



Full Length Article

Polyphenolic contents and antioxidant potential in *Nasturtium officinale*

Sobia Zaman^a, Raza Ahmad^a, Manal Abdulaziz Binobead^b, Mohamed Ragab Abdel Gawwad^c,
 Mohamed Soliman Elshikh^d, Yusufjon Gafforov^{e,f,g}, Arshad Mehmood Abbasi^{h,i,*}

^a Department of Biotechnology, COMSATS University Islamabad, Abbottabad Campus, 22060 Abbottabad, Pakistan

^b Department of Food Science and Nutrition, College of Agriculture Food Science, King Saud university, Riyadh, Saudi Arabia

^c Genetics and Bioengineering, Faculty of Engineering and Natural Sciences, International University of Sarajevo, 71210 Sarajevo, Bosnia and Herzegovina

^d Department of Botany and Microbiology, College of Science, King Saud University, P.O.Box 2455, Riyadh, 11451, Saudi Arabia

^e Central Asian Center for Development Studies, New Uzbekistan University, Tashkent, 100007, Uzbekistan

^f Department of Education and Training Management, Tashkent International University of Education, Tashkent, 100207, Uzbekistan

^g Institute of Botany, Academy of Sciences of Republic of Uzbekistan, Tashkent 100125, Uzbekistan

^h Department of Environmental Sciences, COMSATS University Islamabad, Abbottabad Campus 22060 Abbottabad, Pakistan

ⁱ University of Gastronomic Sciences of Pollenzo, Piazza V. Emanuele II, I-12042, Bra/Pollenzo, Italy

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ABSTRACT

The present study was conducted with the aim to assess the impact of environmental conditions on polyphenolic content and free radical scavenging ability in the leaves of *N. officinale* used as food and medicine in the Himalayan region of Pakistan. Samples were gathered from six different sites located in Himalaya, Hindukush, and Karakorum ranges. Polyphenolic profiling, and in vitro antioxidant potential were determined using standard analytical techniques. The samples collected from Abbottabad had significant levels ($p \leq 0.05$) of total phenolic (331.43 ± 41.43 mg GAE/100g DW) and total flavonoid contents (8.45 ± 1.53 mg RutE/100g DW). Whereas, rutin, quercetin and kaempferol were maximum in samples collected from Dir (132.46 ± 0.31 , 8.80 ± 0.05 and 0.95 ± 0.04 $\mu\text{g/g}$, respectively). Likewise, Gallic and caffeic acids were highest in Haripur and Swat samples, whereas ferulic and *p*-coumaric acids were maximum in samples taken from Dir. The percentage scavenging of DPPH was maximum in samples collected from Chitral ($69.57 \pm 3.39\%$) and that of H_2O_2 at Dir ($57.24 \pm 8.05\%$), whereas samples collected from Abbottabad depicted maximum inhibition of ferric and molybdate ions. Total phenolic and total flavonoids contents exhibited highly significant positive correlations ($p \leq 0.01$) with ferric and molybdate ions reduction capacity. Cluster and principal component analyses also depicted significant associations among samples collected from Abbottabad-Haripur, Dir-Swat, and Mansehra-Chilas. Our findings contribute in understanding the antioxidant potential of *N. officinale* leaves, and highlights the importance of polyphenolic composition and environmental factors in future investigations related to its nutraceutical applications. Further research on detailed phytochemical profiling, their in vitro and in vivo mechanisms of action could unveil the potential of this plant as a functional food.

1. Introduction

Polyphenolics offer a wide range of health benefits to human beings because of their anti-inflammatory, anti-allergic, antiviral, anticancer, antihypertensive, antioxidant, and anticancer properties (Montenegro-Landívar et al., 2021). These compounds are abundant in vegetables, fruits, legumes, grains, spices, medicinal plants, and drinks (Ren et al., 2021). Watercress (*Nasturtium officinale* L.), belonging to family Brassicaceae is a perennial herb with origins in Europe and Asia. This plant is typically found in freshwater springs, streams, and canals (Klimek-

Szczykutowicz et al., 2018). Watercress is eaten as a vegetable, used in different cuisines, soups, meat dishes, pasta, and also as salad (Goo-goolee et al., 2022). The usage of *N. officinale* in conventional medicine includes the treatment of hyperglycemia, hypercholesterolemia, bronchitis, hypertension, arthritis, odontalgia, scurvy, and diuresis. And such medicinal properties of this plant are a result of polyphenolics, glucosinolates, isothiocyanates, and various vitamins (Panahi Kokhdan et al., 2021). Furthermore, these secondary metabolites support antioxidant capacity of *N. officinale*, which reduces cellular lipid peroxidation, superoxide anion, and free radicals' damage (Mahmood et al., 2021). In

* Corresponding author at: Department of Environmental Sciences, COMSATS University Islamabad, Abbottabad Campus 22060 Abbottabad, Pakistan.

E-mail addresses: arshad799@yahoo.com, amabbasi@cuiatd.edu.pk (A. Mehmood Abbasi).

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Pakistan, fresh leaves of *N. officinale* are cooked as vegetable, and is consider as a coolong and appetizing agent, and to control consitpation.

Genetic diversity, growing conditions, agricultural activities, harvesting and post-harvesting techniques, processing and analytical techniques, and consuming methods all have a substantial impact on the functional properties of food and medicinal plants (Prabhu et al., 2021; Tuladhar et al., 2021). The nutritional value, phytochemical concentration, and bioactive potential of the same plant species or varieties of same species grown under diverse environmental circumstances may differ considerably. For instance, Bibi et al. (2022), reported substantial variation in polyphenolic contents and free radical secevning potential of onion varieties grown in diverse agro-ecological zones of Pakistan. In this context, present stduy was based on the hypothesis that there may be variation in the polyphenolic contents and bioactive potential of *N. officinale* which grows naturally in diverse agroecological zones of Pakistan. Our main goal was to determine free radicals' scavenging potential and polyphenolics profiling in the leaves of *N. officinale* gathered from various parts of Pakistan.

2. Materials and Methods

2.1. Sampling and extraction

Fresh leaves of *N. officinale* (≈ 1.0 kg) were collected from Abbottabad, Haripur, Muzaffarabad (Himalayan), Dir, Swat (Hindukush), and Chillas (Karakorum) regions from March 2022 to September, 2022 at full maturity stage (Figure 1). Samples were identified by expert taxonomists and with the help of Flora of Pakistan (Ali and Qaiser, 1993–2018), and voucher speciemen (CUHA-50), were submitted at COMSATS University herbarium in Abbottabad, Pakistan. Scientific and family names were confirmed by "World flora online".

