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- [97]  www.thieme-connect.com/products/ejournals/html/10.1055/a-1487-7642?articleLanguage=en
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1The aptness of entomopathogenic bacteria against sawtoothed grain beetle
2(*Oryzaephilus surinamensis* [L.]) (Coleoptera: Silvanidae) in dates
3

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9Abstract

10Objective

11The sawtoothed grain beetle, *Oryzaephilus surinamensis* (L.)^[8] is known as one of the most
12devastating insect pests in stored dates, infrequently found in newly harvested dates and
13raisins but developed a high plenty in dry fruits particularly after long storage.
14Indiscriminate use of pesticides and fumigants to control insects have resulted in insecticide
15resistance, environmental hazards, residual toxicity, and pest resurgence so the control
16trends have been changed and particularly biological control agents like entomopathogenic
17bacteria *Photorhabdus* temperata and *Xenorhabdus* nematophila being used to manage
18sawtoothed grain beetle.

19Methods

20During present study, the pathogenicity of *P. temperata* and *X. nematophila* was evaluated
21against this crucial pest of stored grains. The bacterial concentrations were applied directly
22to the dates infested with sawtoothed grain beetles and data were recorded.

23Results

24Significant results were observed for both bacterial treatments as the highest concentration
25of 10^8 cells/ml showed to be very effective against the beetle in terms of fecundity, adult
26emergence, and mortality. In terms of fecundity, 8.66 to 4.33^[24] mean number of eggs were

27produced at bacterial *P. temperata* concentrations of 1×10^4 and 1×10^8 cells/ml,
28respectively. Similarly, the F_1 adult emergence for *P. temperata* was recorded 15.33 and 9
29adults at concentrations of 1×10^4 and 1×10^8 cells/ml, respectively. Results showed a similar
30trend for the *X. nematophila* species.

31⁰▶ Conclusions

32These results indicate that utilizing nematodes as biological control agents can be
33advantageous for reducing insect populations in storage facilities and preventing the loss of
34grains and dates that have been stored in storage.

35Keywords: ⁰▶ Stored product insects, microbial control, entomopathogens, date fruits, Saudi
36Arabia

37 Abbreviation

38 First generation (F_1), integrated pest management (IPM), entomopathogenic nematodes
39 (EPNs), toxins complexes (Tc's), gram (g), One-way analysis of variance (ANOVA)

40 1. Introduction

41 Dates contain a high rich protein and good source of rapid energy because of high amount
42 of carbohydrates which is 70-80% (Ghnimi et al., 2017).^[8] The sawtoothed grain beetle is a
43 cosmopolitan pest found globally, it is responsible for sever losses, polyphagous feeding in
44 nature and have broad range of stored goods, as well as cereal products, dried fruits, oil
45 seeds and stored grains (Hashem et al., 2012). The sawtoothed grain beetle, infest dates
46 which has low moisture content, with the calyx removed or those broken or with
47 mechanical damage. Tunnels are made on the outer fruit skin and flesh by feeding of adults
48 and larvae. During the high infestation, sawtoothed grain beetle consume all fruit contents
49 leaving the skin or exo-carp integral. The beetle causes severe damage such as weight
50 decrease, reducing the quality and quantity of the dates. Stored dry and semi dry dates face
51 a basic issue of insect damage (Al-Dosary 2009). In stored products the date varieties are
52 most infected by sawtoothed grain beetle and 40 to 75% losses have been reported (Mallah
53 et al., 2016).

54^[0] Pesticides have been used as protectants all over the world because of their efficient pest
55 control ability but on the other hand, the detrimental effects may include toxicity of stored
56 food commodities causing sanitary and phytosanitary issues as well as life threatening
57 effects on non-targeted organisms (Phillips and Throne 2010).^[0] For the sake of human health
58 and safety, environment friendly the implementation of less expensive control measures has
59 become the need due to the lack of awareness of pesticides cost, their detrimental effects on
60 human health and development of genetic resistance in insect species against these
61 chemicals (Aggarwal et al., 2016).^[9] Many countries are scrutinizing the traditional

62 fumigation procedures as they are posing threats which include ozone depletion, potential
63 risk of carcinogenic methyl Bromide and phosphine.^[0] Alternative strategies must focus on
64 the efficacy against limited target specific species which are biodegradable into non-toxic
65 products and recommended to be use in integrated pest management programs as well as
66 eco-friendly measures development are in discussion. Plant-based chemicals, insect growth
67 regulators and the insect pathogens like entomopathogenic bacteria, fungi, viruses and
68 protozoan are being assessed as better alternatives to chemical based prepared insect
69 control programs.^[0] Biological control has great attention over past few decades, as
70 alternative to chemical insecticides or as a constituent of integrated pest management (IPM)
71 (Subramanyam and Hagstrum 2012). These constitute effective alternatives to chemicals
72 without producing adverse effects on the environment.

