



# Anticancer effects of pH-sensitive carvacrol zinc oxide quantum dots on DMBA induced mammary carcinoma in female sprague dawley rats

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## ABSTRACT

**Objectives:** The current research explores the anticancer effects of pH sensitive CVC-ZnO QDs (Carvacrol-loaded Zinc Oxide Quantum Dots) on DMBA induced mammary carcinoma in rats.

**Methods:** Female SD rats were used, and mammary cancer was induced by chemical carcinogen via subcutaneous injection near the mammary gland. Different concentrations of CVC-ZnO QDs were orally supplemented to evaluate the optimum dose. We assessed the growth rate, body weight changes, tumor volume, tumor incidence and tumor burden in both the inducer and treatment groups. We also evaluated the biochemical parameters (antioxidant status, lipid peroxidation, detoxification enzymes, and lipid profile) and histopathological changes in the kidney and mammary tissues.

**Results:** Our findings indicate that CVC-ZnO QDs treated rats significantly decreased the tumor weight, incidence, burden, lipid peroxidation levels, phase I detoxification enzyme activities and increased the body weight, phase II detoxification enzyme activities, and antioxidant status compared to the DMBA alone treated rats. CVC-ZnO QDs treatment also altered the lipid profile of plasma and mammary tissue. Furthermore, histopathological results confirmed that the CVC-ZnO QDs protect against DMBA-mediated damage to the mammary and kidney.

**Conclusion:** The findings indicate that the CVC-ZnO QDs administered at 4 mg/kg b.w exhibited a significant anticancer effect against DMBA-induced mammary cancer.

## 1. Introduction

Breast cancer begins in the breast tissue and is the most prevalent cancer impacting women globally. Based on the latest statistics, breast cancer represents around 30 % of newly diagnosed cancers among women (Cao et al., 2021). While there have been notable strides in diagnosing and treating breast cancer, it remains a pressing public health issue, underscoring the demand for innovative and efficient methods in both therapy and diagnosis (Anwar et al., 2020).

Quantum dots (QDs) represent nanocrystals possessing exceptional optical and electronic characteristics, rendering them highly prospective for various biomedical purposes such as cancer detection and treatment. QDs are composed of semiconductor materials that emit bright and

stable fluorescent signals when excited by light, making them ideal for imaging and sensing applications (He et al., 2019). Recent investigations have highlighted the potential of QDs for breast cancer therapy and imaging. For instance, QDs have been used for imaging breast cancer cells, which allows the detection of cancerous tissues with high accuracy and specificity (Rajitha et al., 2021). In addition, QDs have also been utilized to deliver therapeutic drugs to breast cancer cells, showing promising results in preclinical studies (Tagde et al., 2020). Edis et al. (2021) reported using QDs to deliver chemotherapeutic agents to mammary cancer cells, showing reduced toxicity and improved efficacy compared to conventional chemotherapy.

DMBA is a potent chemical carcinogen used to induce mammary tumors in animal models. The DMBA induced breast cancer model in

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female SD (Sprague Dawley) rats has been widely used to study the molecular and cellular mechanisms underlying the progression and development of mammary cancer (Kerdelhué et al., 2016). This ideal has additionally been employed to assess the effectiveness of different chemotherapy drugs and identify new therapeutic targets for treating breast cancer.

Carvacrol (CVC), a phenolic compound found in oregano, has been shown to possess anticancer properties (Li et al., 2021). Zinc oxide quantum dots (ZnO QDs), on the other hand, are nanomaterials that have been investigated for their potential use in chemotherapy because of their characteristics, such as optical attributes and a significant (Jiang et al., 2018).

From our initial investigation, it has been noted that nanoformulations containing CVC-ZnO QDs exhibit pH-responsive behavior, leading to their release specifically in the acidic pH conditions found within the tumor microenvironment. Consequently, this targeted drug delivery approach primarily directs therapeutic agents towards cancer cells while having a lesser impact on normal cells. The aim of this study is to explore the anticancer potential of pH-sensitive CVC-ZnO QDs in female SD rats with DMBA-induced mammary carcinoma. This investigation entails an examination of various parameters, including tumor growth rate, biochemical analysis in both plasma and mammary tissue, as well as histological alterations in the kidney and mammary tissue.

## 2. Materials and methods

### 2.1. Chemicals

Carvacrol and DMBA were acquired from Sigma Aldrich Chemical Pvt. Ltd., while all other chemicals utilized in this study were of analytical quality.

#### Synthesis of CVC-ZnO QDs

ZnO QDs were prepared from zinc acetate dihydrate and sodium hydroxide precursors. They were formed by mixing the precursors in anhydrous ethanol at 60 °C for 5 h, followed by centrifugation and dissolution in anhydrous ethanol. CVC was loaded onto the QDs during 12 h of continuous stirring in a colloidal suspension. The prepared CVC-ZnO QDs had spherical shape with an average crystal size of 7.64 nm.

### 2.2. Animal model

We purchased 36 female SD rats from the Biogen Laboratory Animal Facility in Bangalore, India, registered under CPCSEA (Reg No: 971/PO/RcBiBt/S/2006/CPCSEA). The rats were between 6 and 8 weeks old and weighed 100 g and 120 g. These rats were housed in the Central Animal House, RMMC (Rajah Muthiah Medical College), Annamalai University. Prior to initiating the experiment, we ensured compliance with the guidelines by securing the necessary approval from the Institutional Animal Ethics Committee for the Supervision and Administration of Experimental Animals (IAEC Proposal No: AU-IAEC/1321/6/22). The rats were acclimated to the laboratory conditions, which included controlled humidity (50 ± 10 %), a temperature of 24 ± 2°C and a 12 h (light/dark cycle). They were provided with stranded feed (Pellet diet-cereals, animal and vegetal proteins vitamins & minerals) and water throughout the study.