Fresh leaves were cleaned with distilled water, and kept in shade at room temperature. After drying, leaves were grinded into fine powder and stored at 4 °C in refrigerator before extraction. The finely ground powdered material was extracted according to previously established protocol (Abbasi et al., 2015). Briefly, 0.5g of powdered sample was mixed with 10 mL methanol (99.8%), and kept at room temperature for 24 hrs. Afterwards, samples were centrifugated (at 8000 rpm.), and supernatants were combined in labeled flasks. All extracts were stored at 4 °C in refrigerator for subsequent analysis.

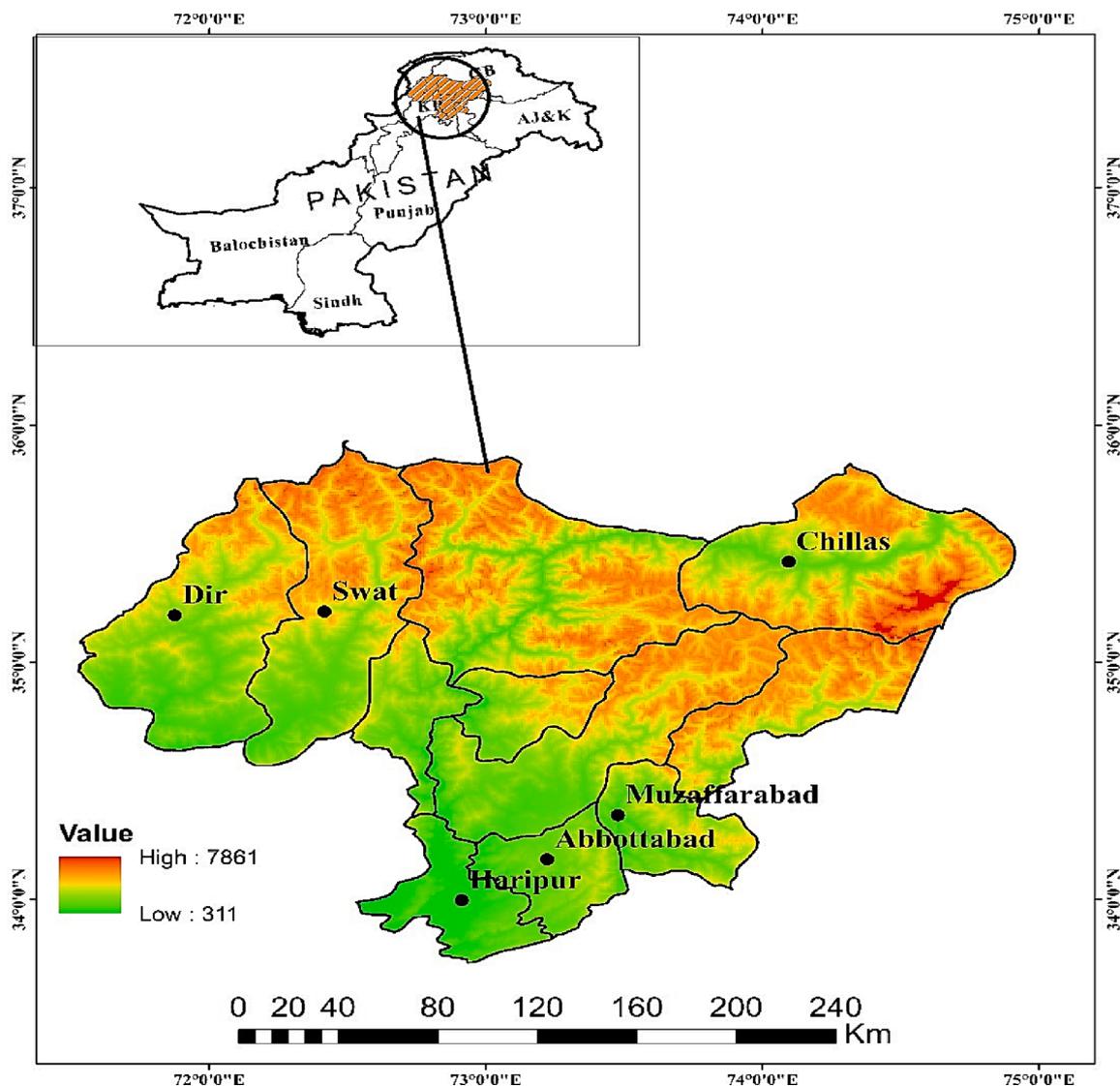


Figure 1. Map indicating collection sites of *N. officinale*

2.2. Quantification of polyphenolics

Determination of total phenolic and total flavonoids contents and was carried out as reported earlier by Zhang et al. (2011), using gallic acid and rutin as standards, respectively. Final values of total phenolic were reported as mg GAE/100g, and that of total flavonoids as mg RutE/100g based on dry weight. Standards curves of gallic acid (GA) and rutin (Rut) are shown in Figure S1 (A & B).

Profiling of phenolic acids and flavonoid compounds by modified method as explained earlier (Abbasi et al., 2015). In HPLC system (UV detector Cecil, UK, and C18 column Waters, USA.), 0.1% formic acid solution was used as mobile phase A, and methanol as mobile phase B at a flow rate of 1 mL/min. Polyphenolics were detected at 280 nm and quantified using the peak area of sample compared to calibration curve of standards, and final values were expressed as $\mu\text{g/g}$.

2.3. DPPH assay

The scavenging assay of DPPH radical was performed according to the method of He et al., (2018). Briefly, 0.1mM solution of DPPH (2 mL) was mixed with equal volume of plant extract, and incubated in dark for 30 min. Absorbance was measured at 517 nm using UV-spectrophotometer, and ascorbic acid was used as positive control. The percentage inhibition was calculated using the equation:

$$\text{DPPH}(\%) = [(As - Ab/Ac) \times 100] \quad (1)$$

Where As indicates sample absorbance, Ab shows absorbance of blank (methanol), and Ac indicates control absorbance.

2.4. Ferric ion reduction assay

The reduction of ferric ions was estimated using the method of Hazra et al. (2008). Shortly, plant extract, potassium ferricyanides (0.1%), and phosphate buffer (0.2 M, pH 6.6), were mixed in equal volume (2 mL of each). This mixture was incubated for 20 min bath at 50°C. Then 2 mL of trichloroacetic acid (10%) was added, followed by the addition of distilled water and 0.01% ferric chloride. Absorbance was measured at 700 nm. Gallic acid was used as standard with different concentrations of 20, 40, 60, 100, 140, 180 $\mu\text{g/mL}$ (Figure S1 C). Final values were expressed mg GAE/100g.