73^[5] Entomopathogenic bacteria, *Photorhabdus temperata* and *Xenorhabdus nematophila* are
74 gram negative, motile rod-shaped bacteria which belongs to family Enterobacteriaceae and
75 form symbiosis with entomopathogenic nematodes (EPNs) *Steinernema* and
76 *Heterorhabdus* (Akhurst and Boemare 1988; Akhurst et al., 1996). Due to this symbiosis,
77 nematodes provide shelter to bacteria in their guts where they seek protection from soil
78 stressors as well as antagonists such as telluric bacterial consortia and bacteria inside insect
79 guts that is why the isolation from soil samples is not possible without their nematode host.
80 This pathogenic symbiosis is able to parasitize as well as to kill the larval stages of host
81 from orders Diptera, Hymenoptera, Lepidoptera, Orthoptera, Coleoptera and Isoptera
82 (Boemare 2002; Belien 2018). The toxins complexes (Tc's) and *Photorhabdus* insect-
83 related toxins show oral activity against insect species despite of the fact that oral infection
84 is not important to the biology of *Photorhabdus* or *Xenorhabdus*. Gram-positive and gram-
85 negative bacteria produced toxins which have high molecular weight and multi subunit
86 with insecticidal properties (Waterfield et al., 2001). Bacteria *Photorhabdus* or

87Xenorhabdus encodes the toxins producing genes which co-exist with entomopathogenic
88nematodes. Strains of bacteria have been discovered with loci that encodes for toxins, some
89of which are associated to insect while others are not. The objective of this research is
90assessment of entomopathogenic bacterial toxicity towards sawtoothed grain beetle
91management and determining mortality of sawtoothed grain beetle in dates due to bacteria.

92. Materials and methods

932.1 Insects rearing

94 *Oryzaephilus surinamensis* adults were reared on dates and its healthy cultures were
95 maintained in plastic jars, covered with the muslin cloth; tightened with the elastic rubber
96 band.^[0] These jars were placed in a SANYO incubator, which was set at $27 \pm 2^\circ\text{C}$ and 70 ± 5
97 percent relative humidity in the laboratory. Adult of sawtoothed grain beetle was identified
98 as per the males have spine like structure on femur of meta-leg, which is absent in females.

992.2 Bacterial culture maintenance

100 Entomopathogenic bacteria namely *Photorhabdus temperata* and *Xenorhabdus*
101 *nematophila* were obtained from Korean Agriculture Collection. At first the culture on
102 supplement agar plates was streaked over at 25°C for 2 to 4 days. Obtained culture was
103 filtered by streaking individual colony on nutrient agar. Refined bacterial culture was
104 replicated in nutrient broth for 24 hours. The serial dilution plate count method was used to
105 draw a dilution curvature between the optical thickness and cells/ml in order to calculate
106 and adjust the colony shaping units per unit volume. Distinctive concentrations of bacteria
107 were prepared to apply against sawtoothed grain beetle.

1082.3 Bacterial pathogenicity Bioassay

109 In each jar, 50g of dates was placed. The jars were secured with muslin cloth, fixed with
110 elastic band and were placed in an incubator at temperature and humidity mentioned
111 earlier.^[0] Following different concentrations, 1×10^4 , 1×10^5 , 1×10^6 , 1×10^7 and 1×10^8 cells/ml
112 of both bacteria were prepared and used for the experiments. In the control treatment
113 distilled water was used. There were three replicates for each treatment, each jar
114 representing one replicate, and there were five pairs of the young beetles in each jar. The
115 effectiveness of entomopathogenic bacteria against sawtoothed grain beetle were studied
116 according to the following parameters.