### 2.3. Induction mammary carcinoma and CVC-ZnO QDs preparation

Mammary carcinoma was induced in rats by administering a single dose of DMBA (25 mg/rat) through an emulsion of physiological saline (0.25 ml) and sunflower oil (0.75 ml). CVC-ZnO QDs were suspended in 0.1 % DMSO.

### 2.4. Experimental design

Thirty-six female SD rats were randomly divided into 6 groups, each

consisting of six. The experimental procedure conducted in this study is illustrated in Fig. 1. After a study duration of 13 weeks, all animals were euthanized through cervical dislocation. Several parameters were measured and calculated, including animal body weight, growth rate, tumor weight, and tumor characteristics. Blood was collected for biochemical analysis and subjected to centrifugal force for 15 min at 1000g to separate the plasma. Mammary tissue was removed from each rat, processed by homogenization with a suitable buffer, and then subjected to centrifugation. The resulting supernatant was utilized for biochemical analyses. Additionally, kidney and mammary tissues were preserved (10 % formalin) for histological studies.

### 2.5. Biochemical analysis

The biochemical analysis was performed in the control and experimental rat plasma and mammary tissue. The Table 1 shows the biochemical parameters and their methods.

### 2.6. Histopathological studies

The control and experimental rats kidney and mammary tissues were examined histopathologically. After fixation in 10 percent buffered formalin, the tissues were regularly treated, embedded in paraffin, and sliced into 2–3 mm sections using a rotary microtome. The resulting tissue slices were placed on glass slides and subjected to H&E (hematoxylin and eosin) staining.

### 2.7. Statistical analysis

The mean ± SD is used to represent experimental values. One-way ANOVA (analysis of variance) was used to compare the mean values across groups, and DMRT was used for multiple comparisons. And the significance was set at  $p \leq 0.05$ .

## 3. Results

### 3.1. Effect of CVC-ZnO QDs on growth rate, body weight, and tumor characteristics

Table 2 represents the growth rate, body weight, and tumor characteristics of the control and experimental rats. The body weight was assessed based on the differences observed in the initial and final stages between the control and experimental rats. Rats treated with DMBA alone exhibited a notable reduction in growth rate and body weight. Nevertheless, administering CVC-ZnO QDs orally at various doses resulted in a substantial enhancement of growth rate and body weight in cancer bringing rats (DMBA-induced). In contrast to the control group of rats, the group administered CVC-ZnO QDs showed no notable variances in their growth rate and body weight.

Tumor burden and volume in rats were typically evaluated by measuring the size of tumors using calipers and calculated the tumor volume using the formula  $V = 4/3\pi (D1/2) (D2/2) (D3/2)$ . In the DMBA alone-induced rats, 100 % tumor development was observed, with tumor burden (200.77 mm<sup>3</sup>) and tumor volume (33.46 mm<sup>3</sup>). Oral supplementation with CVC-ZnO QDs to cancer-induced rats markedly decreased tumor volume, burden and incidence. In control and CVC-ZnO QDs alone animals, no tumors were seen. Hence, among the 2, 4, and 8 mg/kg b.w doses, the activity of CVC-ZnO QDs at 4 mg/kg b.w had an impact in the DMBA-induced rats.

### 3.2. Effect of CVC-ZnO QDs on antioxidant status

The enzymatic and non-enzymatic antioxidant activities of control and experimental rats are shown in Tables 3 and 4. The presence of antioxidant in the plasma and mammary tissues of rats treated with DMBA alone was notably reduced when compared to the control group.

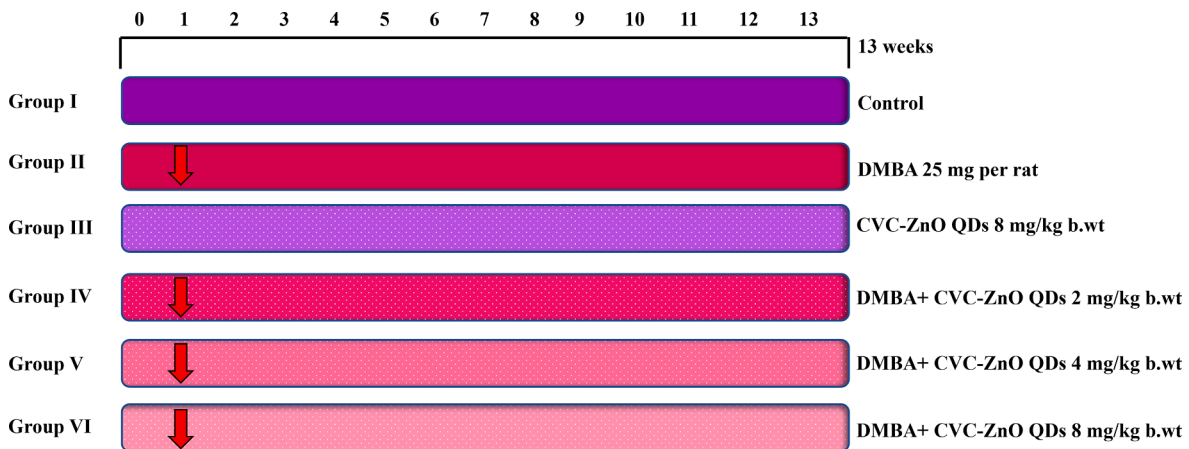


Fig. 1. Experimental design.

