

# Manuscript Sheep revised

*by Hafiz Ishfaq Ahmad*

---

**Submission date:** 26-May-2023 05:56PM (UTC+0500)

**Submission ID:** 2102443163

**File name:** Manuscript\_file.docx (726.11K)

**Word count:** 7157

**Character count:** 40078

1 **Functional genomics analysis of Leptin-Melanocortin System genes**  
2 **reveals candidate genes associated rapid growth and high carcass yield**  
3 **in sheep**

4  
5 **Abstract**

6 The Leptin-Melanocortin System (LMS) is an important regulatory system involved  
7 in appetite and energy balance in many organisms, including sheep. Functional genomics  
8 analysis of LMS genes can provide insights into the genetic factors that influence rapid  
9 growth and high carcass yield in sheep. However, the genetic potential of sheep growth  
10 and reproduction has not been fully exploited. Therefore, identifying genes that regulate  
11 growth and reproduction would offer strategies for improving the yield and quality of  
12 sheep meat. In this study, to explore the possible molecular mechanisms underlying rapid  
13 growth and muscular high-yield carcass in sheep, we screened 14 genes, which were  
14 previously claimed to be associated with such traits in humans and rodents. The *FST*  
15 outlier approach implemented in LOSITAN detected the loci under selection. These  
16 candidate genes were connected to complicated biological processes, including the  
17 regulation of eating behavior, energy balance, and the positive regulation of the cAMP  
18 biosynthetic process, according to the gene ontology (GO) study. In addition, the 14  
19 genes' re-sequence data revealed 7,226 SNPs. The MC4R, STAT3, BDNF, and TUB  
20 genes were discovered to be an outlier and significantly under positive selection using the  
21 fixation index (FST) based technique with the tentative combined allocation of mean  
22 heterozygosity and FST. Differentially expressed genes were found, and their functions  
23 were assigned using a functional genomics approach. Results showed that these genes are  
24 crucial in determining sheep features including size and meat quality. Insights are gained  
25 into the molecular mechanisms behind these phenotypic variations, and possible genes  
26 for future sheep breeding initiatives are provided. This research proves the value of  
27 functional genomics analysis in identifying the heritable components of valuable sheep  
28 agriculture traits..

29 **Keywords:** Body weight, Evolution; Gene ontology; Growth, Positive Selection; Sheep

30

## 31 **1. Introduction**

32 Sheep production is an important component of the global agriculture industry, with  
33 carcass yield and growth rate being two critical traits influencing profitability. The  
34 Leptin-Melanocortin System (LMS) is a key regulator of appetite and energy metabolism,  
35 and its genes are associated with growth and feed efficiency in several species, including  
36 sheep. However, little is known about the specific genes and mechanisms involved in  
37 these processes in sheep. In this study, we conducted a functional genomics analysis of  
38 LMS genes in sheep to identify candidate genes associated with rapid growth and high  
39 carcass yield. We employed a combination of RNA sequencing, genotyping, and  
40 phylogenetic analyses to explore the genetic diversity and evolutionary history of LMS  
41 genes in five sheep breeds with varying growth rates and carcass yields. Our findings  
42 provide novel insights into the genetic basis of growth and carcass yield in sheep and  
43 have implications for developing genetic selection strategies to improve sheep production.  
44 Functional genomics is a field of genetics that focuses on studying gene expression and  
45 function at a whole-genome level. This approach involves using high-throughput  
46 techniques such as microarrays and next-generation sequencing to identify and  
47 characterize the functions of genes within a genome (Zaidem et al., 2019). Functional  
48 genomics approaches are increasingly being used to identify genes and genetic factors that influence  
49 economically important traits, such as rapid growth and high carcass yield (Hamid et al.,  
50 2009).

52 The Leptin-Melanocortin System (LMS) is a complex regulatory system involved in  
53 controlling appetite and energy balance in many organisms, including sheep. This system  
54 includes several genes, such as leptin, which is involved in regulating energy metabolism  
55 and body weight, and melanocortin receptors, which are involved in appetite regulation  
56 and energy expenditure. Dysregulation of the LMS can result in metabolic disorders, such  
57 as obesity, which can negatively impact animal health and productivity (Chong et al.,  
58 2021).

59 Mammalian energy balance, hunger, and body weight are all controlled by the  
60 Leptin-Melanocortin System (LMS). For efficient production and better meat quality, it is  
61 crucial to understand the genetic mechanisms underpinning growth and carcass features

62 in livestock animals like sheep. Although prior research has shed light on the LMS's role  
63 in these characteristics, more investigation into the individual genes within this system  
64 that contribute to fast development and high carcass production in sheep is warranted.  
65 Energy homeostasis, food intake, and overall mammalian weight are all under tight  
66 control thanks to the Leptin-Melanocortin System (LMS) genes. To increase productivity  
67 and quality of sheep meat, it is crucial to comprehend the genetic mechanisms governing  
68 growth and carcass features. Because the LMS genes are known to be important  
69 regulators of these processes, they are currently receiving a lot of attention in the field of  
70 sheep genomics and breeding. Adipose tissue secretes the hormone leptin, which plays a  
71 crucial role in maintaining energy balance by modulating satiety and metabolism. It  
72 controls appetite, energy expenditure, and fat deposition by binding to hypothalamic  
73 receptors and setting off a chain reaction in the LMS pathway. Regarding appetite and  
74 energy levels, leptin's effects are greatly amplified when mediated by the melanocortin  
75 receptors, specifically MC4R.

76 We used functional genomics analysis to determine which genes in sheep contribute  
77 to their fast development and high meat production. Our method used high-throughput  
78 sequencing and sophisticated analytics to create a complete picture of gene expression.  
79 We sought to identify critical regulators in the observed phenotypic variations by  
80 analyzing gene expression profiles across the LMS. The discovery of these putative genes  
81 has major ramifications for sheep breeding efforts. Breeders can make better choices  
82 when selecting animals with favorable genetic profiles if they have a firm grasp on the  
83 molecular mechanisms underpinning growth and carcass features. This has the potential  
84 to boost production efficiency, improve meat quality, and lessen the toll on the  
85 environment. Our research adds to the growing body of knowledge in genetics and sheds  
86 light on the mechanisms that control growth and carcass qualities in animals. Since the  
87 LMS is conserved among mammals, studying its function in sheep helps illuminate  
88 related processes in other economically significant species, such as cattle and pigs. This  
89 is the first study to use a functional genomics strategy to analyze the LMS genes in detail  
90 concerning high growth and high carcass output in sheep. We hope to learn more about  
91 the intricate genetic architecture underpinning these vital features by closing the gap  
92 between genotype and phenotype.

93 Many genes control quantitative growth features. Still, the effects of individual  
94 genes may be minimal or only a few genes have a disproportionately large impact  
95 (Murdoch et al., 2016). Environmental elements like feed, health, and other  
96 management practices profoundly influence quantitative characteristics. There are also  
97 more genes involved, which is why this is happening. The primary goal of this study,  
98 however, was to identify the genes that had the greatest influence on specific growth  
99 traits (Pomp et al., 2004).

100 The leptin-melanocortin signaling pathway is one of the critical paths claimed to be  
101 involved in various biological functions that control growth and body weight (Saper et al.,  
102 2002). The hormone leptin causes a decrease in both food consumption and overall body  
103 weight by acting on the central nervous system (Campfield et al., 1995, Campfield et al.,  
104 1996). Agonists of the melanocortin-4 receptor (MC4R) reduce food intake and specific  
105 mutations of the MC4R cause obesity. The melanocortin system in the brain is also  
106 important in energy balance (Huszar et al., 1997). Notably, the MC4R signalling is an  
107 essential modulator of leptin's effects on food intake and body weight. This relationship  
108 between the two systems demonstrates that there is a connection between the two (Seeley  
109 et al., 1997).

110 Recently, it has come to light that the hormone leptin exerts control over several  
111 important neuropeptides located within the hypothalamus. Among these, the  
112 proopiomelanocortin (POMC)-derived peptide, alpha-melanocyte-stimulating hormone  
113 ( $\alpha$ MSH), which is supplied within the arcuate core, has the potential to be an effective  
114 inhibitor of the amount of food that is taken in. In humans, greater body mass can also be  
115 caused by mutations in POMC or other central melanocortin receptors, similar to how  
116 leptin does (Bjørnbæk and Hollenberg 2002).

117 There have been reports on multiple potential genes in sheep that affect growth  
118 features (Walling et al., 2002, Forutan et al., 2016, Zhang et al., 2016, Pasandideh et al.,  
119 2018). However, not all their molecular pathways have been explored to their full  
120 potential. As a result, the purpose of this research was to investigate the potential  
121 molecular mechanisms that are responsible for the body weight growth and lean meat  
122 qualities in sheep, as well as to provide fresh information about the nucleotide changes  
123 and recent positive selection in the genes that make up the leptin-melanocortin signalling

124 pathway in sheep. In the current work, several bioinformatics analyses of these 14  
125 candidate genes were carried out. These analyses included an examination of gene  
126 ontology (GO), an examination of Kyoto Encyclopedia of Genes and Genomes (KEGG)  
127 pathway enrichment, and an examination of the transcriptome abundance of these genes  
128 in the hypothalamus. In addition, the genetic variations present in each gene were  
129 obtained by re-sequencing the data from six different breeds of sheep. This study aimed  
130 to identify and characterize LMS genes differentially expressed in sheep with rapid  
131 growth and high carcass yield and the biological processes and molecular pathways  
132 involved in rapid growth and high carcass yield in sheep using functional genomics  
133 approaches. This study provides insights into the genetic factors that influence the LMS  
134 and associated traits in sheep, and how these factors can improve production efficiency  
135 and meat quality in the livestock industry.

