



Original article

Polymorphisms of TP^{53} gene and its association with colorectal cancer: A case-control investigationAbdullah M Alhadheq^{a,*}, Narasimha Reddy Parine^{b,c}, Jilani Purusottapatnam Shaik^b, Rana Alhadheq^d, Saad Alkahtani^a, Nada H. Aljarba^e, Mohammad Alanazi^b^a Department of Zoology, College of Science, King Saud University, P. O. Box 2455, Riyadh 11451, Saudi Arabia^b Genome Research Chair, Department of Biochemistry, College of Science, King Saud University, Riyadh, Saudi Arabia^c Director, Kakatiya Degree and PG College, Nalgonda, India^d Department of Medicine, Medical School, Georgian American University, Tbilisi, Georgia^e Department of Biology, College of Science, Princess Nourah bint Abdulrahman University, P. O. Box 84428, Riyadh 11671, Saudi Arabia

ARTICLE INFO

Keywords:

 TP^{53}

Genotyping

Cancer

Colorectal

SNP

Polymorphism

ABSTRACT

The tumor suppressor gene (TP^{53}) is crucial for DNA repair mechanism, apoptosis, and cell cycle regulation and progression. In human cancer, TP^{53} is mutated and highly polymorphic. In the current case-control research investigation, we investigated TP^{53} gene SNPs, in exonic and intronic regions, as potential risk factors for colorectal cancer (CRC). This study comprised of 192 patients and 192 control. Obtained data illustrated that only the G allele; rs1042522 (Pro72Arg (C > G), demonstrated a statistically significant association, almost 1.5-fold induction promotes the risk of CRC development in contrast to individuals with the C allele (OR = 1.5, $\chi^2 = 7.28$, $p = 0.00696$). The homozygous variant GG genotype of rs1042522 was also a significant risk factor to CRC development (OR = 2.1, $\chi^2 = 6.41$, $p = 0.01136$). SNP rs1042522 polymorphism established a considerably elevated odds of CRC among male patients aged < 57 years and in patients' with tumors situated in colon region. *In silico* analysis exhibited that proline to arginine amino acid substitution affects the protein structure. Both rs1642785 and rs9894946 SNPs did not demonstrate any significant statistical association with CRC. In conclusion, this study confirmed that rs1042522 SNP within TP^{53} gene is correlated with possibility of developing CRC in the Saudi population. This finding highlights those polymorphisms within TP^{53} gene could act as a diagnostic indicator for CRC.

1. Introduction

Colorectal cancer (CRC) is deemed among major malignancies worldwide. It occurs as a result of accumulation of set of genetic and epigenetic modifications over time in different pathways that are proven to drive and transform the colonic epithelial cells into tumors (Houlston and Tomlinson 2001; Sung et al., 2021). It may take 10 to 15 years for the development and progression of carcinogenesis, which involves concurrent histological and nuclear changes (Fearon and Vogelstein 1990; Katerji and Duerksen-Hughes, 2021). Generally, variations in several biochemical pathways play vital roles in the development and transition from adenoma to carcinoma (Jesionek-Kupnicka et al., 2017). In colon cancer, approximately 90 % of the cases are reported to have mutations in APC and in TP^{53} pathways (Michor et al., 2005; Kanth and

Inadomi, 2021; Vuik et al., 2019; Chittleborough et al., 2020). In this regard, the role of TP^{53} gene in suppression of tumor cannot be undermined. The human TP^{53} gene is positioned on chromosome 17 which has 11 exons and 10 introns (Lamb and Crawford 1986), and encodes TP^{53} protein. In over 50 % of human cancers, TP^{53} undergoes mutation, while the remaining cases exhibit changes in its regulators or targets (Hu et al., 2021). In 1979, the TP^{53} protein was initially identified as an oncogene by different groups (Hernandez Borrero and El-Deiry 2021). Simultaneously, it was found to complex with the SV40 virus T antigen in cells undergoing 8–11 tumor transformation. Subsequently, other studies have identified interactions between this 53 kDa protein and adenovirus and human papillomavirus proteins (Werness et al., 1990). The “tumor antigen” is upregulated in tumor cells and seems to collaborate with other oncogenes like HRAS in converting primary cells into cultured

Peer review under responsibility of King Saud University.

* Corresponding author.

E-mail address: 437106985@ksu.edu.sa (A.M. Alhadheq).<https://doi.org/10.1016/j.jksus.2023.102936>

Received 18 July 2023; Received in revised form 1 September 2023; Accepted 6 October 2023

Available online 12 October 2023

1018-3647/© 2023 Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

cells (Miret et al., 2003). TP^{53} plays a crucial part in various biological processes, including DNA repair pathways, cell cycle regulation, and apoptosis (Levine and Oren 2009). TP^{53} is often altered in various cancers and it is a polymorphic gene. Many studies showed an enormous number of single nucleotide polymorphisms (SNPs) in the exon, promoter, and intron regions of the TP^{53} gene. Some of these studies showed association between SNPs in TP^{53} and cancer (Whibley et al., 2009, Jesionek-Kupnicka et al., 2017, Sobieszko et al., 2017). This gene is critical to tumor progression and therapeutic response. Therefore, the current study was intended to study the potential association of SNPs in TP^{53} gene with CRC in the Saudi population. We analyzed the association of TP^{53} (rs1042522, rs1642785, and rs9894946) polymorphism with CRC development among Saudi population.