117 2.4 Eggs number

118 Eggs were counted on the third day following the bacterial post treatment. Three randomly
119 selected dates were used to open each jar and count the number of eggs within. Average
120 number of eggs were determined influence of different treatment on egg laying capacity
121 (fecundity). Because the sawtoothed grain beetle's eggs are so tiny and delicate, therefore,
122 only three dates from each jar were opened, and the eggs were counted in order to prevent
123 any damage that would have an impact on the outcomes, such adult emergence.

124 2.5 Sawtoothed grain beetle adult emergence (F_1) and mortality%

125 After four weeks of the bioassay, all the jars were opened, emergence of adults (F_1) and the
126 mortality of former adults employed for the bioassay and the new adults were counted. The
127 number of adults' (F_1) were calculated in individual jar to check the repression of *O.*
128 *surinamensis* appearance by using diverse concentrations of entomopathogenic bacteria. ^[0] In
129 a similar manner, the mortality was assessed at 7, 14, and 21 days following treatment. ^[0] At
130 each observation, the dead adults were removed from the jars, the number of dead and live
131 adults were counted, and the mortality percentage was computed.

132 2.6 Statistical analysis

133 The data were collected, tabulated and analyzed. One-way analysis of variance (ANOVA)
134 was done using bacterial concentrations as the main factor and eggs laid, adult mortality
135 and new adult emergence as the response factor. The SAS 9.2 was used to analyze the data.

1363. Results

1373.1 Number of eggs laid by sawtoothed grain beetle in stored dates treated with various
138 concentrations of *P. Temperata*

139 According to Table 1, all used concentrations of *P. temperata* Entomopathogenic Bacteria
140 viz., 1×10^4 , 1×10^5 , 1×10^6 , 1×10^7 and 1×10^8 cells/ml significantly reduced sawtoothed grain
141 beetle fecundity as compared to the control. The least number of eggs were counted in the
142 dates treated with the bacterial concentrations of 1×10^7 and 1×10^8 cells/ml in which the
143 values were 5.00 and 4.00 respectively. However, the concentration 1×10^5 , 1×10^6 , 1×10^7
144 and 1×10^8 cells/ml were statistically similar to each other while 1×10^4 cells/ml
145 concentration was dissimilar statistically from the all other used concentration. All bacterial
146 concentration proved pathogenic against sawtoothed grain beetle fecundity biological trait
147 and showed good results than the control.^[0] Moreover, the results showed that the number of
148 eggs laid by sawtoothed grain beetles got decreased with the increase in the
149 entomopathogenic bacterial concentrations.

1503.2 Sawtoothed grain beetle adult (F₁) emerged in stored dates treated with different
151 concentrations of *P. temperata*

152 All the bacterial concentrations of *P. temperata* 1×10^4 , 1×10^5 , 1×10^6 , 1×10^7 and 1×10^8 cells/
153 ml were better statistically in the F₁ adult emergence (table 2). The minimum number of
154 9.00 F₁ adults were emerged in the concentration of 1×10^8 cells/ml while 1×10^4 bacterial
155 concentration showed maximum number of the 15.00 emerged adults. Moreover, the
156 concentration 1×10^5 , 1×10^6 and 1×10^7 cells/ml were showed statistically different results
157 with one another. All bacterial concentrations indicated better F₁ adults inhibiting rate in
158 stored dates as compared to the control.

1593.3 Sawtoothed grain beetle adult (F1) emerged in stored dates treated with different
160concentrations of *X. nematophila*

161Table 3, shows that highest numbers of sawtoothed grain beetle eggs 7.00 were observed
162when the date were treated with the bacterial concentration of 1×10^4 cells/ml. Similarly, the
163lowest number of the eggs was shown by the bacterial concentration 1×10^7 and 1×10^8 cells/
164ml. All concentration gives a better result as compared to the control. The concentration
165 1×10^4 , 1×10^5 and 1×10^6 were not statistically dissimilar while the 1×10^7 and 1×10^8 were
166similar among themselves and significantly different from the all other concentrations.
167Along with increase in the bacterial concentrations the number of eggs laid was reduced.

1683.4 Number of adult F1 emerged sawtoothed grain beetle in stored dates treated with
169various concentrations of *X. nematophila*

170The bacteria *X. nematophila*, showed significant inhibition of sawtoothed adult emergence.
171According to the Table 4, most effective concentrations which results in minimum number
172of new adults produced was observed to the 1×10^8 cells/ml while the least effective
173concentration was 1×10^4 cells/ml producing the greatest number of new emerged adults of
174sawtoothed grain beetle. However, all concentrations showed a smaller number of the
175adults as compared to the control. It was observed that when the higher concentration was
176used then a smaller number of sawtoothed grain beetle adults emerged.