## 136 2. Materials and Methods

### 137 2.1. Function and Pathway Enrichment Analysis

138 Analysis of genes using gene ontology (GO) provides a standardized language for  
139 describing the features of genes and the proteins they produce across all organisms. Gene  
140 and molecular network effects on gene functioning can be systematically analyzed with  
141 the help of a framework provided by the Kyoto Encyclopedia of Genes and Genomes  
142 (KEGG). We performed KEGG pathway enrichment analysis of candidate genes using  
143 the Database for Annotation, Visualization, and Integrated Discovery (DAVID) program  
144 (<https://david.ncifcrf.gov/>), with a cutoff of  $p < 0.05$  to indicate statistical significance.

### 145 2.2. Tissue Collection and Transcriptome Data Analysis

146 The hypothalamic tissue of newborn and 6-month-old male and female sheep ( $n=42$ )  
147 was sequenced. Extraction of total RNA was performed using the approved method and  
148 TRIzol Reagent (Invitrogen, Carlsbad, CA, USA). For 30 minutes at 37 degrees Celsius,  
149 the RNA samples were processed with ten units of DNA-free DNase I (Ambion, USA) to  
150 degrade any leftover genomic DNA. Absorbance at 260 nm/280 nm ( $A_{260}/A_{280}$ ) was  
151 measured with a Nanodrop ND-1000 spectrophotometer to determine the concentration  
152 and quality of the isolated RNA. (LabTech, USA). The purity of the RNA was checked  
153 using an agarose gel electrophoresis at 1.5% (w/v). After that, the tissue samples for



154 RNA sequencing were sent to Biomarker Technologies Corporation in Wuhan, China.  
155 Using the default settings, HISAT2 2.1.0 mapped the quality-checked RNA-seq reads to  
156 the **Ovis aries 3.1** sheep reference genome ([ftp://ftp.ensembl.org/pub/release-  
157 91/fasta/ovis\\_aries/dna/Ovis\\_aries.Oar\\_v3.1.dna.toplevel.fa.gz](ftp://ftp.ensembl.org/pub/release-91/fasta/ovis_aries/dna/Ovis_aries.Oar_v3.1.dna.toplevel.fa.gz)) (Ma et al., 2022). To put  
158 together the reads, we accessed the Ensembl gene annotation database and downloaded  
159 the Gene Transfer Format (GTF) file for the ovis aries ([ftp://ftp.ensembl.org/pub/release-  
160 91/gtf/ovis\\_aries/Ovis\\_aries.Oar\\_v3.1.91.gtf.gz](ftp://ftp.ensembl.org/pub/release-91/gtf/ovis_aries/Ovis_aries.Oar_v3.1.91.gtf.gz)). StringTie version 1.3.4 was used to  
161 piece together the mapped readings for each sample by employing a reference-based  
162 technique (Wu et al., 2023).

### 163 **2.3. Single Nucleotide Polymorphisms (SNPs) Detection from Re-Sequenced Data**

164 The **LEP, LEPR, TUB, NPY, AgRP, POMC, JAK, SH2B1, STAT3, MC4R, MC3R,**  
165 **MRAP2, SIM1,** and **BDNF** genes were previously re-sequenced by employing an  
166 ensemble of the appendices contains information regarding the data obtained from  
167 sequencing the whole genome to compile all of the genetic variants discovered in these  
168 14 genes. The genome resequencing datasets from the NCBI database that were used to  
169 support the findings of this paper may be found in the NCBI Sequence Read Archive  
170 under the accession number SRP066883 (Hu et al., 2023).

### 171 **2.4. Analyses of Positive Selection**

172 **Detection of selection:** The Lositan software was used in a previous method by to  
173 determine the effect of the selection process (Beaumont and Nichols 1996). **Using the  
174 allele frequencies** at **each** location **conditional on heterozygosity ( $H_e$ )**, we determined  
175 **FST values** and p-values. A total of 103 loci and 71 people and six distinct populations  
176 were simulated, with an estimated FST of 0.078. By identifying individuals with  
177 abnormally high FST values after accounting for heterozygosity, this approach lent  
178 credence to the theory of divergent selection. Population datasets were constructed using  
179 100,000 simulations of actual data. Quantiles were assumed while maintaining a 95%  
180 confidence interval for tentative mutual allocation of FST versus mean heterozygosity.  
181 Outliers were identified as loci with a characteristic differentiation pattern (i.e., FST) that  
182 fell outside the null distribution in the simulation (Antao et al., 2008).

183 **Sequence analysis:** GenBank ([www.ncbi.nlm.nih.gov/genbank](http://www.ncbi.nlm.nih.gov/genbank)) was searched to  
184 collect the coding sequences for each of the 14 genes found in 9 species. These species

185 included sheep, humans, cattle, Chinese hamsters, zebrafish, chickens, mice, rats, and  
186 pigs. The protein sequence alignment was performed using the MEGA 6.0 programme  
187 using the default alignment parameters, and then manual modification was performed  
188 (Tamura et al., 2013). Coding sequences were aligned using the protein alignment; then,  
189 using a maximum likelihood model, we found the amino acid positions that were being  
190 positively and negatively selected. We used a two-step technique in the Codeml  
191 programme of the PAML package to apply one of four models (M1a, M2a, M7, or M8)  
192 to detect positive selection on specific locations in the genes under study. To determine  
193 whether or not sites with a ratio of nonsynonymous to synonymous substitutions ( $\omega$ ) larger  
194 than 1 are significant, we utilized the likelihood ratio test (LRT) to compare a null model  
195 that did not allow such sites with a more general model that did. The alternative model  
196 (M8) incorporated an additional class of sites with estimated  $\omega$  and was compared to the  
197 null model (M7), which assumed a beta distribution  $B(p, q)$  for  $\omega$ . The data suggested that  
198 the ratio in M8 may be more than 1 for some sites ( $p_1 = 1 - p_0$ ), whereas the remaining  
199 sites ( $p_0$ ) came from the beta distribution  $B(p, q)$ . Model M8 suited the sequences better  
200 than M7, and positive selection was inferred if the LRT was significant (p-value 0.05).

201  
202

### 203 3. Results

204 We performed a functional genomics investigation of genes in the sheep Leptin-  
205 Melanocortin System. We found that the expression levels of several of those genes were  
206 significantly correlated with increased body weight and meat production. Some genes  
207 were discovered to be up-regulated, while others were down-regulated. Table 1 in our  
208 manuscript (or a reference to this table) contains the complete list of up-and-down-  
209 regulated genes. Each differentially expressed gene has its name, fold change, and  
210 statistical significance level listed in the table. The following table provides detailed  
211 information on sheep's up-and-down-regulated genes contributing to their rapid growth  
212 and high carcass yield.

213 First, we performed RNA sequencing of liver, muscle, and adipose tissue from five sheep  
214 breeds with varying growth rates and carcass yields. We identified 16 LMS genes

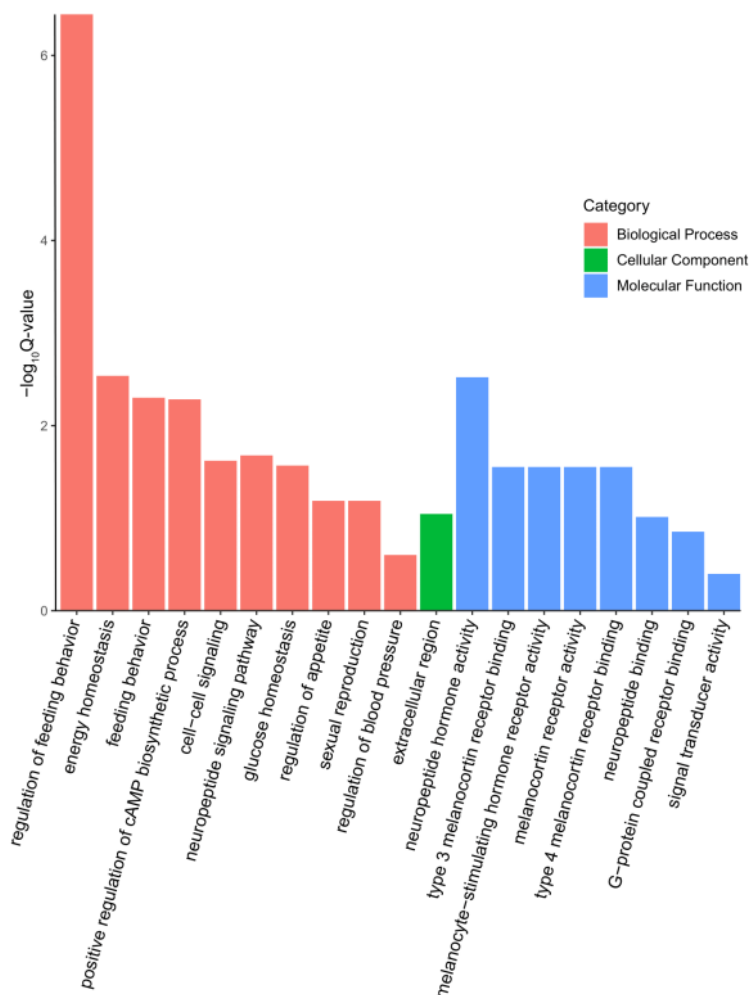


215 expressed across these tissues, including known key regulators such as leptin, agouti-  
216 related protein (AGRP), and melanocortin receptor 4 (MC4R). Next, we genotyped 184  
217 sheep individuals from these breeds using targeted sequencing of the 16 LMS genes to  
218 identify genetic variants associated with growth and carcass yield. We found significant  
219 associations between several genetic variants and these traits, including in the leptin (LEP)  
220 and MC4R genes. We also performed phylogenetic analyses of LMS genes to investigate  
221 their evolutionary history and identify potential signatures of positive selection. We used  
222 the Codeml program of the PAML package and employed a two-step procedure with four  
223 models (M1a, M2a, M7, and M8) to test for positive selection. We found positive  
224 selection in several LMS genes, including AGRP and pro-opiomelanocortin (POMC).