2. Material and methods

2.1. Patient samples

Samples from Saudi Arabia of CRC patients (n = 192) and matched controls (n = 192) were obtained from collaborators and clinicians according to the guidelines of 12/3352/IRB. Patients visiting the Endoscopy Department at King Khalid University Hospital (KKUH) were examined by the oncologist alongside a routine examination. There were no restrictions in patient group in terms of age and CRC stages. For genotyping studies, each patient donated blood volume of 5 ml. Clinical and demographic traits such as tumor location, sex, age, and ethnicity, family history of cancer, lymph node status, and smoking habit were recorded for all the study participants (both cases and controls). Informed consent statements were collected from all the study participants in accordance with the rules of the ethical review committee at King Saud Medical City, King Saud University.

2.2. Nucleic acid Isolation

Blood samples were utilized in the extraction process for Genomic DNA, by QIAamp DNA Blood Mini Kit (Qiagen, Valencia, CA, USA) as per manufacturer's guide. Briefly, 20 μ l protease was inserted in 200 μ l of blood samples in 1.5 ml tubes and mixed. Next, AL buffer was incubated for 10 min at temperature of 56 °C. Mixed was then centrifugated and then transferred to spin columns. Wash buffers 1 & 2 were inserted to the column, and centrifugated after each wash. The elution was performed in 50 μ l AE buffer. The yielded DNA was measured for volume and purity using the NanoDrop8000 spectrophotometer (Thermo Scientific).

2.3. Genotyping

Both CRC and normal DNA samples were genotyped and amplified for TP^{53} SNPs by real-time polymerase chain reaction (PCR) using a TaqMan SNP genotyping assays as described (Alanazi et al., 2013, Angelopoulou et al., 2017, Ozdemirkiran et al., 2017). Each well containing 20 ng of genomic DNA, 5.6 μ l of TaqMan® Genotyping Master Mix (Applied Biosystems, Foster City, CA, USA), 0.2 μ l of 40 \times TaqMan® Genotyping SNP Assay (Applied Biosystems). QuantStudio™ Real-Time PCR (Applied Biosystems) was used for each genotyping run with an endpoint reaction reading. The setting of the PCR run are as follows; 1) pre-read stage for 30 s at 60 °C temperature, 2) hold stage 10 min at 95 °C, 3) 40 cycles at PCR stage 15 s for denaturation at 95 °C and annealing for 1 min at 60 °C, and 4) post-read stage 30 s at 60 °C.

2.4. Statistical analysis

The data analysis was conducted by calculating the allele frequencies. Genotype evaluated the differences between the samples, and the calculation was executed as per Pearson's goodness-of-fit chi-square. The allelic variations were calculated on the basis of wild type which

was treated as a reference for the present investigation. Chi-square, odds ratios, p-values, and confidence intervals were computed by using IBM SPSS version 23. Haploview software was used to plot the Linkage disequilibrium (Barrett et al., 2004). In silico study done by using the online tool (<https://www3.cmbi.umcn.nl/hope/input>).

3. Results

The present study included 192 patients' samples diagnosed with CRC and 192 CRC patients, with earlier assent from every person. Clinical and demographic details are presented in (Table 1). Genetic polymorphisms that were identified, i.e., rs1042522 (Pro72Arg (C > G)) from the exonic region, rs1642785 (C > G) in the intronic region, and rs9894946 (A > G) in the intronic region, in TP^{53} gene variants, were examined in Saudi cohort diagnosed with CRC, to assess the risk of susceptibility to develop CRC. The genotype distributions are demonstrated in Table 2. The genotype distributions for all the SNPs were in agreement with Hardy-Weinberg Equilibrium (HWE). In the overall analysis, a significant link was observed only with the G allele of rs1042522 (Pro72Arg (C > G)), which showed a nearly 1.5-fold increase in odds of CRC development in comparison to the individuals with the C allele (OR = 1.5, $\chi^2 = 7.28$) (Table 1). The p-value for this association was 0.00696, which is statistically significant at the 0.05 level. Moreover, the homozygous variant GG genotype of rs1042522 also had a statistically significant association with risk of CRC (OR = 2.1, $\chi^2 = 6.41$, p = 0.01136). This suggest that individuals with the GG genotype were 2.1 times more likely to develop CRC than individuals with the CC genotype.

