Journal of King Saud University - Science 35 (2023) 102453

Contents lists available at ScienceDirect

Journal of King Saud University – Science

journal homepage: www.sciencedirect.com

Original article

HOSTED BY

Lipid composition and oxidative changes in diabetes and alcoholic diabetes rats



Lin Qin ^{a,b,c,d}, Shaik Althaf Hussain ^e, Narendra Maddu ^f, Chinna Padamala Manjuvani ^f, Bangeppagari Manjunatha ⁱ, Sudhakara Gujjala ^f, Venkata Subba Reddy Gangireddygari ^g, Yingying Fan ^{h,*}

^a School of Pharmaceutical Science, Kunming Medical University, Kunming, Yunnan 650500, China

^b Department of Endocrinology, The First Affiliated Hospital of Kunming Medical University, Kunming, Yunnan 650000, China

^c Yunnan Province Clinical Research Center for Metabolic Diseases, Kunming, Yunnan 650000, China

^d Yunnan Clinical Medical Center for Endocrine and Metabolic Diseases, Kunming, Yunnan 650000, China

^e Department of Zoology, College of Science, King Saud University, Riyadh, Saudi Arabia

^fDepartment of Biochemistry, Sri Krishnadevaraya University, Anantapur 515 055, India

^g Plant Virus Research, Horticultural and Herbal Crop Environment Division, National Institute of Horticultural and Herbal Science, Rural Development Administration, Wanju 55365, Republic of Korea

^h Department of Anesthesiology, Xi'an Fifth Hospital, Xi'an 710082, China

Department of Cell Biology and Molecular Genetics, Sri Devaraj Urs Academy of Higher Education and Research, Sri Devaraj Urs Medical College, Tamaka, Kolar-563103, India

ARTICLE INFO

Article history: Received 7 September 2022 Revised 6 November 2022 Accepted 14 November 2022 Available online 18 November 2022

Keywords: Diabetic Alcoholic Nitric oxide Lipid peroxidation

ABSTRACT

Background: The investigating of study was expected to the lipid composition of diabetes and alcoholic diabetes in plasma and erythrocyte membrane biochemical profile in rats. Diabetic male *Wistar* Streptozotocin (STZ)-induced rats were used experimental models. Control rats (C) were maintained group I, received glucose (i.e., caloric equivalent), diabetic induced rats (STZ) group II, and group III alcoholic treated and IV diabetic and alcoholic treated rats which received 20 % (v/v) alcohol in water, administered through stomach tube (5 g/kg body weight/day).

Results: STZ-induced diabetic hepatic damage in rats was observed which leads to the increased plasma lipid peroxidation, nitrate and nitrite levels. Diabetic and administration alcohol rats also suggestively lesser the activities of antioxidants, glutathione peroxidase, glutathione S-transferase, superoxide dismutase, catalase and reduced glutathione when related with control rats (group I). Plasma enzymes are normal levels and renovate the enzymic and non-enzymatic antioxidants level in experimental groups.

Conclusion: The present data point out that the nitric oxide scavenging levels increased and may possibly protect and adjacent to oxidative stress and free radicals in diabetic hepatopathy rats and histopathological studies were identified in regular hepatic cortex of Group II, III and IV of animals with diabetes, alcoholic and alcohol-induced diabetes rats.

© 2022 Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

The most broadly used and regularly abused psychoactive drug is alcohol in the world with stern communal and well-being insin-

* Corresponding author.

E-mail address: fanyingxa@sina.com (Y. Fan).

Peer review under responsibility of King Saud University.