**Table 1**  
Biochemical parameters and its methods.

Biochemical parameters	Method
Antioxidant status	
SOD (superoxide dismutase)	Kakkar et al. (1984)
CAT (catalase)	Sinha (1972)
GPx (Glutathione peroxidase)	Rotruck et al. (1973)
GSH (reduced glutathione)	Beutler and Kelly (1963)
Vit-E (Vitamin-E)	Omaye et al. (1979)
Vit-C (Vitamin-C)	Palan et al. (1991)
Lipid peroxidation	
TBAR (Thiobarbituric acid reactive substances)	Yagi (1987) in plasma Ohkawa et al. (1979) in mammary tissue
LOOH (lipid hydroperoxide)	Jiang et al. (1992)
CD (Conjugated dienes)	Rao and Recknagel (1968)
Phase I enzymes	
CYP450 (cytochrome-p450)	Omura and Sato (1964)
Cyt-b5 (cytochrome b5)	Omura and Sato (1964)
Phase II enzymes	
GST (Glutathione s-transferase)	Habig et al. (1974)
GR (Glutathione reductase)	Carlberg and Mannervik (1985)
Lipid profile	
TC (Total cholesterol)	Zlatkis et al. (1953)
TG (Triglycerides)	Foster and Dunn (1973)
PL (Phospholipid)	Zilversmit and Davis (1950)
FFA (Free fatty acids)	Falholt et al. (1973)

The addition of CVC-ZnO QDs at various concentrations notably enhanced antioxidant activities in comparison to rats induced with DMBA alone. Particularly observed outcomes were evident in rats that

**Table 2**  
Effect of CVC-ZnO QDs on body weight, growth rate, tumor incidence, tumor volume and tumor burden of control and experimental rats.

Groups	Initial body weight (g)	Final body weight	Growth rate (%)	Total number of tumors (n)	Tumor weight (g)	Tumor incidence (%)	Tumor volume (mm <sup>3</sup> /rat)	Tumor burden (mm <sup>3</sup> /rat)
Control	111.92 ± 6.1	197.00 ± 6.3 <sup>a</sup>	76.01 <sup>a</sup>	0/6	-	-	-	-
DMBA	116.99 ± 3.1	157.90 ± 7.9 <sup>b</sup>	34.96 <sup>b</sup>	6/6	7.53	100	33.46 ± 3.44 <sup>a</sup>	200.77 ± 20.65 <sup>a</sup>
CVC-ZnO QDs (8 mg)	115.26 ± 4.7	205.40 ± 9.3 <sup>a</sup>	78.2 <sup>a</sup>	0/6	-	-	-	-
DMBA + CVC-ZnO QDs (2 mg)	109.40 ± 5.3	174.81 ± 7.6 <sup>c</sup>	59.78 <sup>c</sup>	3/6	6.35	50	15.75 ± 0.59 <sup>b</sup>	47.26 ± 1.79 <sup>b</sup>
DMBA + CVC-ZnO QDs (4 mg)	112.05 ± 4.2	186.62 ± 5.9 <sup>a</sup>	66.55 <sup>a</sup>	2/6	5.49	33.33	8.78 ± 0.5 <sup>c</sup>	17.57 ± 1.05 <sup>c</sup>
DMBA + CVC-ZnO QDs (8 mg)	112.80 ± 2.72	195.13 ± 4.05 <sup>a</sup>	72.98 <sup>a</sup>	0/6	-	-	-	-

Tumor volume was measured using the formula  $V = 4/3\pi (D1/2) (D2/2) (D3/2)$ , where D1, D2 and D3 are the three diameters (in mm) of the tumor; (n) indicates total number of rats bearing tumors. Tumor burden was calculated by multiplying the tumor volume and total number of tumors. Values are expressed as mean ± SD for six rats in each group. Values not sharing a common superscript (a,b,c) differ significantly at p < 0.05 (DMRT).

**Table 3**  
Effect of CVC-ZnO QDs on the plasma antioxidant status of control and experimental rats.

Groups	SOD U <sup>x</sup> /mL	CAT U <sup>y</sup> /mL	GPx U <sup>z</sup> /mL	GSH mg/dL	Vit C mg/dL	Vit E mg/dL
Control	5.23 ± 0.78 <sup>a</sup>	3.12 ± 0.41 <sup>a</sup>	57.67 ± 3.34 <sup>a</sup>	18.42 ± 1.03 <sup>a</sup>	6.67 ± 0.92 <sup>a</sup>	4.84 ± 0.84 <sup>a</sup>
DMBA	2.73 ± 0.56 <sup>b</sup>	1.61 ± 0.24 <sup>b</sup>	28.69 ± 2.71 <sup>b</sup>	9.87 ± 0.82 <sup>b</sup>	2.52 ± 0.76 <sup>b</sup>	1.67 ± 0.61 <sup>b</sup>
CVC-ZnO QDs (8 mg)	5.29 ± 0.71 <sup>a</sup>	3.18 ± 0.38 <sup>a</sup>	57.71 ± 2.43 <sup>a</sup>	18.47 ± 0.75 <sup>a</sup>	6.72 ± 0.43 <sup>a</sup>	4.88 ± 0.72 <sup>a</sup>
DMBA + CVC-ZnO QDs (2 mg)	4.68 ± 0.57 <sup>c</sup>	1.99 ± 0.27 <sup>c</sup>	41.67 ± 3.52 <sup>c</sup>	14.45 ± 0.94 <sup>c</sup>	5.43 ± 0.84 <sup>c</sup>	3.47 ± 0.42 <sup>c</sup>
DMBA + CVC-ZnO QDs (4 mg)	5.02 ± 0.42 <sup>a</sup>	2.96 ± 0.24 <sup>a</sup>	56.28 ± 2.84 <sup>a</sup>	17.99 ± 0.72 <sup>a</sup>	6.26 ± 0.64 <sup>a</sup>	4.55 ± 0.75 <sup>a</sup>
DMBA + CVC-ZnO QDs (8 mg)	5.21 ± 0.28 <sup>a</sup>	3.09 ± 0.26 <sup>a</sup>	57.64 ± 2.76 <sup>a</sup>	18.39 ± 0.93 <sup>a</sup>	6.64 ± 0.52 <sup>a</sup>	4.81 ± 0.48 <sup>a</sup>

Data are expressed as the mean ± SD for six rats in each group. Values not sharing a common superscript (a,b,c) differ significantly at p < 0.05 (DMRT). Units for SOD<sup>x</sup>, CAT<sup>y</sup> and GPX<sup>z</sup> are expressed as the amount of enzyme required to inhibit 50 % of NBT reduction, micromoles of H<sub>2</sub>O<sub>2</sub> utilized/second, and micromoles of glutathione utilized/minute, respectively.