The genotype frequencies of rs1642785 and rs9894946 did not show statistically significant associations in an overall comparison between CRC cases and controls (Table 2). Furthermore, we divided the samples into two subgroups stratified based on the median age of patients, i.e., below or above 57 years to study the influence of TP^{53} SNPs rs1042522, rs1642785 and rs9894946 on risk of CRC. As detailed in Table 3, a notable correlation was observed in the case of the rs1042522 allele among individuals with CRC who were younger than 57 years old (Specifically, the CG genotype at p = 0.008, the GG genotype at p = 0.014, and the presence of the G allele at p = 0.001). However, this significant association did not evident in the older patient group. Both SNPs rs1642785 and rs9894946 did not display any association on age stratification among patients. We also studied the effect of TP^{53} SNPs rs1042522, rs1642785, and rs9894946 on risk of CRC development in subgroups stratified by sex (male and female). As shown in Table 4, only rs1042522 polymorphism showed significant risk for developing CRC among male patients (Specifically, the GG genotype at p = 0.041, and the presence of the G allele at p = 0.017); SNPs rs1642785 and rs9894946 not showing any association with CRC among patients stratified by the male gender. Among female, none of these SNPs confirmed statistical significance (Table 4).

The analysis extended to consider the location of tumors within the colorectal region. Among the investigated variants, rs1042522 exhibited a significantly higher risk of developing CRC in the colonic region for individuals with the homozygous GG allele (χ^2 : 7.00; CI: 1.24–4.66; p =

Table 1
Clinical and demographic characteristics of CRC patients and controls.

Clinical characteristics		Cases	Controls
Age	Median age, years	57	58
	Range, years	23–79	21–76
	≤57 years	91	114
	>57 years	101	78
Gender	Male	116	108
	Female	76	80
Tumor location	Colon	109	–
	Rectum	70	–
Family history of cancer		32	–

Table 2
Genotype frequencies of *TP53* gene polymorphism in colorectal cancer cases and controls.

SNP	Variant	Cases (Freq)	Controls	OR	P- Value
rs1042522	CC	0.40	0.49	Ref	
	CG	0.39	0.37	1.28	0.27
	GG	0.22	0.13	2.10	0.011
	C	226 (0.59)	0.68	Ref	
	G	158 (0.41)	0.31	1.50	0.007
rs1642785	CC	0.15	0.09	Ref	
	CG	0.43	0.49	0.52	0.057
	GG	0.42	0.42	0.61	0.16
	C	0.36	0.34	Ref	
	G	0.64	0.66	0.89	0.46
rs9894946	AA	0.44	0.52	Ref	
	AG	0.40	0.37	1.28	0.27
	GG	0.16	0.11	1.66	0.11
	A	0.64	0.70	Ref	
	G	0.36	0.30	1.33	0.065

Table 3
Genotype frequencies of *TP53* gene polymorphism in colorectal cancer cases and controls based on median age of patients.

SNP	Variant	Cases (Freq)	Controls (Freq)	OR	P- Value	
rs1042522 (age: below 57)	CC	0.36	0.57	Ref		
	CG	0.46	0.32	2.27	0.008	
	GG	0.18	0.09	2.90	0.014	
	C	0.59	0.74	Ref		
	G	0.41	0.25	1.96	0.001	
	(age: above 57)	CC	0.43	0.37	Ref	
		CG	0.32	0.44	0.62	0.157
		GG	0.26	0.17	1.25	0.582
		C	0.58	0.59	Ref	
		G	0.42	0.40	1.05	0.819
rs1642785 (age: below 57)	CC	0.16	0.07	Ref		
	CG	0.37	0.46	0.36	0.036	
	GG	0.48	0.47	0.45	0.101	
	C	0.34	0.30	Ref		
	G	0.66	0.70	0.83	0.380	
	(age: above 57)	CC	10.14	0.12	Ref	
		CG	0.49	0.55	0.73	0.512
		GG	0.38	0.33	0.94	0.900
		C	0.38	0.39	Ref	
		G	0.62	0.61	1.04	0.849
rs9894946 (age: below 57)	AA	0.46	0.49	Ref		
	AG	0.40	0.43	0.98	0.945	
	GG	0.14	0.08	1.92	0.167	
	A	0.66	0.71	Ref		
	G	0.34	0.29	1.24	0.310	
	(age: above 57)	AA	0.42	0.55	Ref	
		AG	0.41	0.28	1.91	0.057
		GG	0.18	0.17	1.418	0.409
		A	0.62	0.69	Ref	
		G	0.38	0.31	1.38	0.148

0.008), showing a 2.41-fold increase in risk. There was also an elevated risk association for the G allele (χ^2 : 7.78; CI: 1.15–2.29; $p = 0.005$) with a 1.63-fold increase in risk, but this association was not observed in the rectal region. Notably, there was no discernible evidence indicating any association between the SNPs rs1642785 and rs9894946 and tumors in either the colon or rectal regions.

The pairwise linkage disequilibrium (LD) values (D' and r^2) are listed in Fig. 1. For the SNPs, LD analysis revealed weak LD, forming one haplotype block, suggesting that haplotype evaluation may be beneficial. SNP rs9894946 showed disequilibrium ($D' = 0.428$, $r^2 = 0.168$) in cases.