ELSEVIER



uations (Xu et al., 2005). As per the worldwide position based on the 2004 report of alcohol (WHO) exposed that extra two billion people have been intense alcohol worldwide with increase in the amount with the adding of new drinkers each year counting adolescent youngsters and young woman (World Health Organization, 2004; Lieber, 2000). DM (diabetes mellitus) is a chronic metabolic illness categorized by imbalances in carbohydrate, protein and lipid metabolisms, due to faulty or shortage in insulin action and secretion (Lu and Cederbaum, 2008). The near relation amid ethanol and liver damage is mostly owing to the fact that 80 % of swallowed liquor is absorbed in the liver. Throughout alcohol/ethanol absorption numerous sensitive oxygen species are produced via cytochrome P450 (Tuma and Casey, 2003). The aim of

https://doi.org/10.1016/j.jksus.2022.102453

1018-3647/© 2022 Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/). work was to expand an investigational model of Streptozotocin (STZ) treated to diabetic male Wistar rats were used as investigational rats, control and Diabetes and alcoholic treated and diabetic + alcoholic treated (body wt/day g/kg 5) for sixty days. Diabetes alcoholic treated and diabetic + alcoholic treated hepatocyte/retinopathy damage. Nitric oxide (NO) is a significant intermediary of a lot of functions of physiological, and its position in the pathogenesis of several diseases is attainment in the identification. Diabetic, alcoholic treated and diabetic + alcoholic treated utilization increased levels of NO and may show the mode to toxicity by nitrites of peroxides, a strong oxidant. Therefore, the work investigates and evaluate the changes in various plasma biochemical parameters SGOT, SGPT and lipid peroxidation. NO is a significant go-between of a lot of physiological function, and its role in the pathogenesis of a lot of disease is attainment credit (Pacher et al., 2007). Diabetic, alcoholic treated and diabetic + alcoholic treated consumption increases NO level and may lead to toxicity by peroxynitrite, a potent oxidant (Venkatraman et al., 2004). Thoughtless nitrogen species/reactive oxygen species (RNS/ROS) let go of overproduction might take place when its production in an arrangement exceed the system's capability to counterbalance and abolish them.

Lipid peroxidation, antioxidant levels of rats to appreciate the position and importance of nitric oxide in diabetes, alcoholic and diabetes + alcoholic treated rats with plasma and erythrocyte membrane chemical and physical alterations and may it lead hepatic damage. In wide-ranging and toxicity of these drugs, with its numerous troubles at a time, have an effect on altered organs causes some disorders. The numerous pathogenicity of diabetic, alcoholic treated and diabetic + alcoholic treated burden many modes of healing come near to fight and adjacent to such trouble by modulating activities of enzymes, metabolism, receptor performance, transduction of signal mechanism and scavenge free radicals at a mixture of levels (Trinder, 1969). There has been a look for safer customary nutritional supplementation of inhabitant plant extracts containing some principles for healing reason with multiple targets for treating multiple pathologies of diabetic plus alcoholic rats.

2. Methods

2.1. Subjects for study and experimental design

60 days male Wistar rats, about 120-140 g weighing, are maintained in animal house. They were feed among pellet diet and tap water ad libitum. The rats were separated into four groups of 8 rats in each. Group I normal rats (C), which inward sugar in its place of diabetic/alcohol (caloric equal), group II diabetic Streptozotocin (STZ) induced rats, III group (5 g/kg body weight/day) alcohol treated rats, and group IV diabetic plus alcohol administered from sideto-side abdomen tube procured from animal house in creature cages in an AC room (25 \pm 1 °C) with daylight from 7:00 a.m to 7:00p.m. Hence the present work determined on the result of diabetes and alcohol induces oxidative injure/alteration in plasma with pressure on its machinery. The study was established by institutional ethical committee. The study for the night fasted blood sample use from subjects. The dose of the sample is set based on our previous study (Allian et al., 1974; Reddy et al., 2009). Administered way out two months by using intragastric pipe every day. Foodstuff and water eating of every rat was record every day and maintained mass on every other day. At the end of the trial time, the rats in each set were fasted during the night and then kill by cervical dislocation.