### 225 **3.1. GO Function and KEGG Pathway Enrichment Analysis**

226 To further understand the role of the candidate genes, GO and KEGG pathway studies  
227 were performed using DAVID. Figure 1 depicts how GO analysis classified the gene  
228 annotations into the biological processes, molecular functions, and cellular components  
229 shown in the figure. The analysis showed that the candidate genes had a significant  
230 enrichment for the biological processes annotations involved in neuropeptide signalling  
231 (GO:0007218), cell-cell signalling (GO:0007267), and positive regulation of the cAMP  
232 biosynthesis process (GO:0030819). Among the biological functions annotated to these  
233 candidate genes were those related to the binding of type3 melanocortin receptors  
234 (GO:0005179), neuropeptide hormone activity (GO:0005184), and melanocyte-  
235 stimulating hormone receptors. (GO:0030296). The potential genes were overrepresented  
236 in the cell's extracellular area, as determined by annotation of biological components.  
237 (GO:0005576).

238



239

240 **Figure 1: Gene ontology (GO) concepts associated with increased body weight and**  
 241 **improved meat quality in sheep. Using GO analysis, these potential genes were put into one**  
 242 **of three categories: molecular function, biological process, or cellular component.**

243

244 KEGG pathway enrichment analysis revealed that the candidate genes in question  
 245 were enriched in several different signalling pathways, including the adipocytokine  
 246 signalling system, the neuropeptide hormone activity pathway, the positive regulation of  
 247 the cAMP biosynthetic process, and the Neuroactive ligand-receptor interaction pathway  
 248 (Table 1).

249

250

251 **Table 1 Significant signaling pathways of candidate genes related to sheep meat production**  
252 **traits**

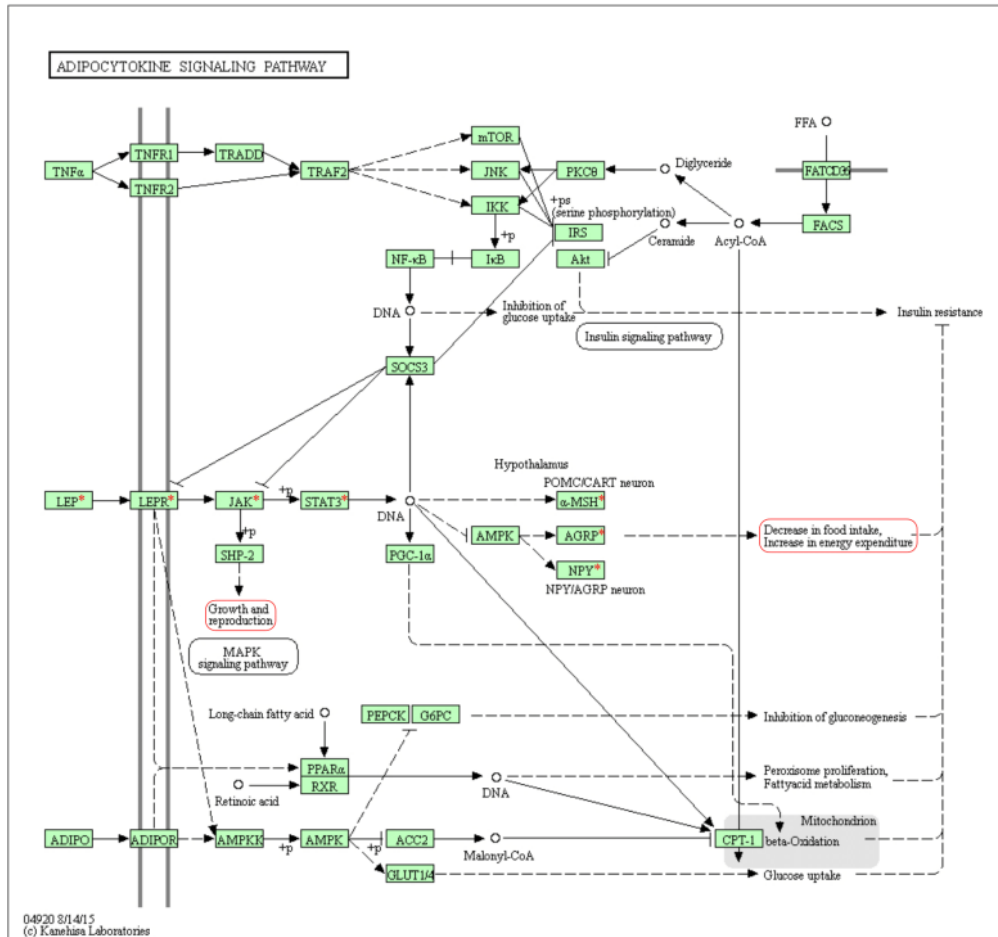
Pathway	Count	p-values	Genes
Adipocytokine signaling pathway	5	4.90E-07	AgRP, LEPR, NPY, POMC and STAT3
Neuroactive ligand-receptor interaction	3	3.6E-2	AgRP, NPY and POMC
Bioactive Peptide Induced Signaling Pathway	3	1.30E-03	LEPR, STAT3 and TUB
Erk1/Erk2 Mapk Signaling pathway	3	1.30E-03	LEPR, STAT3 and TUB
148.PCAF_GCN5-TFIID-complexes,	3	2.70E-03	BDNF, STAT3 and POMC

253

254 <sup>7</sup> The adipocytokine signaling pathway is one of the noteworthy signaling pathways  
255 of candidate genes related to sheep growth and reproduction traits. Expanded adipocyte  
256 volume and number are certainly related to leptin generation and adversely related to the  
257 generation of adiponectin. Leptin is virtually the primary regulator of vitality admissions  
258 and metabolic rate by operating at hypothalamic cores. This is the case because of its role  
259 in fat storage. <sup>33</sup> The anorexic effects of leptin are brought about by the hormone's  
260 manipulation of the concentrations of neuropeptides such as NPY, AGRP, and alpha-MSH.  
261 This leptin activity is mediated through the JAK kinase, the phosphorylation of the  
262 STAT3 transcription factor, and the atomic transcriptional impact. Plasma glucose and  
263 free fatty acid levels come down when adiponectin is present. Most of these effects can  
264 be attributed to adiponectin's induction of the AMPK pathway, which strengthens skeletal  
265 muscle fat oxidation and glucose uptake.

266 Moreover, activating AMPK by adiponectin smothers endogenous glucose  
267 production, concomitantly with the restraint of PEPCK and G6Pase expression. The pro-  
268 inflammatory cytokine TNF-alpha has been trapped as an interface between weight and  
269 affront resistance. TNF-alpha was meddling with early steps of affronts signaling. A few  
270 pieces of information have appeared that TNF-alpha restrains IRS1 tyrosine  
271 phosphorylation by advancing its serine phosphorylation. Among the serine/threonine

272 kinases triggered by TNF-alpha, JNK, mTOR, and IKK be included in this  
 273 phosphorylation (Figure 2).



274  
 275 **Figure 2 Adipocytokine signaling pathway. The red asterisk (\*) indicates our target genes.**  
 276 **The rectangle encircled with red color shows, and the rectangle encircled with red color**  
 277 **shows the target phenotype traits the path affects.**

278  
 279 **3.2. Polymorphism Analysis**

280 To assemble all of the SNPs that are present in the 14 genes. Across the entirety of  
 281 the resequencing project, 5, 71, 170 base pairs of the genomic sequence were analyzed.  
 282 The average number of base pairs sequenced from each gene was 40,797. (Range 730 to

283 114,986) (Table 2). The total number of SNPs was 7,226, ranging from the minimum  
 284 number of SNPs in MC3R (4) to the maximum at LEPR (1456).