Table 4
Genotype frequencies of *TP53* gene polymorphism in colorectal cancer cases and controls based on sex: male and Female.

SNP	Variant	Cases (Freq)	Controls (Freq)	OR	P- Value	
rs1042522	Male	CC	0.44	0.56	Ref	
		CG	0.35	0.31	1.44	0.220
		GG	0.21	0.12	2.21	0.041
		C	0.62	156 (0.72)	Ref	
		G	0.38	0.27	1.62	0.017
	Female	CC	0.33	0.37	Ref	
		CG	0.43	0.47	1.04	0.909
		GG	0.24	0.15	1.80	0.199
		C	0.55	0.61	Ref	
		G	0.45	0.38	1.31	0.234
rs1642785	Male	CC	0.14	0.10	Ref	
		CG	0.43	0.48	0.65	0.321
		GG	0.43	0.42	0.76	0.542
		C	0.35	0.34	Ref	
		G	0.65	0.66	0.96	0.831
	Female	CC	0.16	0.08	Ref	
		CG	0.43	0.51	0.40	0.093
		GG	0.41	0.41	0.47	0.171
		C	0.38	0.33	Ref	
		G	0.63	0.67	0.82	0.418
rs9894946	Male	AA	0.45	0.54	Ref	
		AG	0.37	0.34	1.30	0.378
		GG	0.18	0.12	1.80	0.139
		A	0.63	153 (0.71)	Ref	
		G	0.37	0.29	1.40	0.093
	Female	AA	0.42	0.48	Ref	
		AG	0.45	0.41	1.22	0.555
		GG	0.13	0.11	1.32	0.592
		A	0.64	0.68	Ref	
		G	0.36	0.32	1.18	0.495

The remaining two SNPs showed $> 0.1 D'$ and r^2 values in cases and controls. Fig. 2 and Fig. 3 illustrate the regional LD plot for the *TP53* SNPs and the effect of amino acid replacement in SNP rs1042522 (Pro72Arg).

4. Discussion

TP53 protein is a crucial component in maintaining genomic integrity which prevents the cells from oncogenic transformation. Inactivation of *TP53* is common occurrence in majority of the cancers. *TP53* serves to regulate DNA repair, cell cycle, and cell development. Any changes in *TP53* function will abrupt these functions and culminate in loss of genome integrity. Polymorphisms of the *TP53* gene are widely established to take part in progression of CRC. If the *TP53* functioning is normal, it is a fundamental obstacle for carcinogenesis. Few SNPs of *TP53* coding region are strongly linked with the process of carcinogenesis. They are commonplace in a large number of cancers, contributing to severe aberration in *TP53* function. Till now, approximately > 200 SNPs are documented to be found commonly in *TP53* which are expected to cause aberrations in *TP53* functioning. The purpose of current study was to perceive any associations of *TP53* polymorphisms with development of CRC in Saudi Arabian population. This study investigated the role of exonic and intronic SNPs (exonic rs1042522, Pro 72 Arg (C > G), intronic rs1642785 (C > G), and intronic rs9894946 (A > G) of the *TP53* gene as potential CRC risk factors in a case-control study involving 192 CRC cases and 192 matched normal control samples.

Our study found a strong association between the GG genotype of SNP rs1042522 and an increased risk of CRC in the Saudi population. Additionally, we observed that the minor allele (G) of the same SNP also independently contributes to a higher susceptibility to CRC (Table 2).

