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Susceptibility Pattern of Multi Drug Resistance *Pseudomonas aeruginosa* Isolates from Tertiary care Hospital in Riyadh KSA

Running title: Susceptibility of Multi Drug Resistance *Pseudomonas aeruginosa*

Abstract

Pseudomonas aeruginosa is important pathogens commonly cause nosocomial infections. The occurrence of multi-resistant organisms (MROs) of *Pseudomonas aeruginosa* strains have been increased worldwide and limiting the therapeutic options. The MRO of *Pseudomonas aeruginosa* phenotype can be mediated by a variety of resistance mechanisms and highly versatile property to mutate.^[1] Therefore the our study aimed to evaluate the resistance pattern of *Pseudomonas aeruginosa* collected from Riyadh tertiary care hospital, Kingdom of Saudi Arabia. During the period from 2019 to 2021 clinical samples were collected from microbiology lab at King Khalid University Hospital and analysed for the antibiotic susceptibility pattern. Results suggested that the rates of resistance for the three years were higher for isolates collected from patients older than 50 years if its compared with the strains collected from young age. A total of 1024 *Pseudomonas aeruginosa* isolates were collected during the last three years, the prevalence rate were 44.6%, 32.6% and 22.7% during the period of 2019, 2020, and 2021 respectively. Meanwhile, the highest percentages of multi drug resistance *Pseudomonas aeruginosa* strains were recovered from body fluids; about 38 (47.5%) out of 80 *Pseudomonas aeruginosa* isolates were MRO *Pseudomonas aeruginosa*. The rate of resistances showed that Imipenem was significantly higher in resistant among the clinical isolates (77.8%), then Meropenem (61%), Aztreonam (42%) and Ceftazidime (36%) than other antibiotics.^[2] Most of isolates were sensitive to colistin except (2.7%) were resistance. Moreover, antibiotic resistant bacteria have been observed with increasing frequency over the past three years. The current study reports that the susceptibility among *P. aeruginosa* isolates have been decreased in KSA, perhaps due to the massive use of antibiotics, the lack of adherence to approved infection control practices by hospitals, or due to the changes to the public health infrastructure.

Keywords: *Pseudomonas aeruginosa*, Multi drug resistance, Antibiotic susceptibility, Clinical isolates

1. Introduction^[0]

Pseudomonas aeruginosa has been responsible for wide range of ICU acquired infections in critically ill patients (Gajdács et al., 2020, Tamma et al., 2021) due to the presence of constitutive resistance to many antibiotics and antiseptics and its ability to acquire further resistance mechanism against multiple classes of antibiotics. *Pseudomonas aeruginosa* is a common pathogen in hospitals and particularly in intensive care units (Höfte 2021). It involves in various life-threatening infection in ICU such as endocarditis and septicemia, urinary tract infections, cystitis, pneumonia, surgical wound infections (Pachori et al., 2019). Various mechanism involve in drug resistance of *Pseudomonas aeruginosa*, like presence of over expressed efflux pump, of acquisition of resistance gene through plasmids and transposons, or through mutation in genes encoding porins, efflux pumps, penicillin-binding proteins, and chromosomal β -lactamase, all contributing to resistance to β -lactams, carbapenems, aminoglycosides, and fluoroquinolones (Normark & Normark 2002). Carbapenem-resistant *Pseudomonas aeruginosa* (CRPA) was first reported in 2001 from a cancer patient in Texas (Walters et al., 2019). *Klebsiella pneumoniae* carbapenemase (KPC) and New Delhi metallo- β -lactamase (NDM) have also been identified in healthcare-associated outbreaks (Epson et al., 2014, Ham et al., 2021, Leung et al., 2013). Recent data from a convenience sample of CRPA tested through the Antibiotic Resistance Laboratory Network found that 1.9% of isolates produced a carbapenemase (Walters et al., 2019). *Pseudomonas aeruginosa* with difficult-to-treat resistance (DTR) was proposed in 2018 and defined as nonsusceptibility to all of the following: piperacillin–tazobactam, ceftazidime, cefepime, aztreonam, meropenem, imipenem–cilastatin, ciprofloxacin, and levofloxacin (Kadri et al., 2018, Bader et al., 2020, Pang et al., 2019).