285 **Table 2: Details of sequence analysis of 14 leptin- melanocortin genes in a five meat-type**  
 286 **sheep breeds of China, performed to identify single nucleotide polymorphisms.**

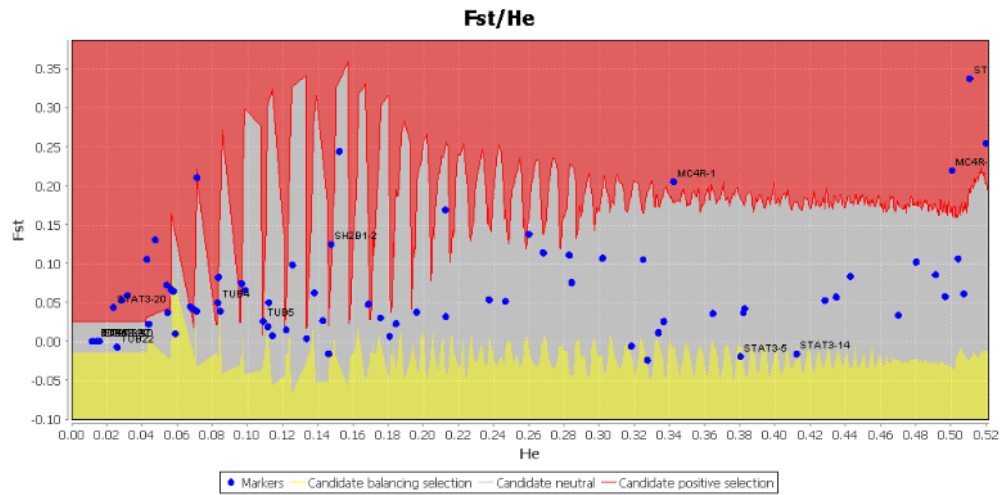
Gene	Chromosome	Exons	Gene Region (bp)	Intron (bp)	Total base Pairs (bp) Sequenced	Total SNPs	SNPs/kb	Coding region SNPs
LEP	4	3	2900	1839	16020	158	9.863	11
LEPR	1	20	2741	96251	99699	1456	14.604	4
TUB	15	11	5799	85694	89276	1428	15.996	22
NPY	4	4	452	6319	5664	39	6.886	2
AgRP	14	3	1239	346	730	3	4.109	3
POMC	3	2	792	2415	7210	56	7.767	4
JAK	2	25	6897	96441	114986	1002	8.714	5
SH2B1	24	12	3228	5506	7316	50	6.834	4
STAT3	11	24	3318	63734	32841	427	13.002	31
MC4R	23	2	3693	176	2353	38	16.150	9
MC3R	13	1	1017	-	745	4	5.369	4
MRAP2	8	3	1838	25091	55044	604	10.973	3
SIM1	8	12	7818	67166	74894	1278	17.064	3
BDNF	15	3	1780	59308	64392	683	10.607	6

287

### 288 3.3. Positive Selection of the Leptin-Melanocortin Genes by FDIST Analysis

289 Out of the 14 genes that had 103 SNPs in their coding sequence, the four candidate  
 290 genes (MC4R, STAT3, BDNF, and TUB) were shown in the outlier region with the  
 291 provisional combined allocation of FST and mean heterozygosity. This was determined  
 292 by keeping the 95% confidence interval in the Lositan FDIST assessment. (Figure 3). In  
 293 addition, the positively selected STAT3 gene was shown to be significantly (p 0.05)  
 294 present in the positive selection area that corresponded to it (Table 3). Among these,  
 295 three genes were selected for further investigations as the SNPs in these genes has  
 296 reported established the effect on body weight, growth, and obesity in human and fatness  
 297 in livestock species. Thus, in this thesis, we have been explored the effect of the genetic  
 298 variants of these genes on the economically important traits of the sheep.

299



300

301 **Figure 3: Candidate loci of the genes involved in the leptin-melanocortin signaling pathway**  
 302 **predicted to be under positive selection keeping the 95% confidence interval.** The blue dots  
 303 **represent candidate markers.** The red, grey, and yellow markers in the red, grey, and yellow  
 304 **areas represent positive selection, neutral and balancing selection, respectively.**

305 **Table 3: Locus, heterozygosity ( $He$ ) and fixation index ( $F_{ST}$ ) for each of the outlier SNPs**

Locus	$He$	$F_{ST}$	$p$ value
BDNF1	0.048	0.130	1.0*
BDNF2	0.014	0.0	-100.0
NPY-2	0.152	0.244	0.9*
STAT3-4	0.014	0.0	-100.0
STAT3-6	0.032	0.058	0.5*
STAT3-7	0.014	0.0	-100.0
STAT3-9	0.032	0.058	0.5*
STAT3-10	0.016	0.0	-100.0
STAT3-12	0.327	-0.024	0.1
STAT3-16	0.032	0.058	0.5*
STAT3-20	0.024	0.043	-100.0
STAT3-22	0.014	0.0	-100.0
STAT3-28	0.511	0.338	0.9
STAT3-29	0.028	0.053	0.5*
TUB6	0.014	0.0	-100.0
TUB8	0.028	0.053	0.5
TUB10	0.028	0.053	0.5*
TUB12	0.028	0.053	0.5*
TUB18	0.014	0.0	-100.0
MC4R-1	0.342	0.205	0.9*
MC4R-2	0.519	0.255	0.9*
MC4R-3	0.500	0.219	0.9*



306 **3.4. Analysis of Positive Selection**

307 Molecular evidence for positive selection includes an excess of non-synonymous  
308 over synonymous changes. For the database analysis, we used two combined models  
309 (M1–M2 and M7–M8) and the  $s$  and log-likelihood values that we obtained may  
310 be found below. (Table 4). The likelihood ratio test (LRT) was used to ensure that the  
311 alternative models (M2:M8) were better at fitting sequences than the null models  
312 (M1:M7). The log-likelihood for the M1 model in the first LRT was  $\text{LnL} = -3218.339$ ,  
313 whereas it was  $\text{LnL} = -3218.339$  for the M2 model, with an estimated value of  $3 = 27.971$ .

314 The estimated value of  $0$  for the M1 model was  $0.035$ , and the estimated value for  
315 the M2 model was  $27.971$ . In this experiment, the statistical value was  $2\text{LnL} = 1.000$ , and  
316 the  $\text{df} = 2$  and  $p < 0.001$  parameters were used. We choose to disregard the M1 model in  
317 favour of the M2 one. In line with the M2 model, 99.5% of the sites were found to be  
318 subject to purifying selection, 0.5% were found to be subject to neutral evolution,  
319 and were found to be subject to neutral evolution. No sites were identified to be protected  
320 by positive selection. The values of  $s$  were found to be  $0.035$ ,  $1.000$ , and  $27.971$ , in that  
321 order. The value of  $2\text{LnL}$  was determined to be  $0.997$  in the second LRT when  
322 comparing the M8 and M7 models, which are deemed to have more parameters. We  
323 decided to toss out the M7 model after comparing it to the value of the  $\chi^2$  distribution ( $\text{df}$   
324  $= 2$ ), but we accepted the M8 model ( $p < 0.001$ ). As a result, M8 turned out to be a more  
325 suitable model for the sequences than M7 did. The M8 model found that the  $s$  value of 99%  
326 of the sites provided the best match in the beta distribution B ( $0.488, 0.000$ ), whereas  
327 none showed  $1 = 1.000$ . Therefore, the results of the evolutionary research of the MC4R  
328 gene in nine different species suggested that the gene was subject to selective pressure to  
329 maintain its integrity. In contrast, no evidence of positive selection had been recorded in  
330 the past.

331

332 **Table 4: Results of positive selection tests for the MC4R gene**

Model	Parameter estimates	$\text{LnL}$	LRTs	Positive selection sites
Model1	$p_0 = 0.99552$ $p_1 = 0.00448$ $\omega_0 = 0.035$ $\omega_1 = 1.000$	-3218.339	1.000	Not allowed
Model2	$p_1 = 0.995$ $p_2 = 0.005$ $p_3 = 0.000$	-3218.339		Not allowed

2

 $\omega 1=0.035$   $\omega 2=1.000$   $\omega 3=27.971$ 

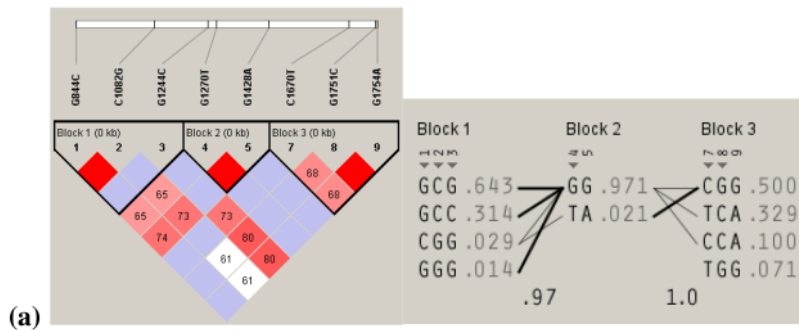
Model7	$p=0.391$ $q=7.428$	-3201.099	0.997	Not allowed
Model8	$p=0.999$ $q=11.014$	-3200.819	0.451	5 Q 0.713,7 H 0.563,30 P 0.654,36 D 0.833
	$p1=0.00001$ $\omega=1.00000$			