**Table 4**

Effect of CVC-ZnO QDs on antioxidant status in the mammary tissues of control and experimental rats.

Groups	SOD U <sup>X</sup> / mg of protein	CAT U <sup>Y</sup> / mg of protein	GPx U <sup>Z</sup> / mg of protein	GSH mg/ 100 g wet tissue	Vit C mg/ 100 g wet tissue	Vit E mg/ 100 g wet tissue
Control	13.65 ± 0.78 <sup>a</sup>	61.04 ± 2.45 <sup>a</sup>	9.65 ± 0.52 <sup>a</sup>	11.86 ± 2.63 <sup>a</sup>	6.42 ± 0.34 <sup>a</sup>	5.63 ± 0.24 <sup>a</sup>
DMBA	7.82 ± 0.63 <sup>b</sup>	38.95 ± 2.84 <sup>b</sup>	4.81 ± 0.49 <sup>b</sup>	5.47 ± 1.55 <sup>b</sup>	2.13 ± 0.29 <sup>b</sup>	1.94 ± 0.26 <sup>b</sup>
CVC-ZnO QDs (8 mg)	13.71 ± 0.71 <sup>a</sup>	61.08 ± 3.10 <sup>a</sup>	9.68 ± 0.53 <sup>a</sup>	11.89 ± 2.61 <sup>a</sup>	6.46 ± 0.32 <sup>a</sup>	5.67 ± 0.30 <sup>a</sup>
DMBA + CVC- ZnO QDs (2 mg)	10.64 ± 0.62 <sup>c</sup>	49.38 ± 2.78 <sup>c</sup>	7.24 ± 0.48 <sup>c</sup>	9.85 ± 2.01 <sup>c</sup>	5.68 ± 0.33 <sup>c</sup>	4.86 ± 0.23 <sup>c</sup>
DMBA + CVC- ZnO QDs (4 mg)	13.23 ± 0.64 <sup>a</sup>	59.06 ± 2.64 <sup>a</sup>	9.37 ± 0.56 <sup>a</sup>	11.54 ± 2.42 <sup>a</sup>	6.09 ± 0.27 <sup>a</sup>	5.28 ± 0.27 <sup>a</sup>
DMBA + CVC- ZnO QDs (8 mg)	13.62 ± 0.66 <sup>a</sup>	60.99 ± 2.45 <sup>a</sup>	9.61 ± 0.55 <sup>a</sup>	11.79 ± 2.46 <sup>a</sup>	6.39 ± 0.29 <sup>a</sup>	5.61 ± 0.21 <sup>a</sup>

Data are expressed as the mean ± SD for six rats in each group. Values not sharing a common superscript (a,b,c) differ significantly at  $p < 0.05$  (DMRT). Units for SOD<sup>X</sup>, CAT<sup>Y</sup> and GPX<sup>Z</sup> are expressed as the amount of enzyme required to inhibit 50 % of NBT reduction, micromoles of H<sub>2</sub>O<sub>2</sub> utilized/second, and micromoles of glutathione utilized/minute, respectively.

received supplementation of CVC-ZnO QDs at a dose of 4 mg/kg b.w.

### 3.3. Effect of CVC-ZnO QDs on lipid peroxidation

The lipid peroxidation levels in the plasma and mammary tissue of both control and experimental rats are detailed in Table 5. The findings suggest that the lipid peroxidation levels were significantly elevated in both the plasma and mammary tissues of rats subjected to DMBA treatment alone when compared to the control group. Oral supplementation of CVC-ZnO QDs to DMBA-treated rats significantly reduced lipid peroxidation levels. In comparison with control rats, CVC-ZnO QDs alone treated demonstrated no significant variation in lipid peroxidation within the plasma and mammary tissue.

### 3.4. Effect of CVC-ZnO QDs on detoxification enzymes

Table 6 displays the enzymatic detoxification activities observed in the mammary tissues of both control and experimental groups of rats. The rats treated with DMBA alone exhibited significantly higher phase I enzyme activity than the control rats. Supplementation with CVC-ZnO QDs reduced DMBA-metabolizing phase I enzyme activities. Compared to the control rats the rats treated with DMBA alone showed a notable decrease in the enzyme activities of phase II. Conversely, treatment through CVC-ZnO QDs significantly improved the phase II enzymes activities in the DMBA-treated rats. Notably, the medium dose of CVC-ZnO QDs at 4 mg/kg b.w demonstrated a more pronounced effect than the other two.

### 3.5. Effect of CVC-ZnO QDs on lipid profile

Table 7 represents the lipid profile of the control and experimental rats in the plasma and mammary tissues. In the DMBA- alone induced rats, concentrations of PL, FFA, TC, and TG were significantly higher in the plasma compared to the control rats. However, in mammary tissues

**Table 5**

Effect of CVC-ZnO QDs on lipid peroxidation in the plasma and mammary tissues of control and experimental rats.