In this regard, a concern for *P. aeruginosa* infections is in the global emergence of multidrug resistant (MDR) and extensively drug resistant (XDR) strains, which limit the selection of effective antimicrobial therapies (Recio et al., 2020, Martis et al., 2014). In addition, Nosocomial infections caused by this organism are often hard to treat because the intrinsic resistance of the specie and its remarkable ability to acquire further mechanisms of β -lactamases enzymes to multiple groups of antimicrobial agents (García-Betancur et al., 2021).

In addition, to a significant population flow from the Middle East, the Kingdom is also annually a host for more than 4 million Muslim pilgrims from over 180 countries worldwide in the Hajj and Umrah seasons. These factors could potentially make Saudi Arabia a hot spot for the collection and spread of multidrug-resistant strains around the world. Bacterial resistance to antimicrobial agents is on the rise in the Kingdom (Memish et al., 2015)

In current study, we are aiming to analyse the prevalence of *Pseudomonas aeruginosa* infection for last 3 years 2019-2021 from Riyadh tertiary care hospital. Moreover, identifying the pattern of multi-drug resistance clinical isolates of *P. aeruginosa* (MRO). This study could lead insight of understanding the treatment options and control of spread of MRO *P. aeruginosa* infection in Saudi Arabia.

2. Methodology

2.1. Sampling and data collection

A total of 1024 *Pseudomonas aeruginosa* isolates from different clinical specimens; wound infections, blood, respiratory tract infections, body fluid and urine) and a total of 221 MRO *Pseudomonas aeruginosa* were collected from clinical microbiology lab, King Khalid University hospital (KKUH) during the period of 2019 to 2021.^[4] The isolates were cultured on blood agar plates and incubated at 37°C for 24 h.^[1] The bacterial colonies were inoculated in Luria-Bertani (LB) broth and incubated in a shaking rack at 37°C for 24 hours.

2.2. Phenotypic analysis

Phenotypic identification using conventional culture methods, colony characteristics, pigment production, grape-like odor, oxidase positivity, motility, Gram-negative character of the bacilli, and to grow at 42°C. (Memish et al., 2015)

2.3. Antibiotic Susceptibility testing

All isolates were evaluated for susceptibility performed by microscan for the (MROs) in different sites of isolation from 2019-2021. The antibiotic susceptibility testing was done for all the isolates using the automated Vitek®2 system. The antibiotic susceptibility profiles of the isolates were determined using antimicrobial agents, imipenem (IPM) (0.002-32ng/ml),

meropenem (MER)(0.002-32ng/ml), ceftazidime (CAZ) (0.016-256ng/ml), amikacin (AN) (0.016-256ng/ml), tobramycin (TM) (0.016-256ng/ml), ciprofloxacin (CIP) (0.002-32ng/ml), colistin (CS) (0.016-256ng/ml) and aztreonam (ATM) (0.016-256ng/ml). Ceftalozan, ceftazidime-avibactam, (4 mg/liter) by an agar dilution method.^[6] The results were compared to that of standard strain and interpreted as sensitive, intermediate resistant or resistant, based on CLSI guidelines.^[1] (Clinical and Laboratory Standards Institute. 2016)

3. Results

3.1. Phenotypic Analysis

Colonies of **Pseudomonas aeruginosa** appeared as smooth, mucoid, greenish color with beta hemolysis and grape-like smell when grown on blood agar with pigment production, oxidase positive and motile.

3.2. Distribution of P. aeruginosa among different body sites

1024 **Pseudomonas aeruginosa** were isolated from different clinical specimens during the last three year from 2019 to 2021. 221 isolates were multi drug resistance **MROS Pseudomonas aeruginosa**, most of the strains recovered from wound infections (32.4%) while the lowest rate of strains were recovered from respiratory tract infections (9.4%) as shown in Table (1)

Table 1: Total numbers of P. aeruginosa recovered from different clinical specimens.