1773.5 Sawtoothed grain beetle adult mortality in stored dates treated with various
178concentrations of *X. nematophila*

179Adult mortality was calculated in percentage by counting the dead and live adults in each
180jar at various time interval of the post treatment. The bacterial content and exposure time
181had a clear correlation with the mortality. The maximum adult mortality with 1×10^8 *X.*
182*nematophila* cells/ml concentrations after the seven days of post treatment was 30%. On the

183other hand, the same treatment resulted in a maximum mortality rate of up to 70% at day 21
184of post treatment (table 5).

1854. Discussion

186 Dates are an extremely valuable fruit because they are only harvested once in a year with
187 proper storage and insect prevention, may be eaten for several years. The usage of
188 entomopathogenic organisms is very safe for people and other animals, including cattle that
189 eat dates incorporated into their diets as an essential nutrient. The ability of
190 entomopathogenic bacteria to inhibit the sawtoothed grain beetle from laying eggs has been
191 proven by current data. *Xenorhabdus* spp., a symbiotic bacterium, is one of the well-known
192 biocontrol agents used in pest management (Zhou et al., 2002; Gulcu et al., 2012).
193 According to (Richards and Goodrich-Blair 2010), *X. nematophila* is an entomopathogenic
194 bacterium with a wide host range and strong toxicity against insect pests; nonetheless,
195 similar results were seen in our investigation. Similar findings were reported that *P.*
196 *temperata* and *X. nematophila* were reported as deadly pathogens that produced proteins
197 and secondary metabolites which prove toxic against a variety of insects (Dowling and
198 Waterfield 2007; Sheets et al., 2011; Ng'ang'a 2015; Namsena et al., 2016). In particular,
199 discussing *X. nematophila*'s pathogenicity against adults of various insects has
200 demonstrated that *Drosophila melanogaster* and *Manduca sexta* adults quickly died after
201 being infected with *X. nematophila* (Kim et al., 2017).^[29] These bacteria produce a number of
202 compounds that are insecticidal and can be utilized to combat a variety of species,
203 including insects (Abd-Elgawad 2022; Tomar et al., 2022).

204 In present study the emergence of F1 adults was very low in the treated dates as compared
205 with the saw-toothed grain beetle adults where distilled water was applied as control
206 treatment. It was noticed during observation that adults were not active in the treated dates
207 as compared to the dates where water was applied. When adults were not active it reflects
208 the deterrent properties of the metabolites produced by the bacteria. This deterrent property
209 of bacteria has affected the normal activities like feeding, copulation, and fecundity. The

210present data has shown the evidence of low fecundity and F1 adult emergence in the treated
211dates which have also been reported in other studies (Bode 2009; Kusakabe et al., 2022).
212Keeping continue, there are several natural products synthesized from the toxic produced
213by several species of Photorhabdus and Xenorhabdus, secretions and being used
214commercially for the management of several issues regarding human health and food
215storage and protection (Cimen et al., 2022). All these findings are in favor of the present
216results outcomes.

217Conclusions

218Present study results showed that bacterial effectiveness was strongly correlated with
219exposure period; larger concentrations of *X. nematophila* were more effective than lower
220concentrations against *O. surinamensis*. An investigation revealed that the
221entomopathogenic bacteria *X. nematophila* was more aggressive against *O. surinamensis*
222when applied to the dates infested with it. The study's findings led to a successful and
223secure biological control procedure and will in the future direct an effective IPM program
224for this crucial pest.

225Acknowledgements

226Author would like to thanks the National Center for Agricultural Technology (NCAT),
227King Abdulaziz City for Science and Technology (KACST), Riyadh, Saudi Arabia.

228Declaration of Competing Interest

229The author declare that he has no known competing financial interests or personal
230relationships that could have appeared to influence the work reported in this paper.

231Disclosure of funding

232The authors declare that they don't have any particular funding for this study.