Groups	Plasma			Mammary tissue		
	TBARs (n mol/ ml)	LOOH (mmol/ ml)	CD (µg/ ml)	TBARs (n mol/mg protein)	LOOH (mmol/ mg protein)	CD (mg/ g protein)
Control	2.56 ± 0.24 <sup>a</sup>	2.57 ± 0.22 <sup>a</sup>	0.94 ± 0.11 <sup>a</sup>	1.92 ± 0.13 <sup>a</sup>	0.96 ± 0.11 <sup>a</sup>	1.67 ± 0.09 <sup>a</sup>
DMBA	5.32 ± 0.22 <sup>b</sup>	4.41 ± 0.28 <sup>b</sup>	2.46 ± 0.18 <sup>b</sup>	3.44 ± 0.11 <sup>b</sup>	1.84 ± 0.14 <sup>b</sup>	2.94 ± 0.11 <sup>b</sup>
CVC- ZnO QDs (8 mg)	2.49 ± 0.27 <sup>a</sup>	2.51 ± 0.21 <sup>a</sup>	0.90 ± 0.12 <sup>a</sup>	1.89 ± 0.16 <sup>a</sup>	0.94 ± 0.14 <sup>a</sup>	1.69 ± 0.09 <sup>a</sup>
DMBA + CVC- ZnO QDs (2 mg)	3.51 ± 0.22 <sup>c</sup>	2.28 ± 0.22 <sup>c</sup>	1.32 ± 0.17 <sup>c</sup>	2.36 ± 0.16 <sup>c</sup>	1.40 ± 0.17 <sup>c</sup>	2.27 ± 0.12 <sup>c</sup>
DMBA + CVC- ZnO QDs (4 mg)	2.61 ± 0.21 <sup>a</sup>	2.73 ± 0.29 <sup>a</sup>	1.01 ± 0.12 <sup>a</sup>	2.09 ± 0.14 <sup>a</sup>	0.99 ± 0.11 <sup>a</sup>	1.84 ± 0.11 <sup>a</sup>
DMBA + CVC- ZnO QDs (8 mg)	2.60 ± 0.27 <sup>a</sup>	2.61 ± 0.27 <sup>a</sup>	0.97 ± 0.19 <sup>a</sup>	1.95 ± 0.16 <sup>a</sup>	0.98 ± 0.16 <sup>a</sup>	1.71 ± 0.14 <sup>a</sup>

Data are expressed as the mean ± SD for six rats in each group. Values not sharing a common superscript (a,b,c) differ significantly at  $p < 0.05$  (DMRT).

**Table 6**

Effect of CVC-ZnO QDs on biotransformation enzymes in the mammary tissues of control and experimental rats.

Groups	Cytochrome P450	Cytochrome b5	GST	GR
Control	0.96 ± 0.09 <sup>a</sup>	0.49 ± 0.03 <sup>a</sup>	1.68 ± 0.24 <sup>a</sup>	4.67 ± 1.65 <sup>a</sup>
DMBA	1.67 ± 0.16 <sup>b</sup>	0.98 ± 0.09 <sup>b</sup>	0.78 ± 0.08 <sup>b</sup>	1.57 ± 0.97 <sup>b</sup>
CVC-ZnO QDs (8 mg)	0.94 ± 0.07 <sup>a</sup>	0.48 ± 0.04 <sup>a</sup>	1.71 ± 0.26 <sup>a</sup>	4.77 ± 1.87 <sup>a</sup>
DMBA + CVC-ZnO QDs (2 mg)	1.29 ± 0.08 <sup>c</sup>	0.76 ± 0.03 <sup>c</sup>	0.98 ± 0.28 <sup>c</sup>	2.99 ± 1.34 <sup>c</sup>
DMBA + CVC-ZnO QDs (4 mg)	1.04 ± 0.08 <sup>a</sup>	0.58 ± 0.05 <sup>a</sup>	1.54 ± 0.27 <sup>a</sup>	3.86 ± 1.48 <sup>a</sup>
DMBA + CVC-ZnO QDs (8 mg)	0.98 ± 0.07 <sup>a</sup>	0.51 ± 0.04 <sup>a</sup>	1.64 ± 0.24 <sup>a</sup>	4.65 ± 1.64 <sup>a</sup>

Cytochrome P450 reductase—nmol/mg of microsomal protein, Cytochrome b5 reductase—nmol/mg of microsomal protein, Glutathione S-transferase—µmol of CDNB-GSH conjugate formed/mg microsomal protein/min, Glutathione reductase—µmol of NADPH oxidized/mg microsomal protein/min. Values are expressed as mean ± SD for six rats in each group. Values not sharing a common superscript (a,b,c) differ significantly at  $p < 0.05$  (DMRT).

of DMBA alone rats the PL and FFA levels were decreased and the TC and TG levels were significantly increased. CVC-ZnO QDs administration to DMBA-induced rats brought back the levels to near those of the control values in both the plasma and mammary tissues.

### 3.6. Histological changes

Figs. 2 and 3 shows the histological changes of kidney and mammary tissue of control and experimental rats. Histopathological examination