Clinical specimens	Years			Total
	2019	2020	2021	
Wound	150	95	87	332 (32.4%)
Blood Culture	37	43	16	96 (9.4%)
Respiratory tract	122	99	34	255 (25%)
Body fluid	44	15	21	80 (7.8%)
Urine	22	15	11	48 (4.6%)
Total	457	334	233	1024
Percentage	44.6%	32.6%	22.7%	-

3.3.^[1] Demographic data

The mean age of the patients included in the study was 37.5 years. The higher numbers of the strains were collected from patients above 50 years old, while the lower no of the strains were collected from young patients even less than 1 year old. Out of 1024 isolates of *P. aeruginosa*; 221 strains were found to be multi drug resistant *P. aeruginosa* (MROs). Data suggested that older patients 50 comprised the highest percentage (46%), adult aged 25-50 were (39%), then patients aged 1-25 years were (17%), followed by patient aged less than 1 years were (1.8%) as shown in Figure (1). The higher rate of isolation occur from samples collected from wound (WO) n=332 and the lowest number of the strains were recovered from urine n=48 as shown in Table (2).

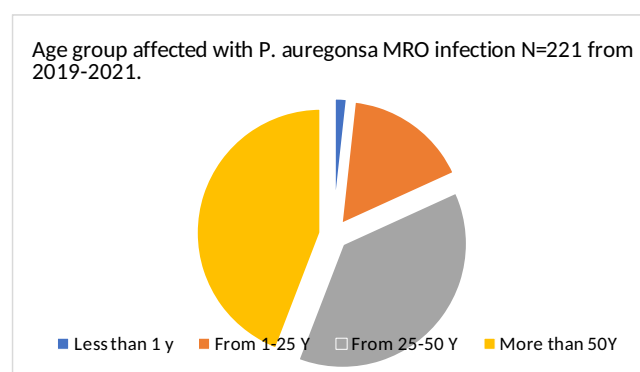


Figure (1): Age distribution of patients suffering from *P. aeruginosa* (MROs) infection during the period of 2019-2021

3.4. *P. aeruginosa* screening by Microscan

The antibiotic susceptibility of all isolates was evaluated by microscan to detect the (MROs) from the strains collected from different clinical specimens during the period of 2019-2021.^[9] **The total number of *P. aeruginosa* (MROs) were N=221. Most MROs *P. aeruginosa* isolates were collected from wound infections (64 isolates, 28.5%), followed by blood culture (10 isolates, 10.4%), then respiratory tract infections (98 isolates, 44.3%), Body fluid (38 isolates, 17%) and urine samples (11 isolates, 5%) as shown in Table (2).**

Table 2: Incidences of P. aeruginosa and multi drug resistance P. aeruginosa (MROs) isolated from different clinical specimens during the period of 2019-2021.

Sites of isolation	Total No of P. aeruginosa	Total No of P. aeruginosa (MRO)	% P. aeruginosa (MRO)
Wound (WC)	332	64	19.2%
Blood (BC)	96	10	10.4%
Respiratory (RR)	255	98	43.5%
Body fluid (BF)	80	38	47.5%
Urine (UR)	48	11	22.9%
Total	1024	221	21.58%

The rate of isolation of P. aeruginosa (MROs) throughout the examination period revealed that the strains recovered from respiratory tract infections were significantly higher (47%) in the strains collected during the period of 2019 than that strains recovered during the period of 2020 and during the period of 2021; 28 (44%) and 21 (21%) respectively. Although, the number of the strains recovered from blood culture were significantly low (5.6%) in 2019, 2 (3%) during the period of 2020, and 2 (3.7%) during the period of 2021, are shown in Table (3) and Figur (2).

Table 3: Incidences of P. aeruginosa (MROs) isolated from different clinical specimens during the period of 2019-2021.