233References

- 234Abd-Elgawad, M.M., 2022. Xenorhabdus spp.: An overview of the useful facets of
235 mutualistic bacteria of entomopathogenic nematodes. Life. 12 (9) 1360.
- 236Aggarwal, N., Thind, S., and Sharma, S., 2016. Role of secondary metabolites of
237 actinomycetes in crop protection. Plant Growth Promoting Actinobacteria, Springer:
238 99-121.
- 239Akhurst, R., and Boemare, N., 1988. A numerical taxonomic study of the genus
240 Xenorhabdus (Enterobacteriaceae) and proposed elevation of the subspecies of X.
241 nematophilus to species. Microbiology. 134 (7) 1835-1845. DOI:[M10.1099/00221287-
242 134-7-1835](https://doi.org/10.1099/00221287-134-7-1835)
- 243Akhurst, R.J., Mourant, R.G., Baud, L., and Boemare, N., 1996. Phenotypic and DNA
244 relatedness between nematode symbionts and clinical strains of the genus
245 Photorhabdus (Enterobacteriaceae). IJSM. 46 (4) 1034-1041.
246 DOI:[M10.1099/00207713-46-4-1034](https://doi.org/10.1099/00207713-46-4-1034)
- 247Al-Dosary, N., 2009. Role of the Saw-toothed grain beetle (*Oryzaephilus surinamensis* L.)
248 (Coleoptera: Silvanidae) in date palm fruit decay at different temperatures. Basra J.
249 Vet. Res. 8 (2) 1-14.
250
- 251Belien, T., 2018. Entomopathogenic nematodes as biocontrol agents of insect pests in
252 orchards. CAB Reviews. 13 (058) 1-11. DOI: [10.1079/pavsnnr201813058](https://doi.org/10.1079/pavsnnr201813058)
253
- 254Bode, H.B., 2009. Entomopathogenic bacteria as a source of secondary metabolites. Curr
255 Opin Chem Biol. 13 (2) 224-230. DOI: [10.1016/j.cbpa.2009.02.037](https://doi.org/10.1016/j.cbpa.2009.02.037)
256
- 257Boemare, N., 2002. Biology, taxonomy and systematics of *Photorhabdus* and
258 *Xenorhabdus*. J. Entomol. Nematol. 35-56. DOI: [10.1079/9780851995670.0035](https://doi.org/10.1079/9780851995670.0035)
259
- 260Cimen, H., M. Touray, S. H. Gulsen, and Selcuk, H., 2022. Natural products from
261 *Photorhabdus* and *Xenorhabdus*: Mechanisms and impacts. Appl. Microbiol.
262 Biotechnol. 1-13. DOI: [10.1007/s00253-022-12023-9](https://doi.org/10.1007/s00253-022-12023-9)
263
- 264Dowling, A., and Waterfield, N.R., 2007. Insecticidal toxins from *Photorhabdus* bacteria
265 and their potential use in agriculture. Toxicon. 49 (4) 436-451. DOI:
266 [10.1016/J.TOXICON.2006.11.019](https://doi.org/10.1016/J.TOXICON.2006.11.019)
267
- 268Ghnimi, S., Umer, S., Karim, A., and Afaf, K., 2017. Date fruit (*Phoenix dactylifera* L.):
269 An underutilized food seeking industrial valorization. NFS journal. 6 1-10. DOI:
270 [10.1016/J.NFS.2016.12.001](https://doi.org/10.1016/J.NFS.2016.12.001)
271
- 272Gulcu, B., Hazir, S., and Kaya, H.K., 2012. Scavenger deterrent factor (SDF) from
273 symbiotic bacteria of entomopathogenic nematodes. J. Invertebr. Pathol. 110 (3) 326-
274 333. DOI: [10.1016/j.jip.2012.03.014](https://doi.org/10.1016/j.jip.2012.03.014)
275
- 276Hashem, M.Y., Ahmed, S.S., El-Mohandes, M.A., and Gharib, M.A., 2012. Susceptibility
277 of different life stages of saw-toothed grain beetle *Oryzaephilus surinamensis* (L.)