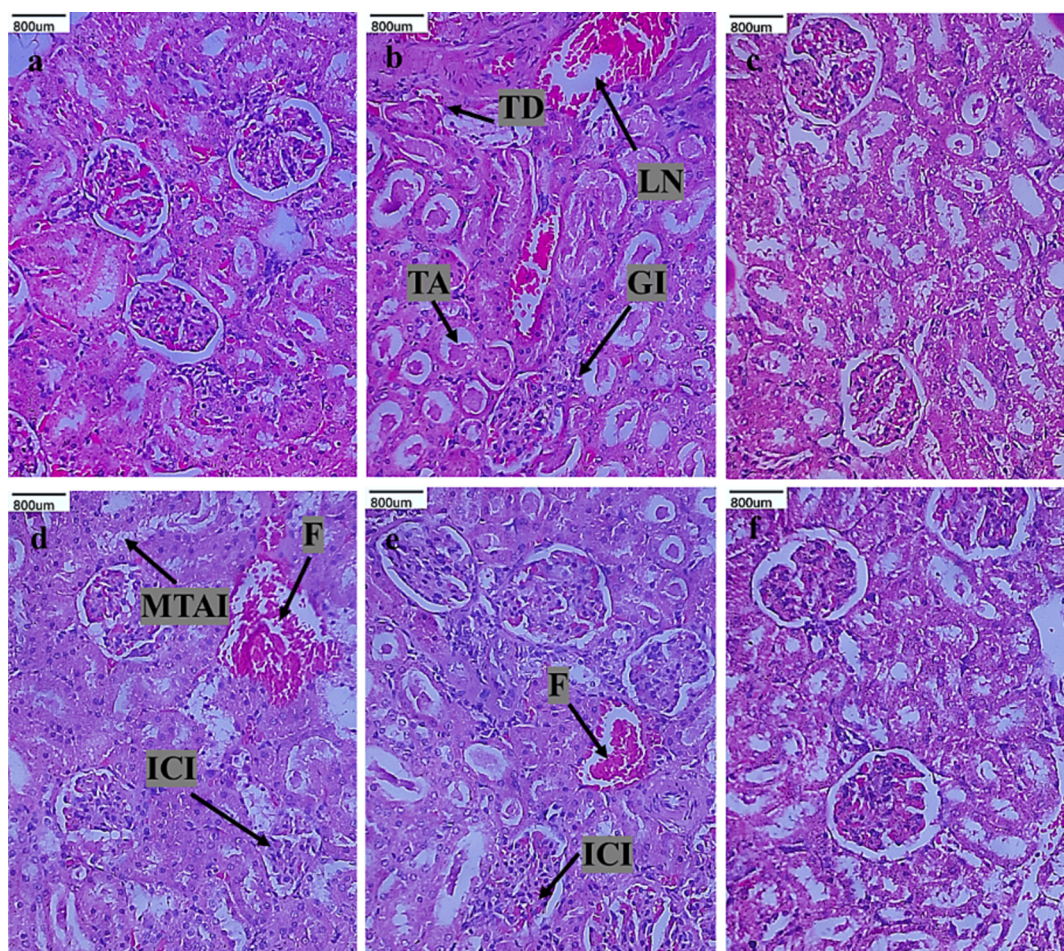


**Table 7**

Effect of CVC-ZnO QDs on the lipid profile in the plasma and mammary tissue of control and experimental rats.

Groups	Plasma (mg/dl)				Mammary tissue (mg/g)			
	TC	TG	PL	FFA	TC	TG	PL	FFA
Control	79.32 ± 6.57 <sup>a</sup>	75.67 ± 6.12 <sup>a</sup>	101.65 ± 9.3 <sup>a</sup>	12.47 ± 1.51 <sup>a</sup>	6.41 ± 0.42 <sup>a</sup>	5.49 ± 0.37 <sup>a</sup>	15.48 ± 0.98 <sup>a</sup>	10.26 ± 0.81 <sup>a</sup>
DMBA	120.41 ± 9.42 <sup>b</sup>	128.34 ± 10.04 <sup>b</sup>	147.68 ± 11.75 <sup>b</sup>	17.92 ± 1.82 <sup>b</sup>	15.74 ± 0.57 <sup>b</sup>	10.57 ± 0.49 <sup>b</sup>	9.82 ± 0.67 <sup>b</sup>	6.78 ± 0.44 <sup>b</sup>
CVC-ZnO QDs (8 mg)	78.75 ± 6.31 <sup>a</sup>	75.10 ± 5.87 <sup>a</sup>	100.97 ± 9.64 <sup>a</sup>	11.97 ± 1.47 <sup>a</sup>	6.37 ± 0.43 <sup>a</sup>	5.37 ± 0.41 <sup>a</sup>	15.73 ± 1.01 <sup>a</sup>	10.31 ± 0.92 <sup>a</sup>
DMBA + CVC-ZnO QDs (2 mg)	101.92 ± 7.57 <sup>c</sup>	108.75 ± 7.68 <sup>c</sup>	131.56 ± 10.57 <sup>c</sup>	15.34 ± 1.58 <sup>c</sup>	10.64 ± 0.47 <sup>c</sup>	8.59 ± 0.39 <sup>c</sup>	11.54 ± 0.92 <sup>c</sup>	8.97 ± 0.86 <sup>c</sup>
DMBA + CVC-ZnO QDs (4 mg)	93.24 ± 5.91 <sup>a</sup>	88.64 ± 6.31 <sup>a</sup>	110.37 ± 9.71 <sup>a</sup>	13.72 ± 1.53 <sup>a</sup>	6.83 ± 0.44 <sup>a</sup>	5.92 ± 0.41 <sup>a</sup>	14.99 ± 0.97 <sup>a</sup>	10.04 ± 0.86 <sup>a</sup>
DMBA + CVC-ZnO QDs (8 mg)	80.68 ± 6.82 <sup>a</sup>	77.21 ± 6.27 <sup>a</sup>	102.34 ± 9.27 <sup>a</sup>	12.81 ± 1.51 <sup>a</sup>	6.57 ± 0.43 <sup>a</sup>	5.54 ± 0.36 <sup>a</sup>	15.99 ± 0.89 <sup>a</sup>	10.21 ± 0.89 <sup>a</sup>

Values are expressed as mean ± SD for six rats in each group. Values not sharing a common superscript (a,b,c) differ significantly at  $p < 0.05$  (DMRT).



**Fig. 2.** Histological changes of kidney tissue of control and experimental rats (H&E staining). Group I (a) control, group II (b) DMBA alone, group III (c) CVC-ZnO QDs alone, group IV (d) CVC-ZnO QDs (2 mg/kg b.w.), group V (e) CVC-ZnO QDs (4 mg/kg b.w.), and group VI (f) CVC-ZnO QDs (8 mg/kg b.w.). Arrows indicate (TD) tubular dilation, (TA) tubular atrophy, (LN) loss of nucleus, (GI) glomerular injury, (ICI) inflammatory cell infiltration, (MTAI) mild acute tubular injury, and (F) fibrosis.