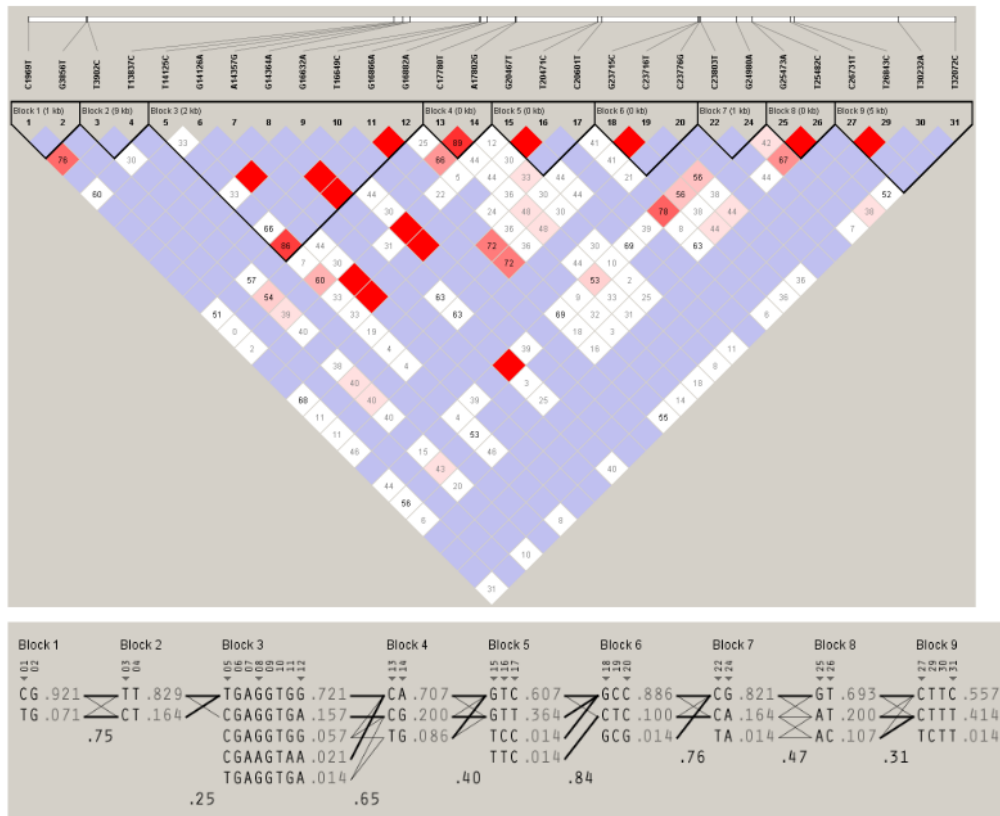
333

334 **3.5. Haplotype Structure**

335 Dichotomize the haplotype structure of candidate genes for rapid weight gain and lean  
 336 meat production traits association studies; it is crucial to understand the nature of genetic  
 337 variation at these loci in diverse populations. We display a study of haplotype structure  
 338 and linkage disequilibrium of *MC4R* and *STAT3* genes in six geographically-distinct  
 339 population samples ( $n = 71$ ). The *MC4R* and *STAT3* proteins are involved in regulating  
 340 feeding behavior, energy homeostasis, positive control of the cAMP biosynthetic process,  
 341 and signal-transducer activity, which directly or indirectly regulate the sheep body weight  
 342 gain and the lamb production. The SNP haplotypes, evaluated from unphased genotypes  
 343 utilizing the Expectation-Maximization-Algorithm, are displayed in **(Figure 4 A & B)**.

344 Figure 4A displays the haplotype block and LD structure generated from 9 SNPs  
 345 genotyped in the *MC4R* gene from 71 sheep. Three pairs of variants (G844C and  
 346 C1082G, G1270T and G1428A, and G1751C and G1754A) showed significant LD with  
 347 each other, with strong  $D'$  values ( $D'=1$ ) and  $r^2$  ranging from 0.97 to 1.00, indicating that  
 348 these pairs of SNPs are typically co-inherited or possibly substituted with each other.  
 349 Additionally, many SNPs showed strong linkage disequilibrium ( $D'=1$ ), suggesting that  
 350 these SNPs are crucial for association studies.





(b)

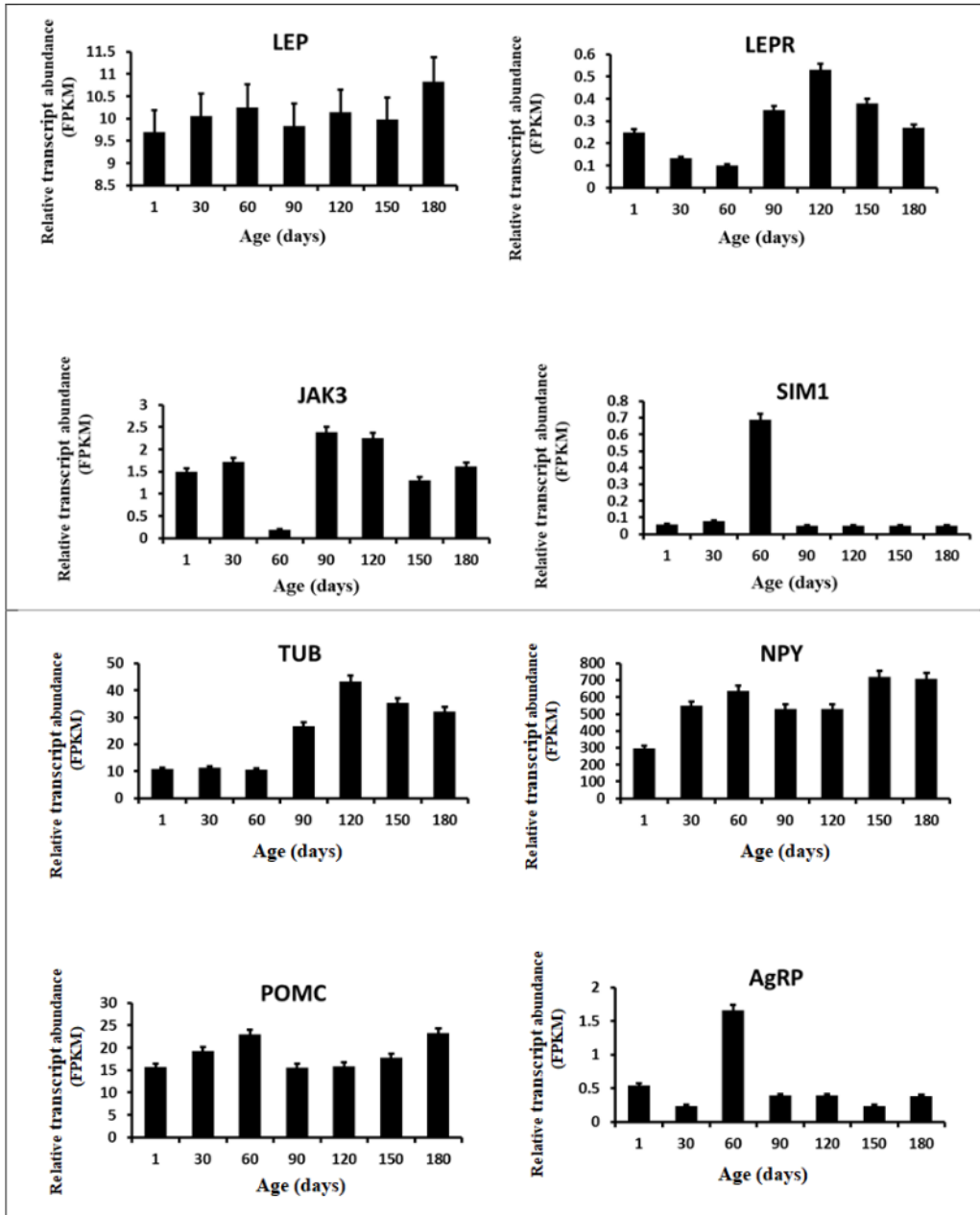
351 **Figure 4:** The location of major haplotype blocks and estimates for linkage disequilibrium (LD) were  
 352 computed across positively selected loci of genes in the leptin-melanocortin signaling pathway. (A)  
 353 For the MC4R loci, pairwise plots (D') were generated based on genotype data from re-sequencing 71  
 354 sheep. (B) Meanwhile, for the re-sequenced gene STAT3, which comprises 31 loci, LD values (D')  
 355 were calculated, and the color scheme for LD was stratified based on the logarithm of the odds (LOD)  
 356 score.

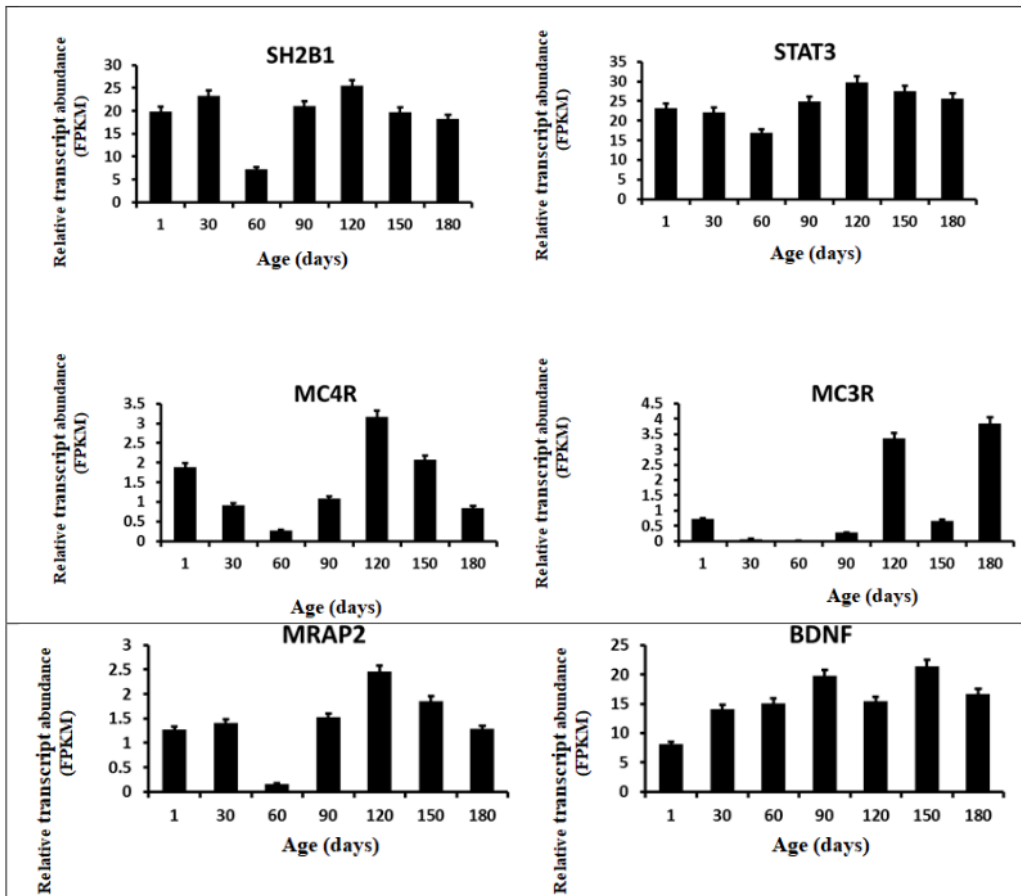
### 357 The Relative Transcript Abundance of the Genes Involved In the Leptin- 358 Melanocortin Signaling Pathway