2.5. Hydrogen peroxide scavenging activity

Previously described method of Yahaya et al., (2021), was used to estimate H_2O_2 radical inhibition potential. Precisely, 2mL plant extract and same amount of H_2O_2 (mixture in phosphate buffer pH 7.4) were mixed. This mixture was kept in water bath for 10 min at 25°C, before taking absorbance at 230 nm. Phosphate buffer without H_2O_2 was used as blank. Percentage scavenging of H_2O_2 radical was determined using the equation:

$$\text{H}_2\text{O}_2(\%) = [(As - Ab/Ac) \times 100] \quad (2)$$

Where As indicates sample absorbance, Ab shows absorbance of blank (methanol), and Ac indicates control absorbance.

2.6. Phosphomolybdenum complex assay

The scavenging of molybdate ion was determined by performing Phosphomolybdenum complex antioxidant assay (PMCA) following the method as explained earlier (Prieto et al., 1999). Shortly, leaf extract (2 mL) was blended with 6.6 mL of reagent mixture containing sodium phosphate, ammonium molybdate and sulphuric acid. Reagent and extract mixture was kept in water bath for 90 min at 85°C. Methanol was used as blank, and absorbance was taken at 695 nm. Different concentrations of ascorbic acid (20 to 180 $\mu\text{g/mL}$) were used in Figure S1 (D) to calculate molybdate ions scavenging and results were presented as

mgAAE/100g on dry weight base.

2.7. Climate data

Data on climate attributes viz. latitude, longitude, and elevation of each sampling site was taken with the help of Global Positioning System (GPS). While temperature, rainfall, and relative humidity data of each location were taken from "Pakistan meteorological department (PMD)."

2.8. Statistical analysis

Data were analyzed using Microsoft Excel 2019; One-way ANOVA and correlation analysis were performed with SPSS-13.0 v. For graphical presentation Sigma Plot 12.1, and OriginPro-2023 were used.

3. Results and discussion

3.1. Variation in total phenolic and flavonoids contents in *N. officinale*

As illustrated in Figure 2A, *N. officinale* leaves collected from Abbottabad relatively had the highest concentration of total phenolic content (331.43 ± 41.43 mg GAE/100g), followed by Haripur and Swat samples (288.27 ± 26.06 , and 65.25 ± 4.14 mg GAE/100g, respectively) on dry weight basis with significant variations at $p \leq 0.05$. However, the

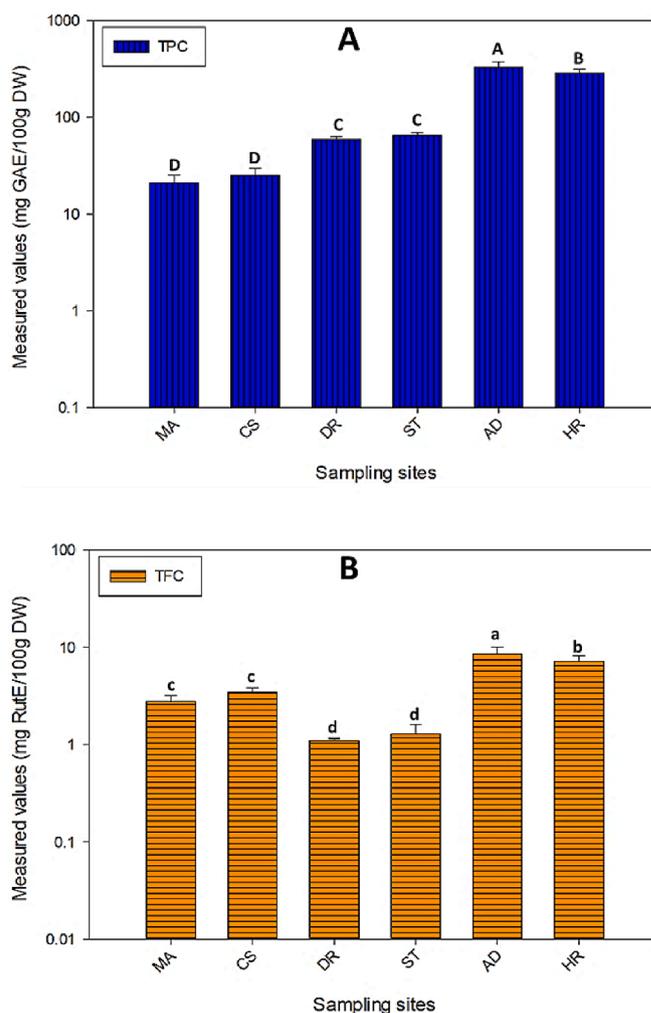


Figure 2. Comparison of TPC (2A) and TFC (2B) in *N. officinale* collected from different locations. Different letters (A-D/a-d) indicate variation in data at $p \leq 0.05$. MA. Muzaffarabad, CS. Chilas, DR. Dir, ST. Swat, AD. Abbottabad, HR. Haripur

lowest concentration of total phenolic content was in the samples collected from Muzaffarabad (21.00 ± 4.34 mg GAE/100g), which was 68% less than from Abbottabad sample. Because of strong bioactive potential against free radicals, inflammation, mutagenesis and cancer, flavonoids are most preferable phytochemicals in nutraceuticals, pharmaceuticals, and cosmetic industries (Karakaya et al., 2020). Measured levels of total flavonoids content (Figure 2B), were significantly higher in *N. officinale* leaves collected from Abbottabad and Haripur (8.55 ± 1.53 and 7.18 ± 0.98 mg RutE/100g, respectively). Total flavonoids in studied samples were relatively lower than reported previously from Iraq (Faizy et al., 2021). However, TPC in *N. officinale* reported from Iran (Mazandarani et al., 2013) were lower than our samples.

It is worth noting that phytochemical components in plants can vary significantly depending on environmental conditions, such as drought, rainfall patterns, temperature changes, and soil characteristics (Prabhu et al., 2021). Therefore, variations in TPC and TFC levels in *N. officinale* could be attributed to factors like different cultivars, geographical origins, genetic variations in the plant material, as well as analytical methods used. For instance, Prabhu et al. (2021) reported that concentration of phytochemicals may vary in the members of same species, because of their origin, and environmental conditions such as sunlight exposure, temperature, soil composition, and altitude. Additionally, plant's age, genetic variability, and agricultural practices could also contribute to the observed variations in phenolic and flavonoids contents. Likewise, in Pakistan *N. officinale* grows naturally in fresh water at different environmental conditions. Therefore, variation in water temperature, pH, concentration of dissolved salts and nutrients, composition of hydric soil and rainfall pattern, may effect the production route of polyphenolics (Amelung et al., 1999). Likewise, variation in total phenolic and flavonoids contents in the leaves of *N. officinale* collected from different areas of Pakistan and those reported from Iraq and Iran (Faizy et al., 2021; Mazandarani et al., 2013) might be due to difference in the geo-climatic conditions, water properties, genetic variations and analytical techniques used. Such as instrumentation, standards and laboratory conditions may affect concentration of phytochemicals and bioactive potential of the same species.