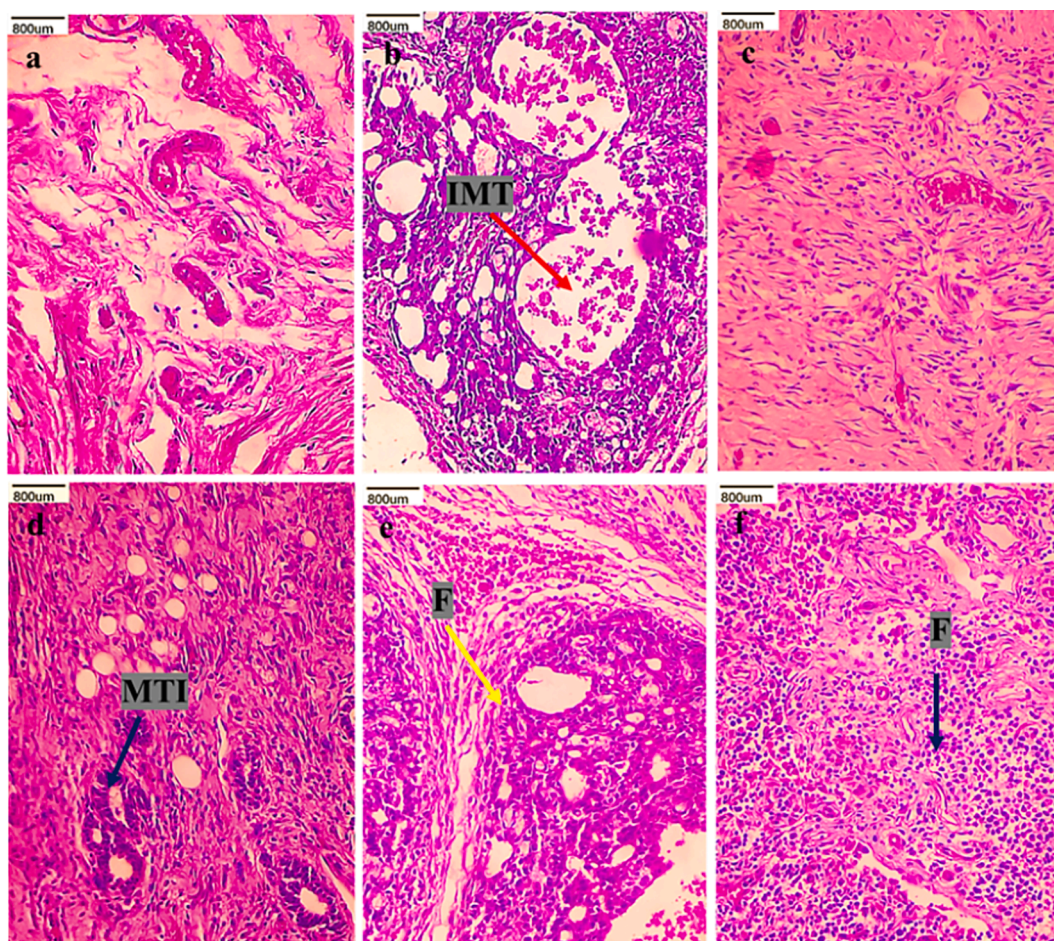
of kidney tissue of DMBA-alone induced rats revealed several features, including tubular dilation, loss of nucleus, tubular atrophy, and glomerular injury. Control and CVC-ZnO QDs alone-rats revealed the normal architecture of the kidney tissue. CVC-ZnO QDs treated DMBA-induced rats showed mild acute tubular injury, inflammatory cell infiltration, and fibrosis. Histology of mammary tissue of DMBA induced rats demonstrated infiltrating malignant tumor. CVC-ZnO QDs treated rats showed mild tumor infiltration and fibrosis with near normal architecture. The control and CVC-ZnO QDs treated alone rats showed normal

architecture of the mammary tissues.

#### 4. Discussion

Nanotechnology enhances cancer treatment by delivering targeted therapies at the molecular level, increasing precision and minimizing damage to healthy cells (Yao et al., 2020). QDs are nanocrystals with unique optical properties that can be used as carriers due to their small size and biocompatibility (Badilli et al., 2020). There have been studies





**Fig. 3.** Histological changes of mammary tissue of control and experimental rats (H&E staining). Group I (a) control, group II (b) DMBA alone, group III (c) CVC-ZnO QDs alone, group IV (d) CVC-ZnO QDs (2 mg/kg b.w), group V (e) CVC-ZnO QDs (4 mg/kg b.w), and group VI (f) CVC-ZnO QDs (8 mg/kg b.w). Arrows indicate (IMT) infiltrating malignant tumor, (MTI) mild tumor infiltration, (F) fibrosis.

on drug-loaded QDs for the chemoprevention of cancerous breast cells. One study used folate receptor–targeted DOX-QD–loaded NPs, to treat female mice with breast cancer induced by a triple-negative cancer cell (Alibolandi et al., 2016). Carvacrol is a natural compound with anticarcinogenic activity (Sivaranjani et al., 2016). The previous study found that CVC caused breast cancer cells to undergo cell death and inhibited the cell proliferation genes involved in cancer (Li et al., 2021). DMBA induces mammary carcinogenesis by acting as a potent chemical carcinogen that causes DNA damage and mutations in mammary epithelial cells. It initiates and promotes the mammary tumors by disrupting the regulatory mechanisms of growth and proliferation of cell (Kerdelhué et al., 2016). The present study investigates the pH-sensitive CVC-ZnO QDs anticancer efficacy in DMBA-induced mammary carcinoma by its ability to scavenge free radicals and detoxify chemical carcinogens.

Antioxidants are essential in protecting cells from the damage caused by ROS (reactive oxygen species), which are highly reactive molecules that can induce oxidative harm to lipids, DNA, and proteins. Lipid peroxidation is a process that occurs when ROS attacks the PUFA (polyunsaturated fatty acids) in cell membranes, leading to the formation of highly reactive lipid radicals that can further damage cell structures (Yaman and Ayhanci, 2021).

Lipid peroxidation and ROS have been implicated in the progression and development of cancer, affecting the metabolisms that lead to weight loss (Davis and Kuttan, 2001). In this research, the growth rate and body weight of DMBA alone rats were reduced because of the changes in biochemical processes during tumor development. In

addition, elevated lipid peroxidation levels play a crucial role in tumor formation. CVC-ZnO QDs-supplemented rats show elevated body weight and growth rate, indicating decreased lipid peroxidation levels. Further, CVC-ZnO QDs decreased the quantity of tumors and the volume of tumors in rats induced with DMBA. This may be due to the targeted drug delivery of nanoparticles (Alibolandi et al., 2016).