Site of isolation	Urine (UR)		Body Fluid (BF)		Respiratory tract (RR)		Blood culture (BC)		Wound (WC)	
	No	%	No	%	No	%	No	%	No	%
Total "2019" (104)	21	20%	6	5.7%	49	47%	24	23%	4	3.8%
Total" 2020" (63)	23	36.5%	2	3%	28	44%	6	9.5%	4	6.3%
Total "2021" (54)	20	37%	2	3.7%	21	38%	8	14.8%	3	5.5%

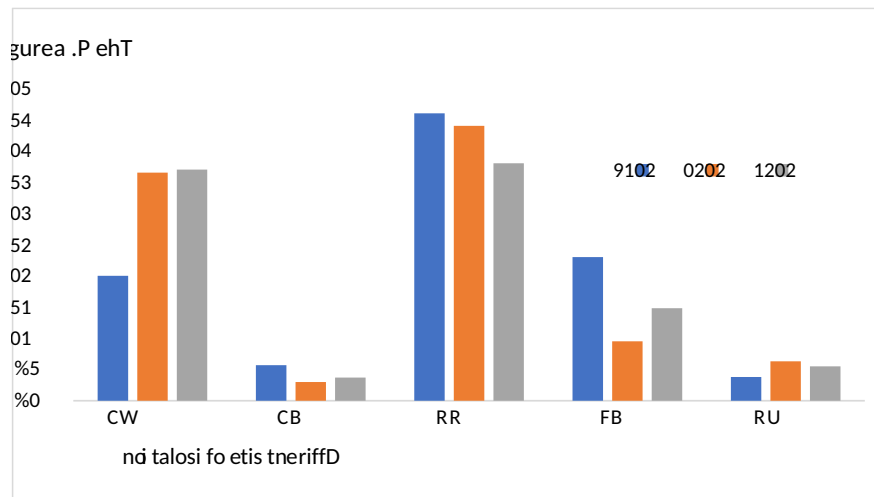


Figure 2: Incidences of *P. aeruginosa* (MROs) isolated from different clinical specimens during the period of 2019-2021.

3.5. Susceptibility testing

The outcomes of the antibiotic sensitivity pattern demonstrated that 77.8% and 61% of the isolates showed resistance against imipenem and meropenem, respectively. Table (4) represents the complete outcomes of the antibiotic resistance pattern for all MROs isolates, the data clearly showed that susceptibility test perform by microscan for all the strains collected from 2019-2021 revealed that (MROs) were resistance against Imipenem (77.8%), Meropenem (61%), Aztreonam (42%) and Ceftazidime (36%) which had been significantly higher than other antibiotics. All isolates sensitive to colistin except (2.7%) of the strains were sensitive, moreover, its of value to said that antibiotic resistant bacteria have been observed with increasing frequency over the past three years shown in Table (4) and Fig (3). Additionally we evaluated the effect of Colistin on 221 clinical isolates of MRO *P. aeruginosa* and it was found that only 6 clinical isolates showed Colistin resistance making 2.7%.

Table (4): Susceptibility perform by microscan for the (MROs) Organisms from 2019-2021

Antibiotic	Years			
	Total Rate of “R” from 2019-2021	2019	2020	2021
Amikacin	15.3%	7.6%	12.6%	33.3%
Gentamicin	20%	13.4%	15.8%	37%
Tobramycin	18%	11.5%	15.8%	33.3%
Piperacillin	30%	16.3%	30%	59.2%
Aztreonam	42%	23%	42.8%	77.7%
Ceftazidime	36%	18.2%	36.5%	70.3%
Cefepime	30%	17.3%	31.7%	53.7%
Ciprofloxacin	11.3%	13.4%	19%	55.5%
Levofloxacin	25.3%	11.5%	20%	57.4%
Imipenem	77.8%	78.8%	74.6%	79.6%
Meropenem	61%	62.5%	68.2%	50%
Colistin	2.7%	2.8%	4.7%	-