3.2. Distribution of phenolic and flavonoid compounds in *N. officinale*

Concentration of phenolic acids and flavonoids in the leaves of *N. officinale* collected from various location is given in Table 1. The overall increasing trend of phenolic acids and flavonoids at all locations was: RUT \geq p-CA \geq GA \geq FA \geq QE \geq CA \geq KEM. The samples collected from Dir had the highest levels of phenolic acids and flavonoids, while the lowest levels were found in samples of Chilas. Measured levels of FA, p-CA, KEM and RUT were significantly high ($p \leq 0.05$) in the leaves of *N. officinale* collected from Dir, followed by Abbottabad and Haripur. However, the increasing order of CA was: Dir \geq Swat \geq Haripur \geq Abbottabad \geq Muzaffarabad \geq Chilas. Gallic acid concentration was maximum at Haripur, and QE was the highest in samples collected from Dir. The differential concentrations of phenolic acids and flavonoids among the samples may attributed to the different structures of these compounds (Lukas et al., 2021). The structure of phenolic acids is

known to influence their activity and involvement in plant environmental responses. Therefore, the variations in the levels of these compounds in the *N. officinale* are likely influenced by their structural differences (Samec et al., 2011).

It was observed that the concentrations of certain compounds were relatively lower or higher in *N. officinale* compared to the reported values for other plant species. Among the phenolic acids, p-CA depicted the highest mean concentration (13.03 ± 5.32 μ g/g) at all locations, followed by GA, FA and CA (12.79 ± 5.22 , 7.09 ± 2.89 , and 1.80 ± 0.73 μ g/g, respectively). Comparatively, average concentration of CA in *N. officinale* (1.80 ± 0.73 μ g/g DW), collected from different locations was relatively lower than reported in *Brassica juncea* from China (Fang et al., 2008), which belongs to the same family "Brassicaceae". Likewise, measured levels of ferulic acid and p-coumaric acid were less than *B. rapa* ssp. *pekinensis* reported from Korea (Seong et al., 2016). Mean concentration of GA was 12.79 ± 5.22 μ g/g, and the highest levels were at Haripur, Dir and Abbottabad locations. However, these values were relatively lower than reported previously in *N. officinale* from Poland (Klimek-Szczykutowicz et al., 2020). Whereas, in the case of flavonoids, the highest average concentration was calculated for RUT (121.32 ± 49.53 μ g/g), followed by QE and KEM (6.29 ± 2.57 and 0.78 ± 0.32 μ g/g) on dry weight basis (Table 1). The contents of flavonoids were relatively high in *N. officinale* samples collected from Dir, while lowest at Chilas. Comparatively, estimated levels of QE, RUT and KEM were higher than reported earlier in the leaves of *N. officinale* and some other medicinal and food taxa of Brassicaceae from Iran and Iraq (Arabi et al., 2018; Al-Mashea et al., 2018). It is well established that concentrations of secondary metabolites can vary due to interactions and responses of plants to environmental conditions, genotypic variations among species and varieties, and even individual differences within the same species (Prabhu et al., 2021). Additionally, sunlight duration, temperature, nutrients and water availability, toxins, seasonal variations, pests, pathogens and herbivory influence on the concentration and synthesis of polyphenolics (Bibi et al., 2022). However, the fundamental molecular mechanisms are still not clear enough.

3.3. Free radicals' scavenging capacity in *N. officinale*.

These antioxidant molecules function as reductants inside the plant, by denoting hydrogen atoms or electrons to scavenge free radicals effectively (Ahmad et al., 2021). Therefore, ability of plants to produce antioxidants is a key factor in their capacity to combat the oxidative stress and damage to cells and tissues due to free radicals. Results showing scavenging potential of DPPH and H₂O₂ radicals in the leaves of *N. officinale* are given in Figure 3. Overall, the percentage inhibition of DPPH varies from 24.81% to 69.57% across different locations. Comparatively, samples collected from Chilas exhibits the highest DPPH scavenging capacity ($69.57 \pm 3.39\%$), followed by Swat and Muzaffarabad. Conversely, *N. officinale* leaves from Haripur exhibited the lowest potential to inhibit DPPH radical. And the percentage variation in the DPPH activity in samples collected from Chilas and Haripur was about 36%. The H₂O₂ scavenging varies significantly among the locations, with Chilas showing the highest percentage inhibition ($57.24 \pm 8.05\%$)

Table 1
Comparative assessment of phenolic acids and flavonoids in *N. officinale*

Localities	Phenolic acids (ug/g DW)				Flavonoids (ug/g DW)		
	CA	FA	p-CA	GA	QE	KEM	RUT
Muzaffarabad	1.65 ± 0.06^c	5.46 ± 0.15^f	12.60 ± 0.06^d	12.86 ± 0.04^c	6.90 ± 0.06^b	0.82 ± 0.11^b	113.14 ± 0.50^c
Chilas	1.47 ± 0.03^d	6.94 ± 0.03^d	10.85 ± 0.03^f	11.25 ± 0.02^e	3.10 ± 0.06^e	0.56 ± 0.05^c	113.47 ± 0.34^e
Dir	2.06 ± 0.08^a	8.89 ± 0.08^a	15.24 ± 0.12^a	13.54 ± 0.09^a	8.80 ± 0.10^a	0.95 ± 0.08^a	132.46 ± 0.62^a
Swat	2.10 ± 0.06^a	6.72 ± 0.03^e	11.24 ± 0.05^e	12.24 ± 0.12^d	6.34 ± 0.12^d	0.67 ± 0.12^c	119.57 ± 1.61^d
Abbottabad	1.71 ± 0.05^c	7.33 ± 0.05^b	14.71 ± 0.03^b	13.23 ± 0.05^b	6.24 ± 0.07^d	0.84 ± 0.06^{ab}	126.52 ± 0.08^b
Haripur	1.83 ± 0.04^b	7.18 ± 0.03^c	13.55 ± 0.05^c	13.64 ± 0.09^a	6.39 ± 0.06^c	0.83 ± 0.07^{ab}	122.80 ± 0.63^c