Evidence suggests high lipid peroxidation levels are associated with cancer progression and development. This is because the oxidative stress caused by free radicals and ROS can damage DNA and other cellular components, leading to mutations and abnormal cell growth (Jelic et al., 2021). In this study, increased lipid peroxidation levels (TBARS, LOOH, and CD) in DMBA-induced rats were observed. However, CVC-ZnO QDs treated rats show decreased lipid peroxidation levels.

Antioxidants work by neutralizing free radicals, which help to protect cells from damage (Jelic et al., 2021). SOD helps to convert the  $O_2$  (superoxide radical) into  $H_2O_2$  (hydrogen peroxide) and  $O$  (oxygen). CAT converts  $H_2O_2$  into  $H_2O$  and  $O$ , thereby preventing the accumulation of  $H_2O_2$ . GPx uses glutathione as a cofactor to reduce  $H_2O_2$  and lipid peroxides into less harmful molecules. GSH (tripeptide), Vit-C, and Vit-E donate electrons to ROS and free radicals, thereby neutralizing their damaging effects (Goodman et al., 2011). Previous research has shown that antioxidants (enzymatic and non-enzymatic) can reduce the severity and incidence of cancer burden in DMBA-induced animals (Latif et al., 2021). In this study, we observed that antioxidants decreased in the cancer-bearing rats. Vayalakkara et al. (2022) study showed that QDs conjugated with a drug antioxidant could selectively target cancer cells and induce cell death by reducing cellular antioxidant defenses.

CVC-ZnO QDs treated DMBA-induced rats showed increased antioxidant levels due to reduced oxidative stress.

Detoxification enzymes play crucial roles in metabolizing and eliminating chemicals from the body. Phase I metabolism involves the oxidation of DMBA by enzymes such as Cyt-b5 and CYP450. Phase II enzymes, including GR and GST, can conjugate these reactive intermediates with glutathione and other molecules to facilitate their excretion from the body, thereby reducing cancer risk (Dhamodharan et al., 2021). According to Latif et al., (2021), rats induced with DMBA, increase the activity of phase I enzymes and decrease the activity of phase II enzymes. In this investigation, we noted an augmentation of phase I enzyme activity and a corresponding decrease in phase II enzyme activity in rats subjected to DMBA treatment. On the other hand, CVC-ZnO QDs treated DMBA-induced rats show significantly altered detoxification enzymes, possibly due to CVC delivery to breast cancer cells, potentially enhancing the bodies to eliminate carcinogens and prevent cancer development. A previous study observed that CVC could increase and decrease in phases I and II enzymes activity in DMH-induced colon carcinogenesis, respectively (Sivaranjani et al., 2016).

Lipid profile typically includes PL, FFA, TC, and TG measurements in the bloodstream. From a study by Dhamodharan et al. (2021), DMBA-induced mammary cancer in rats is associated with significant increases in plasma PL, FFA, TC, and TG levels. High PL, FFA, TC, and TG levels, on the other hand, are associated with increased cancer risk. This may be because cholesterol is a precursor for estrogen, a hormone that can promote the growth of mammary cancer. In our study, we noticed that lipid profile (PL, FFA, TC, and TG) were increased in the plasma of DMBA-induced rats, while in mammary tissues, the PL and FFA levels were decreased. This decrease may result from increased PL degradation, impairing membrane function. Another possible explanation for the reduced PL concentration could be the decreased FFA levels in the mammary tissues, as suggested by Van Hoeven and Emmelot. in 1973. Due to the strong antihyperlipidemic properties of CVC-ZnO QDs, supplemented DMBA-treated rats showed significantly decreased plasma PL, FFA, TC, and TG lipid profiles. In contrast, elevated levels of PL and FFA were observed in the mammary tissues.

Nandakumar and Balasubramanian (2011) reported that DMBA-treated rats showed renal tubular structure enlargement and loss of architecture. In this present study, DMBA-induced rats show significant kidney damage, including tubular dilation, loss of nucleus, tubular atrophy, and glomerular injury. Nanoparticles can be engineered to selectively target the kidneys, allowing for precise delivery of drugs to the affected area. This can be particularly useful in treating conditions such as kidney inflammation. In this context, Ozturk et al. (2018) suggested that CVC could reduce kidney injury induced by bilateral renal ischemia/reperfusion (I/R) in female SD rats. In our study, treatment with CVC-ZnO QDs at different doses significantly reduced these pathological changes, with the 4 mg/kg b.w. dose showing the greatest protective effect.

Histopathology of mammary tissue of DMBA-induced rats exhibited IMT. These results agree with Latif et al. (2021). On the contrary, rats treated with CVC-ZnO QDs displayed no indications of necrosis or cellular growth in the mammary gland. A more noticeable impact was noted in the rats subjected to a dosage of 4 mg/kg b.w. of CVC-ZnO QDs, where the mammary tissue exhibited nearly normal architecture with an increased area of fibrosis representing residual tumor. Therefore, those data suggest that the CVC-ZnO QDs have the capability to provide a secure and efficient approach to treating cancer with significant anticancer activities.

## 5. Conclusion

Based on the research findings, pH-sensitive CVC-ZnO QDs have a significant anticancer effect against mammary carcinoma induced by DMBA in female SD rats. The results demonstrate that CVC-ZnO QDs significantly reduced tumor growth, improved the antioxidant status,

decreased lipid peroxidation, and modulated the biotransformation enzyme activities in both plasma and mammary tissues. CVC-ZnO QDs also exhibited a positive impact on the lipid profile. Furthermore, the histological examination of the mammary and kidney tissue revealed that CVC-ZnO prevents DMBA-induced tissue damage. These findings suggest that CVC-ZnO QDs can be a safe and effective anticancer agent for treating mammary carcinoma. However, further studies are needed to explore the molecular mechanisms underlying the anticancer properties of CVC-ZnO QDs.

## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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## Data availability statement

Data will be provided upon request.

## Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jksus.2023.103029>.

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