2.2. Blood collection and determinations of plasma nitric oxide scavenge action

By Trinder process (Sreejayan and Rao, 1997) glucose was estimated spectrophotometrically by kits. NO estimation by Greiss reaction (Ohkawa et al., 1979) and generated from sodium nitroprusside was measured. NO Scavengers, compete with oxygen important to summary assembly of NO. Phosphate buffered saline in sodium nitroprusside (5 mM) were mixed with various concentration and incubate at 150 min in 25 °C. Greiss reagent (5 % ophosphoric acid, 1 % sulfanilamide and 0.01 % naphthylethylene diamine) and the samples from the above were reacted. The absorbance of the chromophore produced throughout diazotization of nitrite with sulfanilamide and subsequent mixture with napthylethylene diamine was examined in UV-visible spectrophotometer at 546 nm.

2.3. In vivo assays

Tissues dissect out from the liver, wash and weighed with way out of saline with ice cold. Muscles were crushed and homogenized (10 % w/v) in Tris–HCl buffer (0.1 M; pH 7.4) and centrifugation at 10,000 g for 4 °C in 20 min. The resultant upper part was use for different assays.

2.4. Estimation of thiobarbituric acid (TBA) and protein carbonyls

TBARS was calculated by the configuration of malondialdehyde (MDA) by the process of Ohkawa et al. (Reznick and Packer, 1994). The absorption of protein carbonyls was calculated using 2, 4-dinitrophenylhydrazine (DNPH) evaluate as describe formerly (Abei, 1988).

2.5. Antioxidant importance, nitrite and nitrate analysis were calculated

Abei (Mishra and Fridovich, 1972) catalase (CAT) was evaluated as deliberate. The expression of CAT action was as nmol. SOD was assay utilize the process of Mishra and Fridovich (Rotruck et al., 1973). Enzyme was articulated as 50 % reserve of NBT reduction/ min/mg protein. The processes describe by Rotruk (Habig et al., 1974) and action measured GPx (glutathione peroxidase). GPx action was articulated lmol. GST action was calculated according to the method of Habig et al. (Ellman, 1959). GST action articulated as Imol. Whole GSH content was intended by Ellman's (Sastry et al., 2002). By Sastry et al. (Lowry et al., 1951) nitrite and nitrates were determined. Absorption of protein was determined by Lowry et al. (Pigeolot et al., 1990) method. Plasma transaminases, glutamate oxalo acetate transaminase (SGOT) and glutamate pyruvate transaminase (SGPT) were measured by Reitman and Frankel (1957) methods. The histological sections of the hepatocytes of rats were taken by adopting the procedure as described by Humason (1972).

3. Results and discussion

Information presented in plasma enzymes (SGPT) glutamate pyruvate trasminase levels were not changed and (SGOT) glutamate oxalo acetate trasminase propose that an important raise in Table 1, the levels were significantly decreased in diabetes treated and alcoholic and diabetes + alcoholic, administrated rats. Na⁺-K⁺-ATPase actions in erythrocytes were sightly increased the group II and group III, IV experimental rats with group I control

L. Qin, S. Althaf Hussain, N. Maddu et al.

Table 1

Effect of diabetes and alcohol diabetes on the activities of serum enzymes, Activities of Na+- K + ATPase and glycolated enzymes in erythrocytes of rats.

Parameter	Groups				
	Controls Rats	Diabetes treated rats	Alcohol administration rats	Diabetes with alcohol administration rats	
Glutamate oxalo acetate transaminase (GOT) (IU/L) Glutamate Pyruvate Trasminase) (GPT) (IU/L) Hexokinase (IU/gm Hb) Na + -K + ATPase (µg pi liberated/min/mg/protein)	51.6 ± 2.60^{a} 24.6 ± 1.64 ^a 0.93 ± 0.01 ^a 1.31 ± 0.03 ^a	54.2 ± 2.20^{b} 22.2 ± 1.40 ^a 0.95 ± 0.24 ^a 1.42 ± 0.05 ^b	$57.6 \pm 2.38^{\rm b} \\ 24.5 \pm 1.32^{\rm a} \\ 0.94 \pm 0.36^{\rm a} \\ 1.40 \pm 0.04^{\rm b} \\ \end{cases}$	$56.5 \pm 2.40^{b} \\ 24.2 \pm 1.21^{b} \\ 0.95 \pm 0.35^{a} \\ 1.41 \pm 0.04^{b}$	

All Table values are expressed as Mean \pm SEM, in each column followed by the same letter are not significantly different ($P \le 0.05$) from each other according to Duncan's Multiple Range (DMR) test, n = 8.