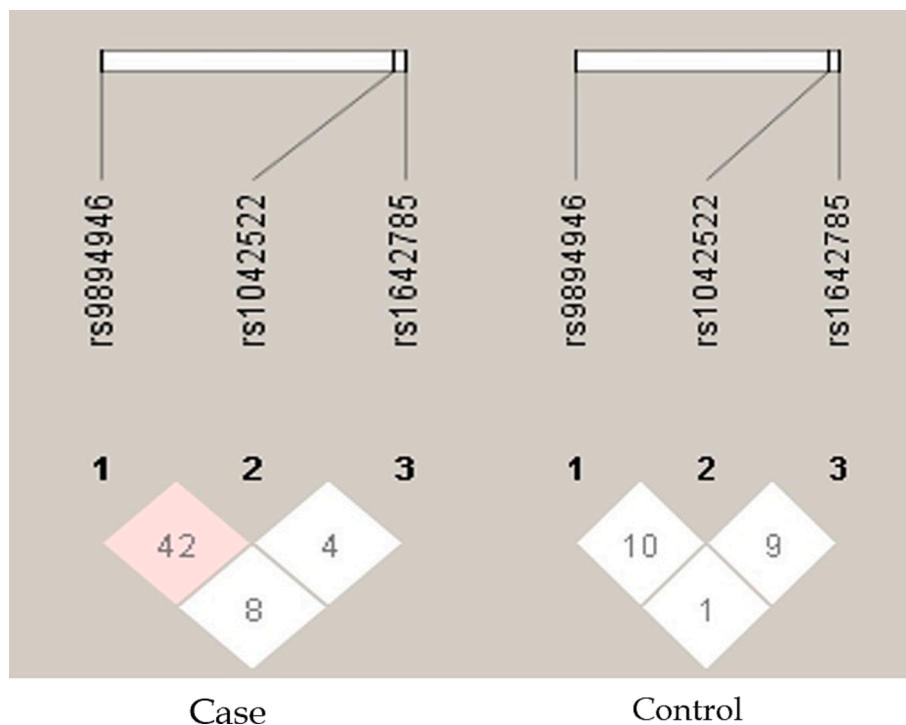


Fig. 1. Linkage Disequilibrium (LD) of studied *TP53* loci in colorectal cancer cases and controls.

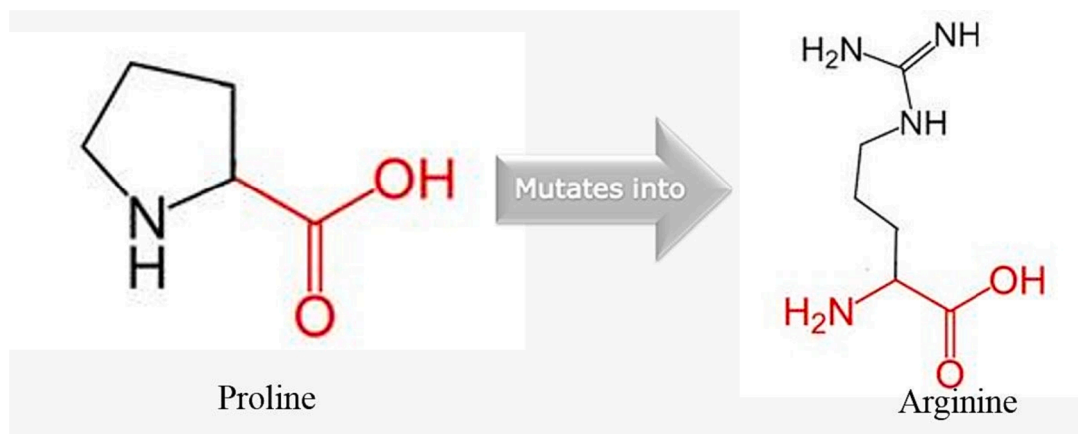


Fig. 2. Effect of amino acid replacement in SNP rs1042522 (Pro72Arg).

Our findings, consistent with previous research, demonstrate that individuals carrying the GG genotype of rs1042522 are more susceptible to CRC (Ashton et al., 2009, Ricks-Santi et al., 2010, Tian et al., 2016). This genotype is associated with a decreased ability of the TP53 protein to induce cell cycle arrest and apoptosis, which are vital mechanisms for controlling and eliminating potentially cancerous cells. Furthermore, our study has also unveiled an independent contribution of the minor allele (G) of SNP rs1042522 to an increased risk of CRC. This observation suggests that even individuals who possess a single copy of the G allele may be more predisposed to CRC than those with a different genetic makeup at this locus. Contrastingly, several other investigations have reported results inconsistent with our own findings. Notably, there is no discernible association between the rs1042522 SNP and cancer risk in their respective studies (Dahabreh et al., 2010; Kodali et al., 2017). In the current study, we sub-analyzed the possible association between three *TP53* SNPs and CRC risk by age, sex, and tumor location. In a prior publication of ours (Alhadheq et al., 2016, Semlali et al., 2017), we made a noteworthy observation concerning the SNP rs8679.

Specifically, we identified a statistically significant protective association between this SNP and the risk of colorectal cancer, particularly in females. The genotype distributions for TT, TC, and CC were as follows: in controls, 0.42, 0.49, and 0.09, respectively, whereas in cases, they were 0.67, 0.30, and 0.03, respectively. These findings indicated that the presence of certain alleles of rs8679 was associated with a lower susceptibility to colorectal cancer in females. It's worth noting that recent research conducted by Nazarian and Kulminski has shed new light on the potential genetic heterogeneity specific to sex in the context of colorectal cancer and lung cancer (Nazarian and Kulminski, 2021). Their work has explored the influence of sex at both the SNP (single nucleotide polymorphism) and pathway levels, revealing intriguing insights into how genetic factors may interact differently in males and females in relation to these cancers. These novel findings underscore the importance of considering sex-specific genetic factors when studying cancer susceptibility and may have significant implications for future research in this field. We noticed a noteworthy association between the rs1042522 polymorphism (CG, GG genotypes, and minor allele G) and

