muhittin
Onderci, Nurhan Sahin, Frederick Khachik,
Soley Seren, Omer Kucuk. "Dietary Tomato
Powder Supplementation in the Prevention of
Leiomyoma of the Oviduct in the Japanese
Quail", Nutrition and Cancer, 2007

Publication

31

Subramanian Kumar, Pichavaram
Prahalthan, Murugesan Saravanakumar,
Boobalan Raja. "Vanillic acid prevents the
deregulation of lipid metabolism, endothelin 1
and up regulation of endothelial nitric oxide
synthase in nitric oxide deficient hypertensive
rats", European Journal of Pharmacology,
2014

<1 %

<1 %

<1 %

<1 %

32

acikerisim.ibu.edu.tr

Internet Source

<1 %

33

jomped.org

Internet Source

<1 %

34

shodhgangotri.inflibnet.ac.in:8080

Internet Source

<1 %

35

www.deswater.com

Internet Source

<1 %

36

www.ijppr.humanjournals.com

Internet Source

<1 %

37

www.phcogres.com

Internet Source

<1 %

38

www.science.gov

Internet Source

<1 %

39

Narasimhanaidu Kamalakkannan.
"Antihyperglycaemic and Antioxidant Effect of
Rutin, a Polyphenolic Flavonoid, in
Streptozotocin-Induced Diabetic Wistar Rats",
Basic & Clinical Pharmacology & Toxicology,
1/2006

Publication

<1 %

40

R. Senthil Murugan, K.V.P. Chandra Mohan, S.
Nagini. "Modulatory Effects of Black Tea
Polyphenols on Rat Forestomach

<1 %

Carcinogenesis", Toxicology Mechanisms and Methods, 2008

Publication

41

Sivaranjani Arivalagan, Nisha Susan Thomas, Balaji Chandrasekaran, Vijay Mani et al. "Combined therapeutic efficacy of carvacrol and X-radiation against 1,2-dimethyl hydrazine-induced experimental rat colon carcinogenesis", Molecular and Cellular Biochemistry, 2015

Publication

<1 %

42

V. Manju, P. Viswanathan, N. Nalini. "Hypolipidemic Effect of Ginger in 1,2-Dimethyl Hydrazine-Induced Experimental Colon Carcinogenesis", Toxicology Mechanisms and Methods, 2008

Publication

<1 %

43

Xueyan Zhou, Ziqiang Zheng, Chang Xu, Juan Wang et al. "Disturbance of Mammary UDP-Glucuronosyltransferase Represses Estrogen Metabolism and Exacerbates Experimental Breast Cancer", Journal of Pharmaceutical Sciences, 2017

Publication

<1 %

44

downloads.hindawi.com

Internet Source

<1 %

45

jeccr.biomedcentral.com

Internet Source

<1 %

46

oaji.net
Internet Source

<1 %

47

pubs.acs.org
Internet Source

<1 %

48

www.ijper.org
Internet Source

<1 %

49

Lakshmi Arivazhagan, Sorimuthu Pillai Subramanian. "Tangeretin, a citrus flavonoid attenuates oxidative stress and protects hepatocellular architecture in rats with 7, 12 - dimethylbenz(a)anthracene induced experimental mammary carcinoma", *Journal of Functional Foods*, 2015

Publication

<1 %

50

Lakshmi, Arivazhagan, and Sorimuthu Pillai Subramanian. "Tangeretin ameliorates oxidative stress in the renal tissues of rats with experimental breast cancer induced by 7,12-dimethylbenz[a]anthracene", *Toxicology Letters*, 2014.

Publication

<1 %

51

Nagarethinam Baskaran, Shanmugam Manoharan, Subramanian Balakrishnan, Pachaiappan Pugalendhi. "Chemopreventive potential of ferulic acid in 7,12-dimethylbenz[a]anthracene-induced mammary carcinogenesis in Sprague–Dawley

<1 %

rats", European Journal of Pharmacology,
2010

Publication

52

link.springer.com
Internet Source

<1 %

Exclude quotes On

Exclude matches Off

Exclude bibliography On