CA. Caffeic acid, FA. Ferulic acid, PCA. P-coumaric acid, GA. Gallic acid, QE. Quercetin, KEM. Kaempferol, RUT. Rutin

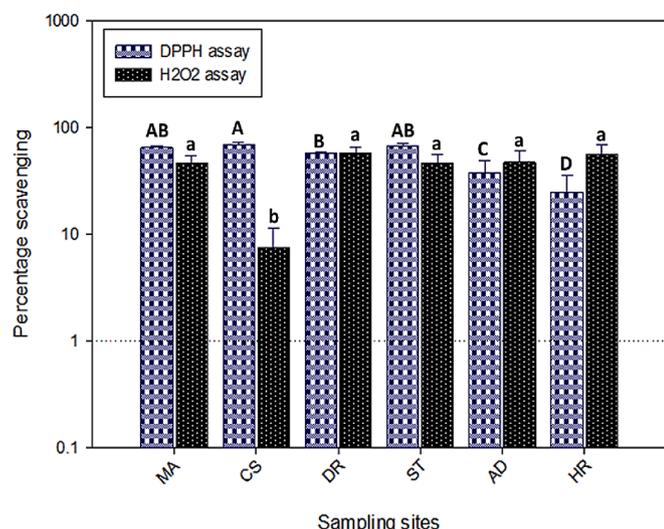


Figure 3. DPPH and H₂O₂ radicals scavenging potential in *N. officinale* collected from different locations. Different letters (A-D/a-b) indicate variation in data at $p \leq 0.05$. MA. Muzaffarabad, CS. Chilas, DR. Dir, ST. Swat, AD. Abbottabad, HR. Haripur

and Dir with minimum ($7.42 \pm 3.92\%$). However, the variations in H₂O₂ scavenging potential between the locations, except for Chilas, were not statistically significant ($p \geq 0.05$).

Relatively, percentage inhibition of DPPH and H₂O₂ radicals in the leaves *N. officinale* was higher than reported in the leaves of *Brassica rapa* from northern Portugal (Fernandes et al., 2016), *Brassica oleracea* var. *capitata* and *Brassica rapa* var. *pekinensis* from Czech Republic (Samec et al., 2011), and in other species of *Brassica* genus of family Brassicaceae cultivated in Spain (Soengas et al., 2012). The observed variations in free radical scavenging activity in the leaves of *N. officinale* collected from different locations could be attributed to several factors. One primary factor is the presence of bioactive molecules within the plant. The higher DPPH and H₂O₂ scavenging capacities in watercress leaves could be linked to the abundance of reductants that efficiently donate hydrogen atoms or electrons to neutralize free radicals.

The reduction of ferric ion (Fe⁺³) and molybdate ion in the leaves of

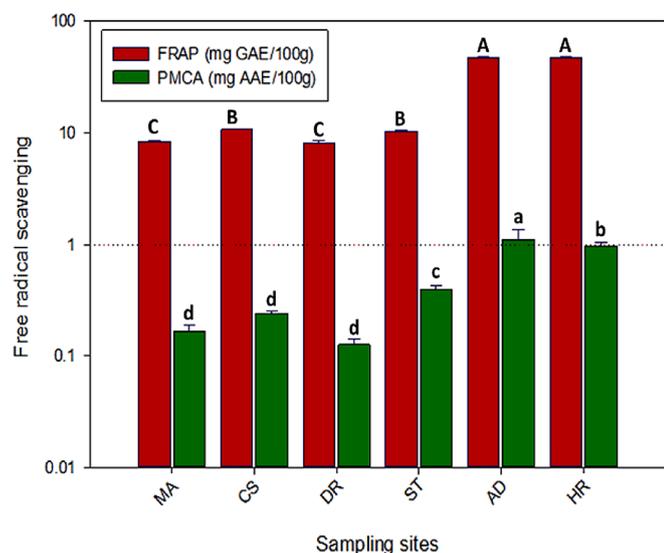


Figure 4. Comparison of ferric and molybdate ions reducing potential in *N. officinale* collected from different locations. Different letters (A-C/a-d) indicate variation in data at $p \leq 0.05$. MA. Muzaffarabad, CS. Chilas, DR. Dir, ST. Swat, AD. Abbottabad, HR. Haripur.

N. officinale as illustrated in Figure 4, provides insights into the antioxidant potential of watercress from different geographic areas. Among these, Abbottabad and Haripur exhibit the highest reduction of both ferric and molybdate ions, indicating potent antioxidant activity in the samples. The ferric ion reduction capacity follows an increasing order of Abbottabad \geq Haripur \geq Chilas \geq Swat \geq Muzaffarabad \geq Dir. Similarly, the inhibition of molybdate ions was maximum in samples collected from Abbottabad and Haripur, with significant variations compared to Muzaffarabad, Chilas, and Dir. *N. officinale* collected from Abbottabad and Haripur depicted maximum scavenging of ferric ions (46.57 ± 1.35 and 46.55 ± 1.21 mg GAE/100g DW, respectively). When compared to other species of the same family, *N. officinale* demonstrates higher ferric ion reducing potential than white and Chinese cabbage (*Brassica oleracea* var. *capitata* and *Brassica rapa* var. *pekinensis*) grown in the Czech Republic (Samec et al., 2011). This suggests that watercress could be a more potent source of antioxidants than some other common vegetables belong to same family Brassicaceae. Likewise, inhibition of molybdate ions was maximum in *N. officinale* collected from Abbottabad and Haripur (1.10 ± 0.20 and 0.96 ± 0.06 mg AAE/100g DW, respectively). These values varied significantly at $p \leq 0.05$, but there was no significant difference in the samples collected from Muzaffarabad, Chilas and Dir.

The differential profile of phytochemical compounds at various sites is responsible for the variations in antioxidant ability of *N. officinale* as well (Thiruvengadam et al., 2016). Because, significant variations were observed in the bioactive potential and phytochemical composition of *N. officinale* collected from different areas. This observation revealed that geo-environmental factors effect on the production of bioactive molecules responsible for antioxidant activity in the plant (Dastoor et al., 2017). Therefore, understanding the variations in antioxidant ability among different locations can guide the selection of optimal sources for nutraceutical applications. Our findings suggest that *N. officinale* possesses potent antioxidant properties compared to the other species within the same family. Therefore, *N. officinale*-derived bioactive molecules may have potential applications in nutraceutical, pharmaceutical, and cosmetic industries in combating oxidative stress and reducing the risk of chronic diseases associated with free radical species. Overall, this study contributes to the growing body of knowledge on the health benefits of *N. officinale* and paves the way for further research in the field of plant-based antioxidants and nutraceuticals.