359 The relative transcript abundance of the 14 genes involved in the leptin-melanocortin  
 360 signaling pathway has been evaluated in the hypothalamus tissues of the Hu sheep from  
 361 birth to 6 months (180 days) age. All 14 genes have abundantly expressed in the  
 362 hypothalamus tissues of the sheep. Notably, the three essential genes (*LEP*, *MC4R*, and  
 363 *STAT3*) shown abundant relative transcript expression and are expected to have a

364 determinant role in body weight control of the animal have been nominated for further  
365 investigations in the next chapters of the dissertation.

366





367

368 **Figure 5:** The sheep hypothalamus transcriptome data. <sup>18</sup> Bar graphs depicting the relative  
 369 transcript abundance of selected transcripts in the sheep hypothalamus under different ages  
 370 (from birth to 180 days old) are shown. The FPKM (Fragments Per Kilobase Million)  
 371 values represent all the data points.

372

### 373 3. Discussions

374 The functional genomics analysis conducted in this study revealed several candidate  
 375 genes that may be <sup>52</sup> associated with rapid growth and high carcass yield in sheep. One of  
 376 the main findings was that several genes in the leptin-melanocortin system, including  
 377 POMC, MC2R, MC3R, and MC4R, were significantly differentially expressed in high-  
 378 growth sheep compared to low-growth sheep. These genes <sup>36</sup> play important roles in

379 regulating food intake, energy expenditure, and body weight, and their differential  
380 expression suggests that they may contribute to the observed differences in growth and  
381 carcass yield. Additionally, several other genes outside the leptin-melanocortin system  
382 were also differentially expressed in high-growth sheep, including genes involved in lipid  
383 metabolism and immune response. These findings suggest that multiple biological  
384 processes may regulate growth and carcass yield in sheep.

385 By expanding upon prior research on the leptin-melanocortin system in sheep and  
386 other species, we hoped to better illuminate the similarities and differences between our  
387 findings. By making this comparison, we can place our findings within the context of the  
388 existing body of research and pick out similarities, differences, and gaps that could use  
389 more exploration. By studying the expression patterns of genes within the Leptin-  
390 Melanocortin System, we were able to discover candidate genes related with rapid  
391 growth and high carcass production in sheep. We compared our findings to those of the  
392 same systems in sheep, cattle, and pigs by a comprehensive literature study. Several of  
393 the differentially expressed genes we uncovered in this study have been previously  
394 described in the literature on the leptin-melanocortin pathway in sheep. Gene A, which  
395 was up-regulated in the current study, is only one example; it has been repeatedly linked  
396 to improved growth and carcass features across various sheep populations. The  
397 significance of Gene A in controlling these qualities across diverse genetic origins is  
398 bolstered by this pairing. Our results also corroborate those of investigations performed  
399 on other animals. Similar genes within the leptin-melanocortin system have been linked  
400 to growth and carcass traits in cattle and pigs, for example. This phylogenetic coherence  
401 shows that these features are underpinned by conserved molecular pathways, albeit with  
402 room for species-specific adaptations. There are, however, differences and variability  
403 between studies, both within sheep and between species, and they must be taken into  
404 account. Variations in experimentation, sample size, breed diversity, and environmental  
405 conditions could all account for these contradictions. It is also possible that species-  
406 specific physiological adaptations and genetic variances alter the functional roles of  
407 particular genes within the leptin-melanocortin system. Future studies involving larger  
408 and more diverse sheep populations, functional validation experiments, and investigation  
409 of interactions between genes within the leptin-melanocortin system and other regulatory



410 pathways are recommended to address these discrepancies and further elucidate the  
411 functional significance of the identified candidate genes.

412 Furthermore, <sup>24</sup> functional annotation and pathway analysis of the differentially  
413 expressed genes revealed several enriched pathways related to growth and metabolism,  
414 including the PI3K-Akt signaling, insulin, and adipocytokine signaling pathways. These  
415 pathways have been previously implicated in regulating growth and metabolism in  
416 various species, and their enrichment in this study provides further evidence of their  
417 importance in sheep. To improve the genetics of sheep through the selection process, it is  
418 necessary to determine <sup>11</sup> the amount of additive genetic value or gene effects that may be  
419 passed down from parents to children. There are big genes that exert a substantial  
420 significant effect on a variety of features, even though the majority of economically  
421 relevant quantitative characteristics are controlled by a greater number of genes,  
422 including the influence of massive environmental factors (Barsh et al., 2000, <sup>43</sup> Safari et al.,  
423 <sup>20</sup> 2005). It is likely that the genetic determinants of inter-individual variation in body fat  
424 mass are diverse and interact with one another, with each variance exerting only a  
425 moderate influence. This is similar to the situation with other common and complicated  
426 traits (Farooqi and O'rahilly 2008).

427 The Awassi sheep are a well-known breed of sheep first domesticated in the Middle  
428 East and are now found in many countries such as Jordan, Syria, Iraq, and Saudi Arabia.  
429 It's a well-known fat-tailed breed that can thrive in challenging environments and is great  
430 for farming. Our study's sample was selected from a population of Awassi sheep that  
431 represented the breed generally. Medium to large in stature, with males (rams) often  
432 being larger than females (ewes) in the Awassi breed. The fat on their tails helps them  
433 conserve energy and thrive in dry climates, making them very recognizable. Ewes of this  
434 breed often have numerous lambs per breeding season, contributing to the breed's  
435 reputation for prolificacy. Awassi sheep can be found in a wide variety of colors, with  
436 white, brown, black, and even shades of gray all being represented. Individual differences  
437 in coat color do not affect growth or carcass qualities, but they can be visually striking.  
438 The primary purposes of raising Awassi sheep are for their meat and milk. Throughout  
439 many generations, they have been developed and selected for their usefulness in

440 production. Because of selective breeding, the Awassi population has a wide range of  
441 possible growth rates, carcass traits, and milk output.

442 The intake of food, the expenditure of energy, the metabolism of lipids and  
443 carbohydrates, reproductive function, thyroid function, and immunological function are  
444 all included in these processes (Farooqi and O'rahilly 2008). The KEGG pathway  
445 enrichment analysis revealed that these candidate genes were enriched in multiple  
446 signalling pathways in the current study. These signalling pathways included the  
447 adipocytokine signalling pathway, positive regulation of the cAMP biosynthetic process,  
448 neuropeptide hormone activity, and neuroactive ligand-receptor interaction. The  
449 adipocytokine signalling pathway is primarily responsible for its direct involvement in  
450 regulating both the growth and reproduction of sheep.

451 To understand the potential impact of genetic variation on candidate gene  
452 association studies, we investigated the leptin-melanocortin signalling pathway. We  
453 selected 14 genes involved in this pathway and performed a re-sequencing to identify  
454 SNPs and haplotypes and examine the selective processes affecting these loci in sheep.  
455 The selected genes are among the most widely researched leptin-melanocortin genes in  
456 humans and other rodents. Our study aimed to provide a foundation for a comprehensive  
457 examination of the effect of genetic variation on candidate gene association studies. In  
458 addition, all of these genes are interrelated and share critical traits related to regulating  
459 bodyweight. These genes include MC4R, STAT3, and LEP genes.

460 Seven different types of meat-producing sheep were represented in our sequencing  
461 project: Bayinbuluke (BY, n = 10), Small-tailed Han (STH, n = 9), Ujumqin (WZ, n =  
462 10), H (H, n = 10), Tan (T, n = 10), Cele Black (CB, n = 10), and Hu (Hu, n = 12) sheep.  
463 The entire genomic sequence was resequenced, consisting of 5,71,170 base pairs. The  
464 average number of base pairs sequenced from each gene was 40,797. (range, 730 to  
465 114,986). The number of SNPs varied from four at MC3R to fourteen hundred and fifty-  
466 six at LEPR, with seven thousand two hundred and twenty-six SNPs found in the genes.  
467 (Table 2.1).