Table 2

Diabetes, alcohol and alcoholic diabetes rats with effect of antioxidant enzymes.

Parameter	Groups				
	Controls Rats	Diabetes treated rats	Alcohol administration rats	Diabetes with alcohol administration rats	
Superoxide Dismutase (SOD) (Units/min/mg Hb)	5.7 ± 0.62 ^a	5.4 ± 0.76^{b}	$5.3 \pm 0.54^{\rm b}$	5.5 ± 0.55 ^b	
Catalase (CAT) (IU/10 ⁴ /gm Hb)	8.2 ± 0.42^{a}	7.6 ± 0.81^{a}	7.7 ± 0.62^{b}	7.6 ± 0.63^{b}	
Red Cell Reduced glutathione (GSH) (µ moles/gm Hb)	3.4 ± 0.16^{a}	3.3 ± 0.09^{b}	3.4 ± 0.07^{b}	3.4 ± 0.08^{b}	
Glutathione S-transferase (^mol/mg/min)	2.89 ± 0.49	2.87 ± 0.44	2.69 ± 0.47	2.59 ± 0.42	
Glutathione peroxidase(G-Px) (IU/gm Hb)	16.5 ± 1.04^{a}	15.2 ± 1.21 ^b	14.3 ± 1.20^{b}	14.7 ± 1.22 ^b	

All Table values are expressed as Mean \pm SEM, in each column followed by the same letter are not significantly different ($P \le 0.05$) from each other according to Duncan's Multiple Range (DMR) test, n = 8.

Table 3 Diabetes, alcohol and alcoholic diabetes rats with effect of plasma LPO and NO₂ and NO₃ levels.

Parameter	Groups				
	Controls Rats	Diabetes treated rats	Alcohol administration rats	Diabetes with alcohol administration rats	
Plasma glucose (mg/dl)	81.12 ± 0.17a	102.16 ± 0.30b	85.12 ± 0.32b	104.12 ± 0.35b	
Plasma lipid peroxidation (p mole of MDA formed/mg protein)	1.92 ± 0.01a	2.11 ± 0.20b	2.15 ± 0.28b	2.40 ± 0.26b	
Plasma NO ₂ (µ mole /L)	2.53 ± 0.01a	3.82 ± 0.16b	4.50 ± 0.42b	4.17 ± 0.40b	
Plasma NO ₃ (μmole /L)	24.66 ± 0.01a	25.74 ± 0.01b	32.52 ± 0.01b	39.62 ± 0.01b	

All Table values are expressed as Mean \pm SEM, in each column followed by the same letter are not significantly different ($P \le 0.05$) from each other according to Duncan's Multiple Range (DMR) test, n = 8.

rat's summarization and glycolated enzymes in erythrocyte and no alter of hexokinase in groups of (group II and group III, IV). Table 3 revealed the differences in plasma parameters viz., plasma glucose, plasma LPO, nitrate and nitrite, in normal male and experimental subjects. Additionally, the effect of diabetic rats and alcoholic rats and diabetic + alcoholic on the on top of parameter supervision at certain quantity in male rats. Reported antioxidant enzymes decreased significantly namely GST, GPx, CAT, SOD and GSH the contented of in liver were specific in Table 2. A significant (P < 0.05) in the performance was observed decreased in treated diabetes, alcoholic and diabetic + alcoholic rats compare to control of antioxidant enzymes.