3.4. Correlations between polyphenolics and antioxidant activities

The results, presented in Figure 5, reveal several significant associations that provide valuable insights into the role of polyphenolics in the antioxidant potential of plant species. For instance, in the current study highly significant ($p \leq 0.01$), positive correlations were noted between total phenolic contents in *N. officinale* leaves with ferric and molybdate ions at $r = 0.986$ and $r = 0.979$, respectively. Likewise, total flavonoids also depicted a highly significant ($p \leq 0.01$) direct relationship with ferric and molybdate ion reducing potential ($r = 0.952$ and $r = 0.927$, respectively). These findings confirm that phenolic compounds, and flavonoids contributes significantly to the antioxidant potential of food and medicinal plants (Karakaya et al., 2020). Interestingly, total phenolic and flavonoids contents exhibited highly significant inverse correlations with DPPH activity ($\approx 92\%$). This implies that higher levels of phenolic compounds and flavonoids may not contribute as effectively to the scavenging of DPPH radicals. In contrast to the DPPH scavenging, these compounds showed a weak positive association with hydrogen peroxide radical scavenging potential. This suggests that polyphenolics may play a role in neutralizing hydrogen peroxide radicals, albeit to a lesser extent compared to their role in reducing ferric and molybdate ions. The reasons behind this inverse relationship may be complex and warrant further investigation. However, it might be due to the synergistic role of different phytochemicals such as anthocyanins, carotenoids, vitamins, and antioxidant metals (Suleria et al., 2020).

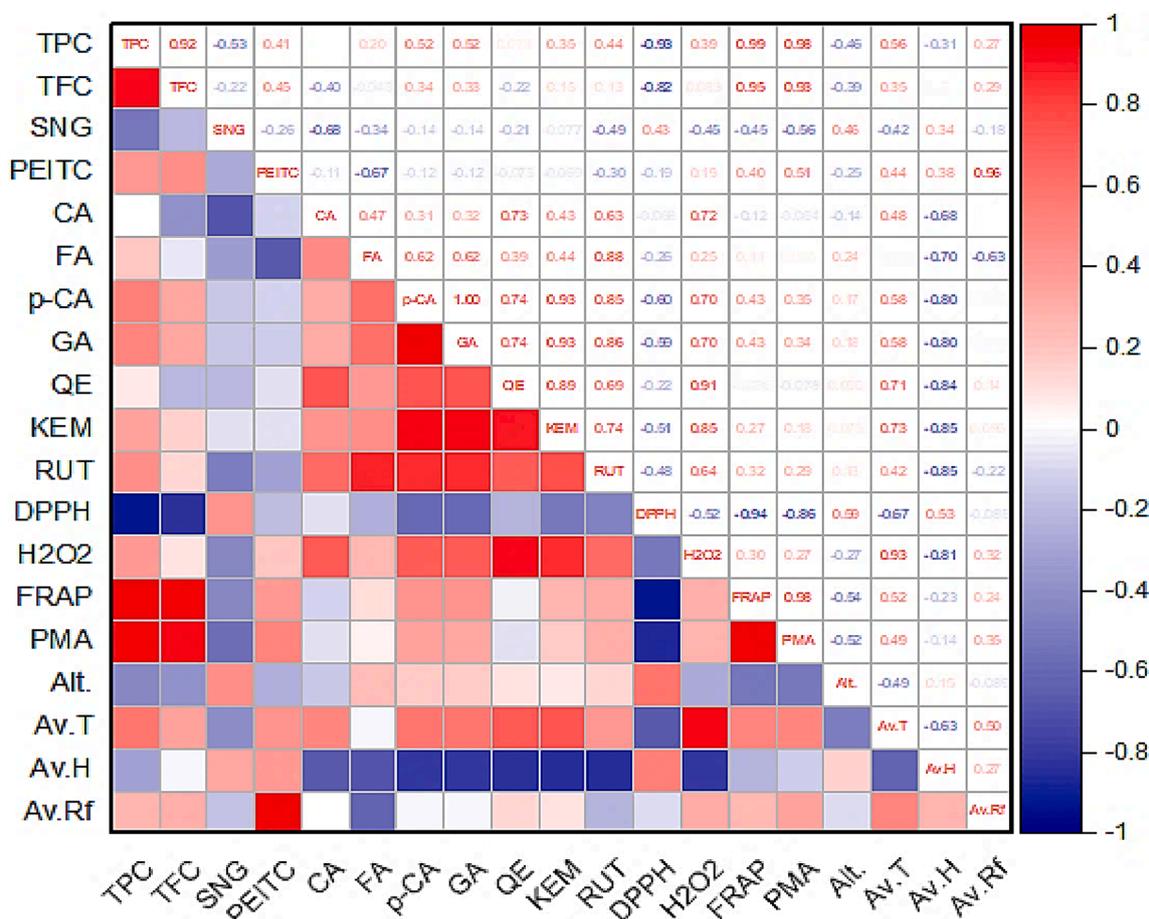


Figure 5. Correlation analysis of polyphenolics and antioxidant activities in *N. officinale* collected from different locations. Circles in boxes indicate significant associations ($p \leq 0.01$ and $p \leq 0.05$) among the variables.

The relationships between individual phenolic acids and flavonoid compounds with different antioxidant activities were also evaluated (Figure 5). Notably, GA, QE, and KEM showed significant ($p \leq 0.05$), positive interactions with inhibition of hydrogen peroxide in *N. officinale* leaves. Correspondingly, FA, p-CA, and RUT exhibited positive associations with inhibition of ferric ion, molybdate ion, and hydrogen peroxide radicals. However, these phenolic acids and flavonoid compounds showed negative relationships with DPPH scavenging potential. This suggests that certain phenolic compounds may be more effective in scavenging specific types of radicals, leading to varying relationships with different antioxidant assays.

Correlation analysis revealed that higher levels of the total phenolic content in the plant species are associated with increased ferric and molybdate ion reducing capacities, which are indicators of antioxidant potential. Similarly, total flavonoids also show highly significant positive correlations with ferric and molybdate ion reducing potential. Interestingly, the study finds highly significant inverse relationships between total phenolic, total flavonoids and DPPH radical scavenging activity. This inverse correlation implies that while higher levels of polyphenolic compounds positively influence ferric and molybdate ion reducing potential, they are associated with lower DPPH radical scavenging activity. Furthermore, it could indicate that the antioxidant mechanisms of polyphenolics in *N. officinale* may differ depending on the type of radical species involved.