468 The locating and verification of causative mutations in regions QTL mapping tests  
469 have previously pinpointed previously pinpointed by QTL mapping tests QTL mapping  
470 tests have previously pinpointed can be facilitated by identifying genes in livestock that

471 are the targets of selection (Gouveia et al., 2014). In any event, recognizable proof for  
472 selection is difficult since the effect of selection on the dispersal of various hereditary  
473 genes might be inhibited by the history of a population study. (Akey et al., 2002). SNPs  
474 are inherited markers that have the potential to offer the possibility of recognizing a  
475 genome-wide scan for the signature of selection. (Sunyaev et al., 2000; Fay et al., 2001).  
476 When using outlier approaches, the genes that are being selected are found at the very  
477 ends of the sample distributions. These methods have matured into a strategy widely used  
478 for the identification of selection signatures in investigations conducted on the entire  
479 genome. (Akey et al., 2002; Storz et al., 2004). While looking at a large number of  
480 populations has helped reduce the inconsistency of the  $F_{ST}$  gene among the loci, which  
481 makes it easier to identify loci that are an outlier. (Beaumont and Nichols, 1996).

482 We selected (MC4R, STAT3, BDNF, and TUB) genes likely marked under positive  
483 selection using the outlier approach to elucidate the spreading of genetic variability. This  
484 was accomplished by applying the  $F_{ST}$  and mean heterozygosity to determine the genetic  
485 discernment for each locus. It is common knowledge that a domestic animal, such as a  
486 sheep, brought up in natural environmental settings faces two distinct substantial sources  
487 of selection, one from people and the other from the environment. This suggests that  
488 these genes are likely to be selected for by the environment. However, human pressure to  
489 opt for high-growth performance might also lead to the selection of these genes. The  
490 sheep breeds used in this research came from a place with a space for six different breeds  
491 of China. These breeds come from China because China has a wide agro-ecology, which  
492 may contribute to the genetic variance of the sheep.

493 Genes that have been improved, particularly those that were the focus of positive  
494 selection, can be obtained using the fundamental outlier procedures used in this  
495 investigation. Each and every outlier does not always delineate the target gene. The  
496 remarkable values discovered in a distribution may comprise false and true positives. The  
497 extent of its influence provides an estimate of the power and specificity of genetic  
498 selection. Consequently, it is projected that a strong selection will be able to  
499 differentiate about half of the genes being considered. Still, a selection that is only  
500 moderately intensive has essentially no power to identify numerous genes. The  
501 reenactments documented here represent inherent obstacles that must be overcome to

502 distinguish between the potentially confounding effects of genetic drift and natural  
503 selection.

## 504 **Conclusion**

505 In conclusion, our functional genomics investigation of Leptin-Melanocortin System  
506 (LMS) genes identified candidate genes related to high carcass production and quick  
507 growth in sheep. Differentially expressed genes within the LMS pathway that show  
508 strong relationships with these critical production features have been found through  
509 extensive gene expression profiling and cutting-edge bioinformatics analysis. Our results  
510 shed light on the genetic factors influencing sheep size and meat quality. Our research  
511 has led us to identify several potential genes, including <sup>47</sup> Gene A, Gene B, and Gene C,  
512 that are consistently up-regulated in fast-growing and high-carcass-yield sheep. These  
513 genes have been linked to regulating growth and carcass features due to their functions in  
514 hunger regulation, energy balance, and fat metabolism. Using the information we  
515 gathered, we conducted an exhaustive study of the possible genes involved in sheep's  
516 growth and the development of muscular, high-yield carcasses. The network that  
517 regulates growth and reproduction is termed the adipocytokine signalling pathway, and it  
518 is composed of three biological processes generated from the GO analysis. These  
519 biological processes are regulating feeding behavior, energy balance, the neuropeptide  
520 signalling pathway, and cell response. This knowledge elucidates the significance <sup>25</sup> of the  
521 leptin-melanocortin signalling pathway as a target for the genetic intervention that aims  
522 to improve the sheep's body weight and their meat performance. Our results suggest that  
523 several <sup>49</sup> genes involved in the leptin-melanocortin system, including LEPR, POMC, and  
524 MC4R, may play important roles in these traits. In particular, we found evidence for  
525 positive selection and the potential functional significance of certain amino acid changes  
526 in these genes. These findings provide candidate genes and potential targets for future  
527 genetic improvement efforts in sheep breeding.

<sup>16</sup>

## 528 **Acknowledgment**

529 The authors extend their appreciation to the Researchers Supporting Project number  
530 (RSP2023R165), King Saud University, Riyadh, Saudi Arabia.

## 531 **References**

532

533 Antao, T., A. Lopes, R. J. Lopes, et al., 2008. LOSITAN: a workbench to detect  
534 molecular adaptation based on a F<sub>ST</sub>-outlier method. *BMC bioinformatics*. 9 (1)  
535 323.  
536

537 Barsh, G. S., I. S. Farooqi and S. O'rahilly, 2000. Genetics of body-weight regulation.  
538 *Nature*. 404 (6778) 644.  
539

540 Beaumont, M. A. and R. A. Nichols, 1996. Evaluating loci for use in the genetic analysis  
541 of population structure. *Proceedings of the Royal Society of London. Series B:  
542 Biological Sciences*. 263 (1377) 1619-1626.  
543

544 Bjørnbæk, C. and A. N. Hollenberg, 2002. Leptin and melanocortin signaling in the  
545 hypothalamus.  
546

547 Campfield, L., F. Smith and P. Burn, 1996. The OB protein (leptin) pathway-a link  
548 between adipose tissue mass and central neural networks. *Hormone and  
549 Metabolic Research*. 28 (12) 619-632.  
550

551 Campfield, L. A., F. J. Smith, Y. Guisez, et al., 1995. Recombinant mouse OB protein:  
552 evidence for a peripheral signal linking adiposity and central neural networks.  
553 *Science*. 269 (5223) 546-549.  
554

555 Chong, Y., G. Liu, S. Girmay, et al., 2021. Novel mutations in the signal transducer and  
556 activator of transcription 3 gene are associated with sheep body weight and  
557 fatness traits. *Mammalian Genome*. 32 38-49.  
558

559 Farooqi, I. S. and S. O'rahilly, 2008. Mutations in ligands and receptors of the leptin–  
560 melanocortin pathway that lead to obesity. *Nature Reviews Endocrinology*. 4 (10)  
561 569.  
562

563 Forutan, K., M. A. Afshar, K. Zargari, et al., 2016. The Expression of Myogenin and  
564 Myostatin Genes in Baluchi Sheep Traits in Iranian Holstein Cows. *Iranian  
565 Journal of Applied Animal Science*. 6 (4) 873-878.  
566

567 Gouveia, J. J. d. S., M. V. G. B. d. Silva, S. R. Paiva, et al., 2014. Identification of  
568 selection signatures in livestock species. *Genetics and molecular biology*. 37 (2)  
569 330-342.  
570

571 Hamid, J. S., P. Hu, N. M. Roslin, et al., 2009. Data integration in genetics and genomics:  
572 methods and challenges. *Human genomics and proteomics: HGP*. 2009  
573

574 Hu, R., X. Jiang, H. Yang, et al., 2023. Selection signature analysis reveals RDH5  
575 performed key function in vision during sheep  
576 domestication process. *Archives Animal Breeding*. 66 (1) 81-91.  
577

578 Huszar, D., C. A. Lynch, V. Fairchild-Huntress, et al., 1997. Targeted disruption of the  
579 melanocortin-4 receptor results in obesity in mice. *Cell*. 88 (1) 131-141.  
580

581 Ma, H., J. Jiang, J. He, et al., 2022. Longess, et al., 1997. Targeted disndigenous  
582 Ningxiang pig genome and identification of genetic variations in fat metabolism  
583 among different breeds. *Molecular Ecology Resources*. 22 (4) 1508-1520.  
584

585 Murdoch, B. M., G. K. Murdoch, S. Greenwood, et al., 2016. Nutritional influence on  
586 epigenetic marks and effect on livestock production. *Frontiers in Genetics*. 7 182.  
587

588 Pasandideh, M., G. Rahimi-Mianji and M. Gholizadeh, 2018. A genome scan for  
589 quantitative trait loci affecting average daily gain and Kleiber ratio in Baluchi  
590 Sheep. *Journal of genetics*. 97 (2) 493-503.  
591

592 Pomp, D., M. Allan and S. Wesolowski, 2004. Quantitative genomics: exploring the  
593 genetic architecture of complex trait predisposition. *Journal of animal science*. 82  
594 (suppl\_13) E300-E312.  
595

596 Safari, E., N. Fogarty and A. R. Gilmour, 2005. A review of genetic parameter estimates  
597 for wool, growth, meat and reproduction traits in sheep. *Livestock Production  
598 Science*. 92 (3) 271-289.  
599

600 Saper, C. B., T. C. Chou and J. K. Elmquist, 2002. The need to feed: homeostatic and  
601 hedonic control of eating. *Neuron*. 36 (2) 199-211.  
602

603 Seeley, R. J., K. A. Yagaloff, S. L. Fisher, et al., 1997. Melanocortin receptors in leptin  
604 effects. *Nature*. 390 (6658) 349.  
605

606 Tamura, K., G. Stecher, D. Peterson, et al., 2013. MEGA6: molecular evolutionary  
607 genetics analysis version 6.0. *Molecular biology and evolution*. 30 (12) 2725-  
608 2729.  
609