278 (Coleoptera: Silvanidae) to modified atmospheres enriched with carbon dioxide. J
279 Stored Prod Res. 48 46-51. DOI: [10.1016/J.JSPR.2011.09.002](https://doi.org/10.1016/J.JSPR.2011.09.002)
280

281Kim, I.H., Aryal, S.K., Aghai, D.T., Casanova-Torres, Á.M., Hillman, K., Kozuch, M.P.,
282 Mans, E.J., Mauer, T.J., Jean-Claude, O., and Ensign, J.C., 2017. The insect
283 pathogenic bacterium *Xenorhabdus innexi* has attenuated virulence in multiple insect
284 model hosts yet encodes a potent mosquitocidal toxin. BMC genomics. 18 (1) 1-25.
285 DOI: [10.1186/s12864-017-4311-4](https://doi.org/10.1186/s12864-017-4311-4)
286

287Kusakabe, A., Wang, C., Xu, Y.-m., Molnár, I., and Stock, S.P., 2022. Selective Toxicity of
288 Secondary Metabolites from the Entomopathogenic Bacterium *Photorhabdus*
289 *luminescens sonorensis* against Selected Plant Parasitic Nematodes of the Tylenchina
290 Suborder. Microbiol. Spectr. 10 (1) e02577-02521. DOI: [10.1128/spectrum.02577-21](https://doi.org/10.1128/spectrum.02577-21)
291

292Mallah, N.A., Sahito, H.A., Kousar, T., Kubar, W.A., Jatoi, F.A., and Shah, Z.H., 2016.
293 Susceptibility of different varieties of stored date palm fruits infested by saw tooth
294 grain beetle, *Oryzaephilus surinamensis* (L., 1758) under laboratory conditions. J.
295 Entomol. Zool. Stud. 4 438-443.
296

297Namsena, P., Bussaman P., and Rattanasena, P., 2016. Bioformulation of *Xenorhabdus*
298 *stockiae* PB09 for controlling mushroom mite, *Luciaphorus perniciosus* Rack. BIOB.
299 3 (1) 1-7. DOI: [10.1186/s40643-016-0097-5](https://doi.org/10.1186/s40643-016-0097-5)
300

301Ng'ang'a, P., 2015. In Silico and Molecular Analysis of Xpta Protein Toxin Genes of
302 *Xenorhabdus* Sp. and Efficacy of the Bacterium on *Sitophilus Zeamais* and
303 *Prostephanus Truncatus*, University of Nairobi.
304

305Phillips, T.W., and Throne, J. E., 2010. Biorational approaches to managing stored-product
306 insects. Annu. Rev. Entomol. 55 375-397. DOI:
307 [10.1146/annurev.ento.54.110807.090451](https://doi.org/10.1146/annurev.ento.54.110807.090451)
308

309Richards, G. R., and Goodrich-Blair, H., 2010. Examination of *Xenorhabdus nematophila*
310 lipases in pathogenic and mutualistic host interactions reveals a role for *xlpA* in
311 nematode progeny production. Appl. Environ. Microbiol. 76 (1) 221-229. DOI:
312 [10.1128/AEM.01715-09](https://doi.org/10.1128/AEM.01715-09)
313

314Sheets, J. J., Hey, T. D., Fencil, K. J., Burton, S.L., Ni, W., Lang, A.E., Benz, R., and
315 Aktories, K., 2011. Insecticidal toxin complex proteins from *Xenorhabdus*
316 *nematophilus*: structure and pore formation. J. Biol. Chem. 286 (26) 22742-22749.
317

318Subramanyam, B., and Hagstrum, D.W., 2012. Alternatives to pesticides in stored-product
319 IPM, Springer Science & Business Media.
320

321Tomar, P., Thakur, N., and Yadav, A.N., 2022. Endosymbiotic microbes from
322 entomopathogenic nematode (EPNs) and their applications as biocontrol agents for
323 agro-environmental sustainability. Egypt J Biol Pest Control. 32 (1) 1-19. DOI:
324 [10.1186/s41938-022-00579-7](https://doi.org/10.1186/s41938-022-00579-7)
325

326 Waterfield, N.R., Bowen, D.J., Fetherston, J.D., and Perry, R.D., 2001. The tc genes of
327 Photorhabdus: a growing family. Trends Microbiol. 9 (4) 185-191. DOI:
328 [10.1016/S0966-842X\(01\)01978-3](https://doi.org/10.1016/S0966-842X(01)01978-3)
329
330 Zhou, X., Kaya, H.K., Heungens, K., and Goodrich-Blair, H., 2002. Response of ants to a
331 deterrent factor (s) produced by the symbiotic bacteria of entomopathogenic
332 nematodes. Appl. Environ. Microbiol. 68 (12) 6202-6209. DOI:
333 [10.1128/AEM.68.12.6202-6209.2002](https://doi.org/10.1128/AEM.68.12.6202-6209.2002)