3.5. Relationships of growing conditions with bioactive potential in *N. officinale*

Although, complex genetic pathways are involved in the synthesis of

various bioactive compounds in plants, but influence of environmental factors could not be underestimate [34]. Furthermore, understanding the impact of environmental factors on the production of secondary metabolites and their bioactive potential can have implications for both ecological and human health perspectives (Dastoor et al., 2017). Therefore, we have also investigated the influence of environmental factors, such as mean minimum and maximum temperature, humidity, rainfall, and altitude on polyphenolic content and antioxidant activities in *N. officinale*.

On the whole, mean minimum and maximum temperature depicted positive association with polyphenolics and antioxidant activities, except for FA content and DPPH radical scavenging activity (Figure 5) that showed inverse relations. The mean maximum temperature had highly significant ($p \leq 0.01$) positive correlation with GA (95.5%), and hydrogen peroxide scavenging potential (91.0%, $p \leq 0.05$) collected from different locations. This suggests that higher temperature may favor the production of polyphenolic compounds and enhances the antioxidant potential in watercress leaves. Our findings are compatible with previous reports (Moreira et al., 2020). However, according to Boussaa et al., (2020), antioxidant activities enhance in cold weather. For instance, other workers (Bibi et al., 2022; Wang, 2006) also reported inverse relationships between temperature and polyphenolics. Overall, rain fall usually has no significant association with polyphenolic synthesis in plants (Wang, 2006). Correspondingly, in the present study, humidity, rainfall, and altitude mostly exhibit negative or weak positive associations with polyphenolics and antioxidant activities in the leaves of *N. officinale*. This indicate that these environmental factors might have a less pronounced effect on the antioxidant capacity of this plant. These findings were similar to previous reports (Bibi et al., 2022; Bernal et al.,

2013). However, Boussaa et al. (2020), reported positive correlations between relative humidity and bioactive potential. Similarly, according to Bernal et al. (2013), concentration of phytochemicals and bioactive potential in medicinal plants decrease with increase in altitude. However, in some food and medicinal plants altitude exhibited significantly positive relationships with synthesis of secondary metabolites and antioxidant activities (Mpofu et al., 2006).

The correlation analysis in this study provides valuable insights into the interactions between polyphenolics and antioxidant potential in the leaves of *N. officinale* collected from different locations. The findings suggest that polyphenolic compounds, including total phenolics and flavonoids, play a significant role in the antioxidant activity of the plant species. However, their impact on different types of radical scavenging activities, such as DPPH and hydrogen peroxide, varies, indicating the complexity of antioxidant mechanisms in *N. officinale*. Additionally, certain phenolic acids and flavonoid compounds are found to have specific associations with different antioxidant activities. Furthermore, environmental factors, particularly temperature, appear to influence polyphenolic content and antioxidant activities in the plant.

3.6. Cluster and principal component analysis

The statistical techniques provide valuable insights into the grouping patterns and the contribution of specific metabolites to the overall antioxidant activity. The study employs clustering analysis (CA) and principal component analysis (PCA) to explore the relationships between polyphenolic contents and antioxidant potential in the leaves *N. officinale*. As illustrated in Figure 6, two main clusters were identified: the first group exhibits a significant association between samples collected from Abbottabad and Haripur. In contrast, the second cluster is subdivided into two groups, with Swat and Dir in one group, and Muzaffarabad and Chilas placed in the other. The clustering of samples may be attributed to the proximity of these sampling sites, which allows for the dispersion of *N. officinale* seeds through wind and running water. This possibility of seed dispersion among plants in close proximity can lead to genetic similarity and, consequently, similarity in polyphenolic profiles and antioxidant activities (Mpofu et al., 2006).

The PCA analysis, as shown in Figure 7, further explores the relations among polyphenolic compounds and antioxidant activities in

N. officinale. The data is grouped into three main components: PC1, PC2, and PC3. As shown in (Table S1), PC1 accounts for the highest percentage of variance (54.48%), followed by PC2 (30.19%) and PC3 (8.54%). In PC1, GA exhibits the maximum loading value (93.4%), indicating a strong positive association of this phenolic acid with other variables in this component. Similarly, *p*-CA, RUT, KEM, hydrogen peroxide scavenging activity, TPC, ferric ion reducing activity, and molybdate ion reducing activity also showed high loading values ($\geq 60\%$). This implies that these variables are positively associated with each other and collectively contribute to the antioxidant potential of *N. officinale*. The findings of PC1 align with the results of the correlation analysis, where TPC and TFC depicted highly significant positive relationships with reduction of ferric and molybdate ions. In PC2, QE, CA, and DPPH activity are grouped together, with QE and CA showing strong positive association ($\geq 60\%$), while DPPH exhibits negative relationships with all metabolites. PC3 primarily involves FA, which shows a loading value of 70.9%. Ferulic acid depicted negative relationships with antioxidant potential in *N. officinale* but has positive correlations with most of the phenolic acids and flavonoid compounds.

The cluster analysis and PCA results provide valuable insights into the diversity of polyphenolic profiles and antioxidant potential in *N. officinale* samples collected from different locations that suggests the dispersion of seeds as a possible contributing factor. Moreover, it reveals the key metabolites responsible for the observed differences in antioxidant activities and highlights their interrelationships. Understanding the factors that contribute to these variations can have implications for the selection and breeding of plants with enhanced antioxidant properties. Further research could focus on identifying the specific genetic and environmental factors that influence the production of polyphenolic compounds in *N. officinale* and exploring their potential applications in functional foods and natural antioxidants. Additionally, investigating the impact of different growing conditions on the antioxidant potential of *N. officinale* could provide valuable information for optimizing its cultivation and utilization as a nutraceutical resource.

4. Conclusions

The present study emphasizes the significance of *N. officinale* as a potential nutraceutical, and its applications in the food, pharmaceutical,