Enzymes of free radical scavenge like GPx, SOD, GST and CAT, are the major line of defence against the injury of oxidatives were the catalase and superoxide ions (SOD) scavenges transfer H_2O_2 to water. In this study it is observed that there is a less impact of GPx, SOD and CAT, at the same time GSH in diabetes, diabetic treated rats, alcoholic treated rats and diabetes + alcoholic tested rats while compare with control rats. Owing to the oxidative inactivation of the enzyme there is a reduction in activity of SOD therefore as result there is an excessive oxygen species generation (Wilce and Parker, 1994). The isozymes that catalyze the conjugation of

GSH to a diversity of electrophilic compound where GSTs are a multigene family of isozymes, whereby it as decisive role in protecting cellular touching ROS (Hayes and Pulford, 1995; Alin et al., 1985). Since the GST activity is reduced there is a compromise detoxification of a toxic aldehyde, 4-hydroxynonenal and a creation of LPO. Hence it leads to decrease in enzyme action or appearance may supply to drugs hepatotoxicity specifically targeting GST isoenzymes by drugs its metabolic products (Gueeri, 1995). As GSH preservation of usual cell arrangement and function from side-to-side detoxification reaction and its redox (Das, 1997). Thus, GSH acting a major role in coordinate the antioxidant protection procedure as it is the central non-protein thiols. Glutathione is a tripeptide there in approximately all the cells and acting a significant position in metabolism, cellular production and transport beside oxidation by reactive oxygen and free radical's intermediates in investigational groups II, III and IV with compare to group I control (Das et al., 1997; Saxena et al., 1993). These reactive oxygen class levels of are required by viz., glutathione reductase antioxidant enzymes (Oberley, 1998); catalyzing the reduce of GSSG to GSH and NADPH utilize, make in path of HMP shunt, glutathione peroxidase catalyzing the reduce of organic hydro peroxides and hydrogen peroxide (Jacob, 1995; Dincer et al., 2002) by



Control rats

Diabetes Treated rats



Alcohol Administration rats

Diabetes with Alcohol Administration rats

Fig. 1. Histopathological changes in Hepatocytes.

SOD, GSH catalyzes the reduce of superoxide radical to hydrogen peroxide (Ashour et al., 1999) and CAT the reduction of hydrogen peroxide to water (Deisseroth and Dounce, 1970) and multifunctional protein, glutathione-S-transferase is catalyzing the conjugation between GSH with a lane variety of minor substrates and contribute, an very important place in the detoxification of xenobiotics (Denake and Fanburg, 1989). The endogenous glutathione and glutathione peroxidase association and catalase are significant antioxidants and cytoprotective apparatus in the hepatocytes expose to on top of drugs. GR is worried with the preservation of cellular level of concentrated GSH.

Catalase and GR performance are enlarged as importance of drugs exposure. Diabetes alcoholic and diabetes + alcoholic induced injuries by oxidative and heart failure rat have been reported to associated with risk of cardiovascular diseases. Rats were constantly exposed to 3 months constantly per day by oral gavages into circulation and change in cardiac markers. The present study was aimed to diabetes and alcoholic and diabetes + alco holic induced lipid composition changes in male albino Wistar rats, demonstrates an important raise in the level in liver activities and asparate transaminase, alanine transaminase in serum with subsequent decrease with cardiac markers, and also show an increase significant in the levels of glucose in blood. Hence, the results of our study demonstrate that it makes lipid composition changes and lipid toxicity. Fig. 1 showed the histopathological examination changes observed in the diabetes, alcohol treated and diabetes with alcohol treated rats compared to control rats in hepatocytes cells.

4. Conclusion

Increase glucose in blood levels and significant lift in serum SGOT, SGPT values were decrease drastically in diabetes treated and alcoholic and diabetes + alcoholic, administrated rats, NO scavenging compounds presently increased and may protect beside free radical mediate in rat hepatocytes oxidative stress. Reduce act of GPx CAT and SOD as well GSH in diabetes treated and alcoholic and diabetes + alcoholic treated rats compared to control rats. Further, the biochemical activities have been supported by the histopathological studies. Additional study is essential to associate the toxic property of long-lasting make use of diabetes treated and alcoholic and diabetes + alcoholic on rat lipid composition.