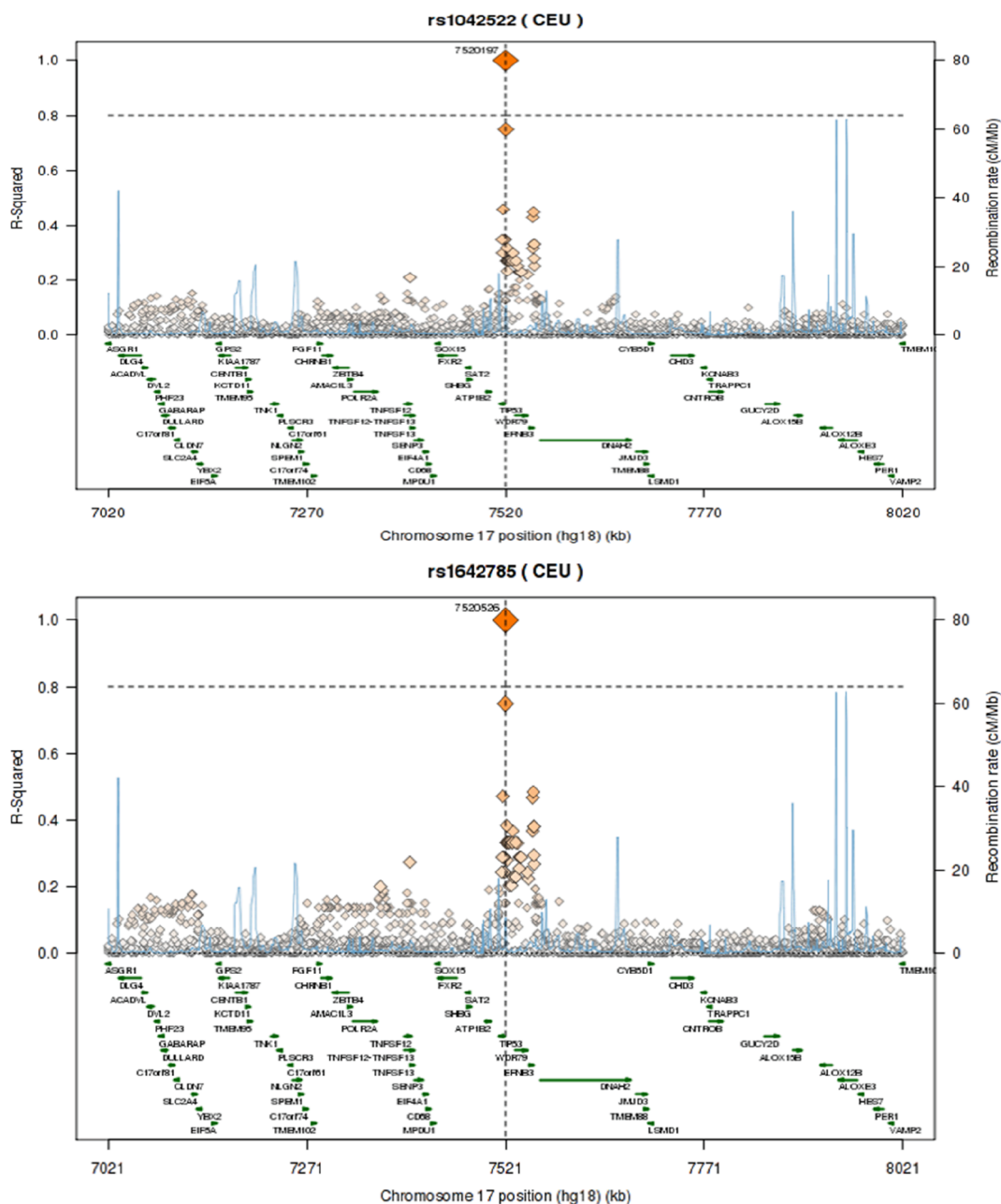


Fig. 3. Regional LD plot for the *TP53* SNPs (A) rs1042522; (B) rs1642785; (C) rs9894946.

an increased risk of CRC in individuals aged <57 years. Surprisingly, SNP rs1642785 demonstrated a significant protective effect against CRC in patients <57 years. Altogether, this finding offers valuable insights into the intricate interplay of genetic factors in colorectal cancer susceptibility, especially concerning age-related considerations. Further hold potential implications for the field of personalized medicine and refined risk assessment. The identification of individuals with distinct genetic profiles has the potential to facilitate early detection, precision screening, and proactive preventive strategies. When comparing sexes, we observed that the rs1042522 SNP was linked to a higher risk of CRC in males in contrast to females. This finding aligns with a prior study that highlighted the influence of sex in susceptibility to CRC (Micheli et al., 2009). Purim et al. reported that females are more protected against CRC development, with a lower related mortality rate when compared to males (Purim et al., 2013). The observed phenomenon may be attributed to variances in physiology, diet, and hormones between males

and females (Jacobs et al., 2007). Furthermore, SNP rs1042522 exhibited a noteworthy correlation with an elevated colorectal cancer (CRC) risk, particularly among patients with tumors located in the colon. It is conceivable that this SNP influences CRC risk by heightening the susceptibility of colon cells to DNA damage. Such DNA damage events can instigate mutations that foster the growth of cancer cells. Consequently, our findings imply that the rs1042522 SNP could serve as a valuable marker for the identification of individuals at an escalated risk of CRC. This insight holds the potential to guide the development of focused strategies for prevention and treatment tailored to this specific group of patients.