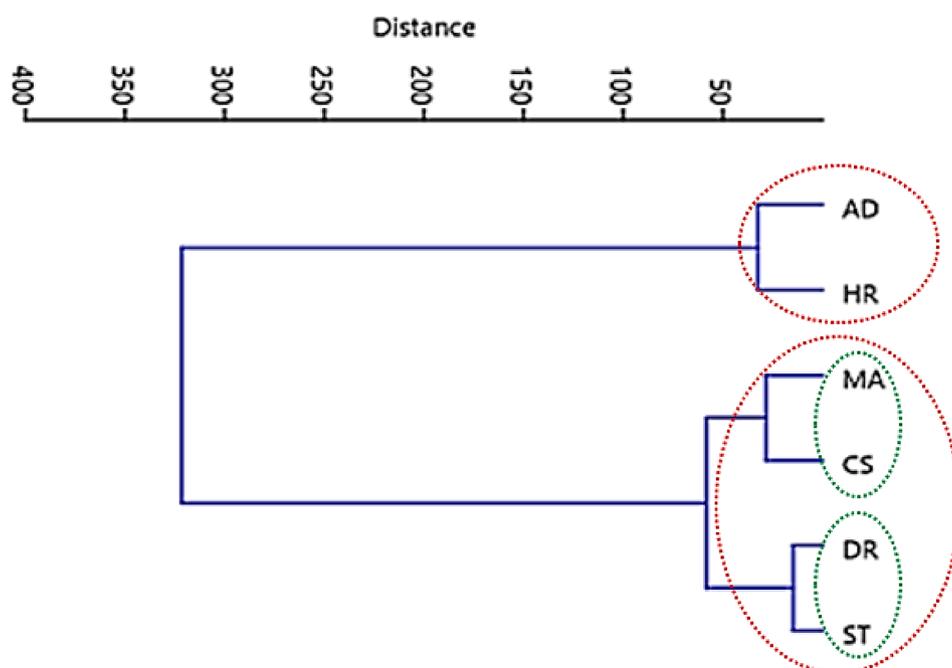


Figure 6. Cluster analysis indicating associations between sampling sites based on polyphenolic contents and antioxidant activities in *N. officinale*.

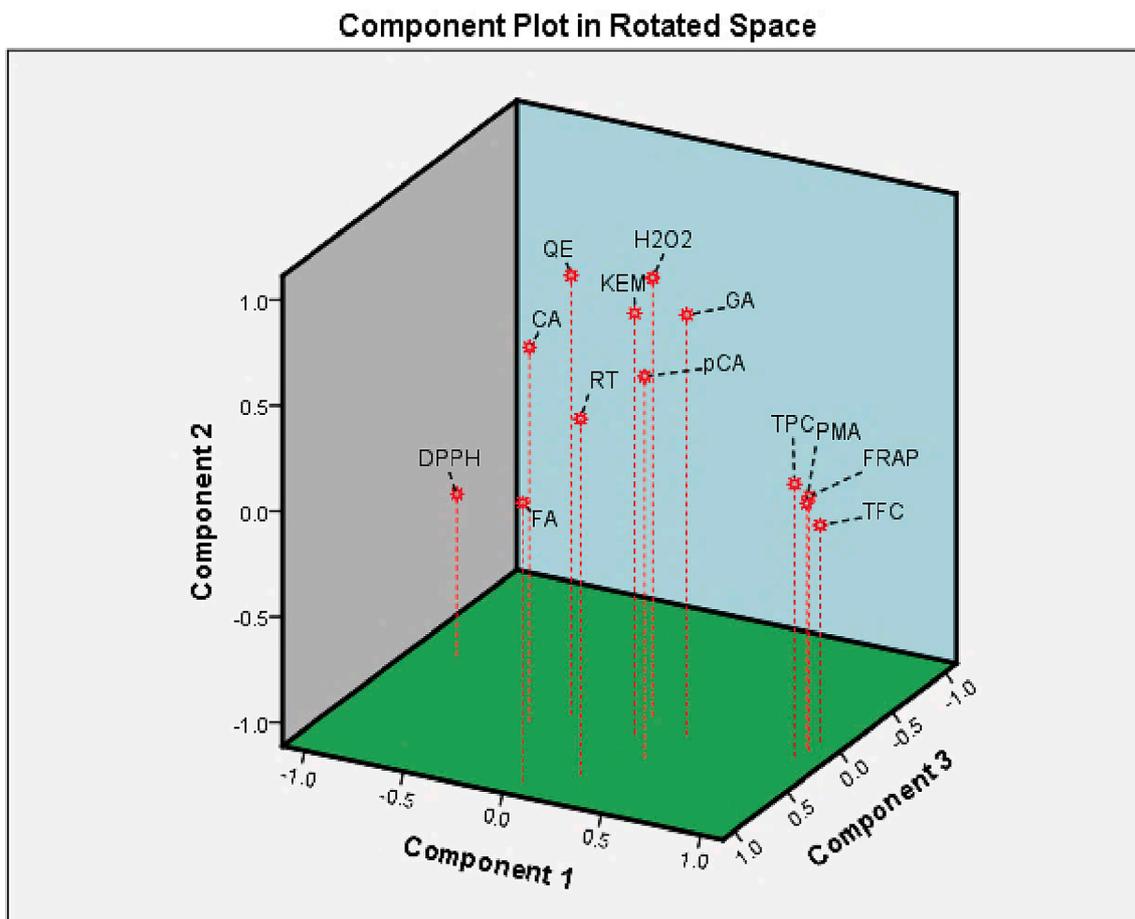


Figure 7. Principal component analysis of polyphenolics and antioxidant activities in *N. officinale* collected from diverse areas.

and cosmetic industries. Comparative assessment of polyphenolic content and antioxidant potential provides valuable insights into the variability of bioactivities in the leaves of this plant and highlights the importance of considering geographic variations. The correlation analysis reveals significant associations between polyphenolics and antioxidant activities in *N. officinale*. Although, Temperature influences the polyphenolic content and antioxidant activities in the leaves of *N. officinale*, but there was no significant impact of humidity, rainfall and altitude which indicates that these environmental factors might have a less effect on the antioxidant capacity of this plant. However, understanding the factors influencing on the synthesis of bioactive compounds in plants can have implications for agricultural, ecological, and human health perspectives. Overall, this study contributes to the understanding of the antioxidant potential of *N. officinale* and highlights the importance of considering both polyphenolic composition and environmental factors in future investigations related to its nutraceutical applications. Further research on the identification and quantification of the specific bioactive compounds in *N. officinale*, their mechanism of action and interactions could unveil the full potential of this plant as a functional food, and natural source of antioxidants with possible health benefits.

CRediT authorship contribution statement

Sobia Zaman: Data curation, Formal analysis, Writing – original draft. **Raza Ahmad:** Conceptualization, Methodology, Project administration, Validation, Writing – review & editing. **Manal Abdulaziz Binobeid:** Funding acquisition, Visualization, Writing – review & editing. **Mohamed Ragab Abdel Gawwad:** Funding acquisition,

Software, Visualization, Writing – review & editing. **Mohamed Soliman Elshikh:** Funding acquisition, Software, Visualization, Writing – review & editing. **Yusufjon Gafforov:** Methodology, Visualization, Writing – review & editing. **Arshad Mehmood Abbasi:** Conceptualization, Data curation, Funding acquisition, Methodology, Project administration, Resources, Software, Supervision, Writing – review & editing.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data Availability Statement

All data is provided in the manuscript; however, the corresponding author may be requested for any additional information.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jksus.2024.103223>.

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