5. Contributions of authors'

All authors were concerned in the scientific assessment, analytic for the cases describe in this document. All authors read and accepted the concluding version.

Funding

The authors extend their appreciation to the Researchers Supporting Program for funding this work through Researchers Supporting Project number (RSP-2021/371), King Saud University, Riyadh, Saudi Arabia. This work also financially supported by Yunnan Province Clinical Research Center for Metabolic Diseases (202102AA100056), Science and Technology Innovation Team of Diagnosis and Treatment for Glucolipid Metabolic Diseases in Kunming Medical University (CXTD202106) and The National Natural Science Foundation of China (Grant No 82160164).

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

The authors are highly thankful and acknowledge the funding support by the Researchers Supporting Project number (RSP-2021/371), King Saud University, Riyadh, Saudi Arabia. The authors also thankful for financial support to Yunnan Province Clinical Research Center for Metabolic Diseases (202102AA100056), Science and Technology Innovation Team of Diagnosis and Treatment for Glucolipid Metabolic Diseases in Kunming Medical University (CXTD202106) and The National Natural Science Foundation of China (Grant No 82160164).

References

Abei, H., 1988. Catalase in vitro. MethodsEnzymol 10, 121–126.

- Alin, P., Danielson, U.H., Mannervik, B., 1985. 4-Hydroxyalk-2-enals are substrates for glutathione transferase. FEBS Lett. 179, 267–270.
- Allian, C.C., Poon, L.S., Chan, C.S.G., Richmans, W., Fu, P., 1974. Enzymatic determination of total serum cholesterol. Clin Chem 20, 470–475.
- Ashour, M., Salem, S., Hassaneen, H., EL. Gadban, H., Elwan, N., Awad A., Basu T.K. 1999. Antioxidant status and insulin dependent diabetes mellitus. Journal of Clinical Biochemistry. 29, 99-107.
- Das, D., Bandyopadhyoy, D., Bhatterjee, N., Baneijee, R.K., 1997. Hydroxy radical is the major causative factor in stress induced gastric ulceration. Free Radic. Biol. Med. 23, 8–18.
- Das, M., 1997. Glutathione status of some homeothermic vertebtate species and its relation with diabetogenesis. Indian J. Exp. Biol. 35, 66–662.
- Deisseroth, A., Dounce, A., 1970. Catalase: physical and chemical properties. Mechanism of catalysis and physiological role. Physiological review. 50 (3), 319–375.
- Denake, S.M., Fanburg, B.L., 1989. Regulation of cellular glutathione. American Journal of Physiology. 257 (1163–1), 173.
- Dincer, Y., Alademir, Z., likova, H., Akcay, T., 2002. Susceptibility of glutathione and glutathione related antioxidant activity to hydrogen peroxide in patients with type 2 diabetes: effect of glycemic control. Clin. Biochem. 35, 297–301.
- Ellman, G.L., 1959. Tissue sulfhydryl groups. Arch Biochem Biophys 82, 70-77.
- Gueeri, H., 1995. Influence on prolonged ethanol intake on the level and turnover of alcohol and aldehyde dehydrogenase and glutathione. Adv Exp Med Biol. 23, 12–14.