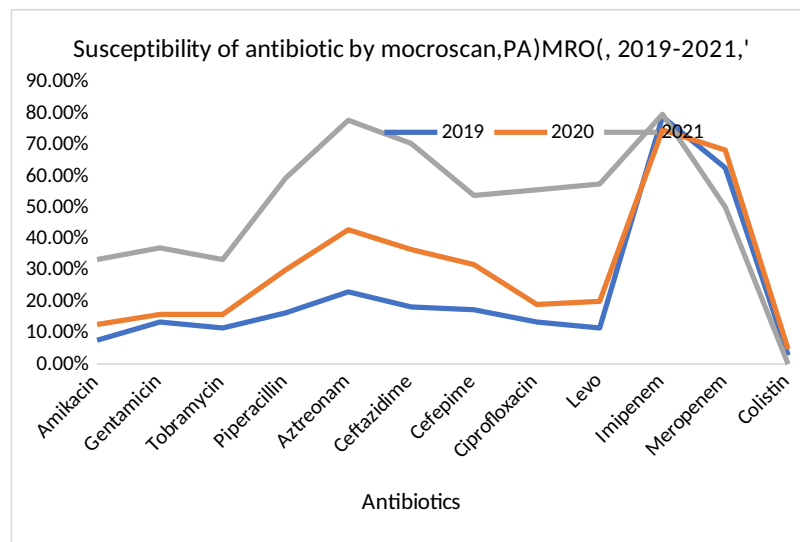


Figure (3): Susceptibility performs by microscan for the (MROs) Organisms from 2019-2021

4. Discussion:

In this study, 1024 isolates of *P. aeruginosa* from clinical samples received from year 2019 - 2021 from KKHU hospital Riyadh, the percentage prevalence of infection with *P. aeruginosa*

seems to be declining yearly with percentage prevalence of 44.6%, 32.6%, and 22.7% in 2019, 2020 and 2021 respectively that is the sign that infection is being controlled from spreading. However, this study is generalized based on specimens from all clinical units of the hospital that is considered as larger study sample. Most isolates in this study were obtained from urine samples, accounting for 51.1% of the total.^[11] Unfortunately, it could not be determined which of the urine samples were catheter specimens, post catheterisation specimens or mid-stream urine samples

In addition, it was also analyzed that the highest percentage of multi drug resistant *P. aeruginosa* strains were associated with Wound (WO) (28.5%), respiratory tract is second most common specimens (44.3%) that showed association with MRO *P. aeruginosa* strains. Whereas, 17% of body fluids, 10.4% of Blood Culture and 5% of Urine isolates showed lowest percentage MRO.^[19] Our findings consent with the findings conducted in Riyadh Saudi Arabia were collected from microbiology lab hospitals in Jazan region during 2017-2018, which documented that most of the MRO *P. aeruginosa* strains were associated with wound (20.2%) and respiratory tract infections (24%) (Kam et al., 2020; Fatima et al., 2019, Hashem et al., 2016, Tamma et al., 2021)

In the present study the most common MRO phenotypes were resistance against carbapenem antibiotics including imipenem & meropenem with 77.8% and 61% respectively. This is much higher than studies conducted by Hashem et al., (2016), who revealed 26.5% to (imipenem & meropenem) resistance in *P. aeruginosa*.

Moreover, In our study pattern of susceptibility demonstrated the resistances from 2019-2021 (MROs) of ceftazidime, cefepime, Amikacin piperacillin and aztreonam and were 36%, 30%, 15.3%, 30% and 42% respectively. Reported 70% resistance to ceftazidime, 75% to piperacillin, 59% to piperacillin/tazobactam, 74% to amikacin, 81% to cefepime, and 69% to aztreonam (Behera et al., 2008, Khan et al., 2016, Nivitha 2016).

In recent years colistin has been used extensively to treat multidrug resistance *P. aeruginosa* infections therefore we also evaluated the effect of Colistin on 221 clinical isolates of MRO *P. aeruginosa*. 6 clinical isolates showed Colistin resistance which is in contrary to results stated in study carried out in Jazan hospitals region during 2017-2018. The potential for encountering

antimicrobial resistance is an important concern for clinicians treating patients with confirmed or suspected *P. aeruginosa* infections. Resistance in *P. aeruginosa* has been shown to lead to increased morbidity and mortality (Carmeli et al., 1999, Harris et al., 1999).

5. Conclusion

MRO *P. aeruginosa* is responsible for serious nosocomial infections among the patients admitted in the hospital, it appears that prevalence of overall infection is decreasing yearly but in contrast, antibiotic susceptibility among *P. aeruginosa* isolates is decreasing in the KSA, perhaps because of increasing or massive use of antibiotics and the lack of adherence to approved infection control practices by hospitals.

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