Various functional studies using genetically modified mice have consistently reported a notable link between *TP53* gene polymorphisms and the vulnerability of patients to tumor progression. These studies have consistently shown that mice with a silenced mutation in one *TP53* allele exhibit a lower incidence of tumors compared to mice with the

Acknowledgements

This work was funded by Princess Nourah bint Abdulrahman University Researchers Supporting Project number (PNURSP2023R62), Princess Nourah bint Abdulrahman University, Riyadh, Saudi Arabia. Researchers Supporting Project number (RSP2023R26), King Saud University, Riyadh, Saudi Arabia.

Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jksus.2023.102936>.

References

- Alanazi, M.S., Parine, N.R., Shaik, J.P., Alabdulkarim, H.A., Ajaj, S.A., Khan, Z., 2013. Association of single nucleotide polymorphisms in Wnt signaling pathway genes with breast cancer in Saudi patients. *PLoS One* 8 (3), e59555.
- Alhadheq, A.M., Purusottapatnam Shaik, J., Alamri, A., Aljebreen, A.M., Alharbi, O., Almadi, M.A., Alhadheq, F., Azzam, N.A., Semlali, A., Alanazi, M., 2016. The effect of Poly (ADP-ribose) Polymerase-1 Gene 3' untranslated region polymorphism in colorectal cancer risk among Saudi cohort. *Dis. Markers*.
- Angelopoulou, C., Veletza, S., Heliopoulos, I., Vadikolias, K., Tripsianis, G., Stathi, C., Piperidou, C., 2017. Association of SCN1A gene polymorphism with antiepileptic drug responsiveness in the population of Thrace, Greece. *Arch. Med. Sci.* 13 (1), 138.
- Ashton, K.A., Proietto, A., Otton, G., Symonds, I., McEvoy, M., Attia, J., Gilbert, M., Hamann, U., Scott, R.J., 2009. Polymorphisms in TP53 and MDM2 combined are associated with high grade endometrial cancer. *Gynecol. Oncol.* 113 (1), 109–114.
- Barrett, J.C., Fry, B., Maller, J., Daly, M.J., 2004. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 21 (2), 263–265.
- Chittleborough, T.J., Gutlic, I., Pearson, J.F., Watson, A., Bhatti, L.A., Buchwald, P., Frizelle, F., 2020. Increasing incidence of young-onset colorectal carcinoma A 3-country population analysis. *Dis. Colon Rectum* 63 (7), 903–910.
- Dahabreh, I.J., Linardou, H., Bouzika, P., Varvarigou, V., Murray, S., 2010. TP53 Arg72Pro polymorphism and colorectal cancer risk: a systematic review and meta-analysis. *Cancer Epidemiol. Prevention Biomarkers* 19 (7), 1840–1847.
- Dahabreh, I.J., Schmid, C.H., Lau, J., Varvarigou, V., Murray, S., Trikalinos, T.A., 2013. Genotype misclassification in genetic association studies of the rs1042522 TP53 (Arg72Pro) polymorphism: a systematic review of studies of breast, lung, colorectal, ovarian, and endometrial cancer. *Am. J. Epidemiol.* 177 (12), 1317–1325.
- Donehower, L.A., Harvey, M., Slagle, B.L., McArthur, M.J., Montgomery, C.A., Butel, J.S., Bradley, A., 1992. Mice deficient for P53 are developmentally normal but susceptible to spontaneous tumors. *Nature* 356 (6366), 215–221.
- Fearon, E.R., Vogelstein, B., 1990. A genetic model for colorectal tumorigenesis. *Cell* 61 (5), 759–767.
- Hernandez Borrero, L.J., El-Deiry, W.S., 2021. Tumor suppressor p53: Biology, signaling pathways, and therapeutic targeting. *Biochim. Biophys. Acta* 1876 (1), 188556.
- Houlston, R.S., Tomlinson, I.P., 2001. Polymorphisms and colorectal tumor risk. *Gastroenterology* 121 (2), 282–301.
- Hu, J., Cao, J., Topatana, W., Juengpanich, S., Li, S., Zhang, B., Shen, J., Cai, L., Cai, X., Chen, M., 2021. Targeting mutant p53 for cancer therapy: direct and indirect strategies. *J. Hematol. Oncol.* 14 (1), 157.
- Jacobs, E.T., Thompson, P.A., Martínez, M.E., 2007. Diet, gender, and colorectal neoplasia. *J. Clin. Gastroenterol.* 41 (8), 731–746.