- Habig, W.H., Pabst, M.J., Glutathione-S-transferases, J.WB., 1974. The first enzymatic step in mercapturic acid formation. J Biol Chem 249, 7130–7139.
- Hayes, J.D., Pulford, D.J., 1995. The glutathione S-transferase supergene family: regulation of GST and the contribution of the isoenzymes to cancer chemoprotection and drug resistance. Crit Rev Biochem Mol Biol. 30, 445–600.
- Humason, G.L., 1972. Animal tissue techniques. 3rd. Freeman San Francisco, CA. Jacob, R.A., 1995. The integrated antioxidant system. Nutr. Res. 15, 755–766.
- Lieber, C.S., 2000. Alcohol and the liver: metabolism of alcohol and its role in hepatic and entrahepatic diseases. The mount Sinai journal of medicine. 67 (1), 84–94.
- Lowry, O.H., Rosebrough, N.J., Farr, A.L., Randall, R., 1951. Protein measurement with the Folin-phenol reagent. J Biol Chem 193, 263–275.
- Lu, Y., Cederbaum, A.I., 2008. CYP2E1 and oxidative liver injury by alcohol. Free Radic Biol Med 44, 723–738.
- Mishra, P.H., Fridovich, I., 1972. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. J Biol Chem 247, 3170–3175.
- Oberley, L.W., 1998. Free radicals and diabetes. Free Radical Biolog}' and Medicine. 5, 113–124.
- Ohkawa, H., Ohishi, N., Yagi, K., 1979. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal Biochem 95, 351–358.
- Pacher, P., Beckman, J.S., Liaudet, L., 2007. Nitric oxide and peroxynitrite in health and disease. Physiol Rev 87, 315–424.
- Pigeolot, E., Corbisier, P., Houbion, A., Lambert, D., Michiels, C., Raes, M., 1990. Glutathione peroxidase, superoxide dismutase and catalase inactivation by peroxide and oxygen derived radicals. Mech Ageing Dev. 51, 283–297.
- Reddy, V.D., Padmavathi, P., Paramahamsa, M., Varadacharyulu, N.C., 2009. Modulatory role of *Emblica officinalis* against alcohol induced biochemical and biophysical changes in rat erythrocyte membranes. FoodChem Toxicol 47, 1958–1963.
- Reitman, S., Frankel, S., 1957. Transaminases. Am. J. Clin. Pathol. 28, 56. In: Methods of Enzymatic Analysis, 2 1x1 edn. Bergmeyer, H.U. (ed.), vol 2, Verlag Chemie Weinheim, Academic Press Inc, New York, pp. 735–739, 760–764.
- Reznick, A.Z., Packer, L., 1994. Oxidative damage to proteins: spectroscopic method for carbonyl assay. MethodsEnzymol 233, 357–363.
- Rotruck, J.T., Pope, A.L., Ganther, H.E., Swanson, A.B., Hafeman, D.G., Hoekstra, W.G., 1973. Selenium: biochemical role as a component of glutathione peroxidase. Science 179, 588-590.
- Sastry, K.V.H., Moudgal, R.P., Mohan, J., Tyagi, J.S., Rao, G.S., 2002. Spectrophotometric determination of serum nitrite and nitrate by coppercadmium alloy. Anal Biochem 306, 79–82.
- Saxena, A.K., Srivastava, P., Kale, R.K., Baquer, N.Z., 1993. Impaired antioxidant status in diabetic rat liver. Effect of vanadate. Biochemistry Pharmacology. 45 (3), 569–572.
- Sreejayan, N., Rao, M.N.A., 1997. Nitric oxide scavenging by curcuminoids. J Pharm Pharmacol 49, 105–107.
- Trinder, P., 1969. Determination of glucose-by-glucose oxide method. Ann Clin Biochem 6, 24–26.
- Tuma, D.J., Casey, C.A., 2003. Dangerous byproducts of alcohol breakdown-focus on adducts. Alcohol Res Health 27, 285–290.
- Venkatraman, A., Shiva, S., Wigley, A., Ulasova, E., Shhieng, D., Bailey, S.M., et al., 2004. The role of iNOS in alcohol-dependent hepatotoxicity and mitochondrial dysfunction in mice. Hepatology 40, 565–573.
- Wilce, M.C., Parker, M.W., 1994. Structure and function of glutathione stransferases. Biochim Biophys Acta. 1205, 1–18.
- World Health Organization. 2004. Global status report on alcohol: alcohol policy: Geneva: World Health Organization.
- Xu, J., Zheng, Y.N., Sung, C.K., 2005. Natural medicines for alcoholism treatment: a review. Drug Alcohol Rev. 24, 525–536.