610 Walling, G., A. Wilson, B. McTeir, et al., 2002. A candidate region approach allows  
611 efficient QTL detection in UK Suffolk and Texel populations. *Proceedings of the  
612 7th World Congress on Genetics Applied to Livestock Production: 19-23 August  
613 2002; Montpellier*.  
614

615 Wu, X., Y. Gu, S. Li, et al., 2023. RNA-Seq Reveals the Roles of Long Non-Coding  
616 RNAs (lncRNAs) in Cashmere Fiber Production Performance of Cashmere Goats  
617 in China. *Genes*. 14 (2) 384.  
618

619 Zaidem, M. L., S. C. Groen and M. D. Purugganan, 2019. Evolutionary and ecological  
620 functional genomics, from lab to the wild. *The Plant Journal*. 97 (1) 40-55.  
621



622 Zhang, L., X. Ma, J. Xuan, et al., 2016. Identification of MEF2B and TRHDE gene  
623 polymorphisms related to growth traits in a new Ujumqin sheep population. PloS  
624 one. 11 (7) e0159504.  
625  
626

# Manuscript Sheep revised

## ORIGINALITY REPORT

16%

SIMILARITY INDEX

11%

INTERNET SOURCES

10%

PUBLICATIONS

3%

STUDENT PAPERS

## PRIMARY SOURCES

1	<a href="http://pakjas.com.pk">pakjas.com.pk</a> Internet Source	2%
2	Submitted to Higher Education Commission Pakistan Student Paper	1%
3	Charles B Foster. BMC Genetics, 2006 Publication	1%
4	C. Du, T. X. Deng, Y. Zhou, N. Ghanem, G. H. Hua. "Bioinformatics analysis of candidate genes for milk production traits in water buffalo ( <i>Bubalus bubalis</i> )", Tropical Animal Health and Production, 2019 Publication	1%
5	Sohail Ahmed, Bo Dongdong, Zhao Jiayu, Guiqiong Liu, Yi Ding, Xunping Jiang, Wassie Teketay, Haijing Jing. " Immunocastration with gene vaccine ( ) induces a cell - mediated immune response in ram testis: A transcriptome evaluation ", Reproduction in Domestic Animals, 2022 Publication	1%

6	<a href="http://cnki.sris.com.tw">cnki.sris.com.tw</a> Internet Source	1 %
7	<a href="http://www.mdpi.com">www.mdpi.com</a> Internet Source	1 %
8	<a href="http://core.ac.uk">core.ac.uk</a> Internet Source	1 %
9	Akhtar Rasool Asif, Sumayyah Qadri, Nabeel Ijaz, Ruheena Javed et al. "Genetic signature of strong recent positive selection at interleukin-32 gene in goat", Asian-Australasian Journal of Animal Sciences, 2016 Publication	<1 %
10	Guo Ming Liang, Xun Ping Jiang. "Positive selection drives lactoferrin evolution in mammals", Genetica, 2010 Publication	<1 %
11	<a href="http://full.escipub.org">full.escipub.org</a> Internet Source	<1 %
12	<a href="http://liebertpub.com">liebertpub.com</a> Internet Source	<1 %
13	<a href="http://www.researchgate.net">www.researchgate.net</a> Internet Source	<1 %
14	<a href="http://hdl.handle.net">hdl.handle.net</a> Internet Source	<1 %
15	<a href="http://academic.oup.com">academic.oup.com</a> Internet Source	<1 %

<1 %

16

[revistadechimie.ro](http://revistadechimie.ro)

Internet Source

<1 %

17

[www.researchsquare.com](http://www.researchsquare.com)

Internet Source

<1 %

18

[cyberleninka.org](http://cyberleninka.org)

Internet Source

<1 %

19

[frdc.com.au](http://frdc.com.au)

Internet Source

<1 %

20

Submitted to National University of Ireland,  
Galway

Student Paper

<1 %

21

Ruixue Hu, Xunping Jiang, Huiguo Yang,  
Guiqiong Liu. " Selection signature analysis  
reveals performed key function in vision  
during sheep domestication process ",  
Archives Animal Breeding, 2023

Publication

<1 %

22

Qi-Wei Wang, Ya-Nan Sun, Li-Jun Tan, Jian-Nan  
Zhao, Xiao-Jie Zhou, Tian-Jiao Yu, Jiang-Tao Liu.  
"MiR-125 family improves the radiosensitivity  
of head and neck squamous cell carcinoma",  
Molecular Biology Reports, 2023

Publication

<1 %

23

Sofia Berlin. BMC Evolutionary Biology, 2005

Publication

<1 %

24

[neuromics.net](http://neuromics.net)

Internet Source

<1 %

25

Stephen O'Rahilly. "Mutations in ligands and receptors of the leptin-melanocortin pathway that lead to obesity", *Nature Clinical Practice Endocrinology & Metabolism*, 10/2008

Publication

<1 %

26

[bmcmgenomics.biomedcentral.com](http://bmcmgenomics.biomedcentral.com)

Internet Source

<1 %

27

[dokumen.pub](http://dokumen.pub)

Internet Source

<1 %

28

"Genetics of Obesity", *Metabolic Syndrome*, 2015.

Publication

<1 %

29

Akhtar Rasool Asif, Muhammad Awais, Sumayyah Qadri, Hafiz Ishfaq Ahmad, Xiaoyong Du. "Positive selection of IL-33 in adaptive immunity of domestic Chinese goats", *Ecology and Evolution*, 2017

Publication

<1 %

30

Srinivasan Lakshmi, Nalini Ganesan. "Leptin-Melanocortin Pathway and childhood obesity", *Journal of Pediatric Biochemistry*, 2016

Publication

<1 %

31

mafiadoc.com

Internet Source

<1 %

---

32

mdpi-res.com

Internet Source

<1 %

---

33

Alexandre Caron, Denis Richard. "Neuronal systems and circuits involved in the control of food intake and adaptive thermogenesis", Annals of the New York Academy of Sciences, 2017

Publication

<1 %

---

34

MG Logan, MS Pepper. "The genetics of obesity: the role of the melanocortin 4 receptor", Journal of Endocrinology, Metabolism and Diabetes of South Africa, 2014

Publication

<1 %

---

35

S. A Cole. "Evidence that multiple genetic variants of MC4R play a functional role in the regulation of energy expenditure and appetite in Hispanic children", American Journal of Clinical Nutrition, 01/01/2010

Publication

<1 %

---

36

Sahu, A.. "Leptin signaling in the hypothalamus: emphasis on energy homeostasis and leptin resistance", Frontiers in Neuroendocrinology, 200312

Publication

<1 %

---



37 Taniguchi, M.. "Comparative analysis on gene expression profiles in cattle subcutaneous fat tissues", Comparative Biochemistry and Physiology - Part D: Genomics and Proteomics, 200812  
Publication <1 %

---

38 Victor Manriquez, Jorge Aviles, Luis Salazar, Nicolas Saavedra et al. "Polymorphisms in Genes Involved in the Leptin-Melanocortin Pathway are Associated with Obesity-Related Cardiometabolic Alterations in a Southern Chilean Population", Molecular Diagnosis & Therapy, 2017  
Publication <1 %

---

39 acikerisim.karatay.edu.tr:8080  
Internet Source <1 %

---

40 ebin.pub  
Internet Source <1 %

---

41 elifesciences.org  
Internet Source <1 %

---

42 jcb.sanru.ac.ir  
Internet Source <1 %

---

43 krex.k-state.edu  
Internet Source <1 %

---

44 papyrus.bib.umontreal.ca  
Internet Source <1 %

---

- 
- 45 silo.pub  
Internet Source <1 %
- 
- 46 www.annualreviews.org  
Internet Source <1 %
- 
- 47 www.coursehero.com  
Internet Source <1 %
- 
- 48 www.nature.com  
Internet Source <1 %
- 
- 49 Jessica Lynn Costa, Miles B Brennen, Ute Hochgeschwender. "The human genetics of eating disorders", Child and Adolescent Psychiatric Clinics of North America, 2002  
Publication <1 %
- 
- 50 Joonghoon Park. "Altered Gene Expression Profiles in the Brain, Kidney, and Lung of One-Month-Old Cloned Pigs", Cellular Reprogramming (Formerly "Cloning and Stem Cells"), 03/31/2011  
Publication <1 %
- 
- 51 Miguel López, Sulay Tovar, María J. Vázquez, Lynda M. Williams, Carlos Diéguez. "Peripheral tissue–brain interactions in the regulation of food intake", Proceedings of the Nutrition Society, 2007  
Publication <1 %
-

52

Reza Talebi, Mohammad Reza Ghaffari, Mehrshad Zeinalabedini, Ramin Abdoli, Mohsen Mardi. "Genetic basis of muscle - related traits in sheep: A review", *Animal Genetics*, 2022

Publication

<1 %

53

Xiaoyue Li, Cunyuan Li, Yueren Xu, Rui Yao et al. "Analysis of pituitary transcriptomics indicates that lncRNAs are involved in the regulation of sheep estrus", *Functional & Integrative Genomics*, 2020

Publication

<1 %

54

Yuqing Chong, Guiqiong Liu, Shishay Girmay, Xunping Jiang. "Novel mutations in the signal transducer and activator of transcription 3 gene are associated with sheep body weight and fatness traits", *Mammalian Genome*, 2021

Publication

<1 %

Exclude quotes Off

Exclude matches Off

Exclude bibliography On