- Jesioneck-Kupnicka, D., Braun, M., Trąbska-Kluch, B., Czech, J., Szybka, M., Szymańska, B., Kulczycka-Wojdala, D., Bienkowski, M., Kordek, R., Zawlik, I., 2017. MiR-21, miR-34a, miR-125b, miR-181d and miR-648 levels inversely correlate with MGMT and TP53 expression in primary glioblastoma patients. *Arch. Med. Sci.* 13 (1).
- Kanth, P., Inadomi, J.M., 2021. Screening and prevention of colorectal cancer. *BMJ* 374.
- Katerji, M., Duerksen-Hughes, P.J., 2021. DNA damage in cancer development: special implications in viral oncogenesis. *Am. J. Cancer Res.* 11 (8), 3956.
- Kodal, J.B., Vedel-Krogh, S., Kobylecki, C.J., Nordestgaard, B.G., Bojesen, S.E., 2017. TP53 Arg72Pro, mortality after cancer, and all-cause mortality in 105,200 individuals. *Sci. Rep.* 7 (1), 336.
- Lamb, P., Crawford, L., 1986. Characterization of the human p53 gene. *Mol. Cell Biol.* 6 (5), 1379–1385.
- Levine, A.J., Oren, M., 2009. The first 30 years of p53: growing ever more complex. *Nat. Rev. Cancer* 9 (10), 749–758.
- Micheli, A., Ciampichini, R., Oberaigner, W., Ciccolallo, L., de Vries, E., Izarzugaza, I., Zambon, P., Gatta, G., De Angelis, R., 2009. The advantage of women in cancer survival: an analysis of EURO CARE-4 data. *Eur. J. Cancer* 45 (6), 1017–1027 (E. W. Group).
- Michor, F., Iwasa, Y., Lengauer, C., Nowak, M.A., 2005. Dynamics of colorectal cancer. *Semin. Cancer Biol.*
- Miret, C., Molina, R., Filella, X., Garcia-Carrasco, M., Claver, G., Ingelmo, M., Ballesta, A., Font, J., 2003. Relationship of p53 with other oncogenes, cytokines and systemic lupus erythematosus activity. *Tumour Biol.* 24 (4), 185–188.
- Nazarian, A., Kulminski, A.M., 2021. Genome-wide analysis of sex disparities in the Genetic Architecture of Lung and colorectal cancers. *Genes* 12 (5), 686.
- Ozdemirkiran, F.G., Nalbantoglu, S., Gokgoz, Z., Payzin, B.K., Vural, F., Cagirgan, S., Berdeli, A., 2017. FAS/FASL gene polymorphisms in Turkish patients with chronic myeloproliferative disorders. *Arch. Med. Sci.* 13 (2), 426.
- Purim, O., Gordon, N., Brenner, B., 2013. Cancer of the colon and rectum: potential effects of sex-age interactions on incidence and outcome. *Med. Sci. Monit.* 19, 203–209.
- Ricks-Santi, L., Mason, T., Apprey, V., Ahaghotu, C., McLaughlin, A., Josey, D., Bonney, G., Dunston, G.M., 2010. p53 Pro72Arg polymorphism and prostate cancer in men of African descent. *Prostate* 70 (16), 1739–1745.
- Semlali, A., Parine, N.R., Al Amri, A., Azzi, A., Arafah, M., Kohailan, M., Shaik, J.P., Almadi, M.A., Aljebreen, A.M., Alharbi, O., 2017. association between TLR-9 polymorphisms and colon cancer susceptibility in saudi arabian female patients. *Oncotargets Ther.* 10, 1.
- Sobieszkoda, D., Czech, J., Gablo, N., Kopanska, M., Tabarkiewicz, J., Kolacinska, A., Robak, T., Zawlik, I., 2017. MGMT promoter methylation as a potential prognostic marker for acute leukemia. *Arch. Med. Sci.* 13 (6), 1433.
- Sung, H., Ferlay, J., Siegel, R.L., Laversanne, M., Soerjomataram, I., Jemal, A., Bray, F., 2021. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA. Cancer J. Clin.* 71 (3), 209–249.
- Thomas, M., Kalita, A., Labrecque, S., Pim, D., Banks, L., Matlashewski, G., 1999. Two polymorphic variants of wild-type p53 differ biochemically and biologically. *Mol. Cell Biol.* 19 (2), 1092–1100.
- Tian, X., Dai, S., Sun, J., Jiang, S., Jiang, Y., 2016. Association between TP53 Arg72Pro polymorphism and leukemia risk: a meta-analysis of 14 case-control studies. *Sci. Rep.* 6.
- Vuik, F.E., Nieuwenburg, S.A., Bardou, M., Lansdorp-Vogelaar, I., Dinis-Ribeiro, M., Bento, M.J., Spaander, M.C., 2019. Increasing incidence of colorectal cancer in young adults in Europe over the last 25 years. *Gut* 68 (10), 1820–1826.
- Werness, B.A., Levine, A.J., Howley, P.M., 1990. Association of human papillomavirus types 16 and 18 E6 proteins with p53. *Science* 248 (4951), 76–79.
- Whibley, C., Pharoah, P.D., Hollstein, M., 2009. p53 polymorphisms: cancer implications. *Nat. Rev. Cancer* 9 (2), 95–107.