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Original article

## Distribution of *Candida* infections in patients and evaluation of the synergic interactions of some drugs against emerging fluconazole- and caspofungin-resistant *C. auris*



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### ABSTRACT

Pathogenic multidrug-resistant *Candida* species are considered some of the most important health risks. This work aimed to evaluate and monitor the prevalence of the human pathogenic *Candida* strains isolated from patients in King Fahd Medical City (KFMD), Riyadh, Saudi Arabia, and to evaluate the synergy of some antimicrobial agents against *Candida* species' resistance to antifungal drugs. The retrospective analysis, identification using biochemical tests, minimal inhibitory concentrations using E-tests, determination of the fraction inhibitory concentration index value for synergic testing, and simulation of 100 experiments using Monte Carlo simulation methods were performed according to standard protocols. The findings showed that all age groups of males and females can be infected by *Candida* species; furthermore, human pathogenic *Candida* species can be isolated from several clinical samples and different human body sites. The minimal inhibitory concentration results showed that many multidrug-resistant *Candida* strains, such as *C. albicans*, emerged in 2020 compared to 2018. *Candida albicans* remains the most important pathogen among all *Candida* species, found in 51.7 % and 42.4 % of the isolates in 2018 and 2020, respectively. In 2018, many isolates of *C. albicans* showed resistance to itraconazole, fluconazole, anidulafungin, amphotericin B, ketoconazole, voriconazole, caspofungin, and flucytosine. In 2018, all *C. auris* isolates ( $N = 94$ ) were resistant to fluconazole, and more than 85 % ( $N = 76$ ) of *C. albicans* isolates were resistant to itraconazole, while only 5.9 % ( $N = 2$ ) were resistant in 2018. The study concluded that the resistance to antifungal drugs among pathogenic yeasts is increasing and constantly changing and that surveillance of these pathogens must continue. Also, the synergy between drugs remains an appropriate option for confronting this risk, especially between natural extracts and drugs. Despite the lack of evidence for any antifungal and antibacterial drug's ability to synergistically suppress the fluconazole- and caspofungin-resistant *C. auris* strains diagnosed in this study, the surveillance and synergic tactics continue to be viable options for dealing with these human pathogenic yeasts.

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### 1. Introduction

*Candida* species are eukaryotic microbes that belong to the fungi kingdom. These fungi (yeasts) live naturally on the skin and inside the human body and have the ability to occupy various natural habitats. *Candida* species can also cause serious diseases if they find a way to penetrate the internal parts of the human body, such as the kidney, heart brain, and bloodstream (Lynch 1994, Jabra-Rizk et al., 2016).

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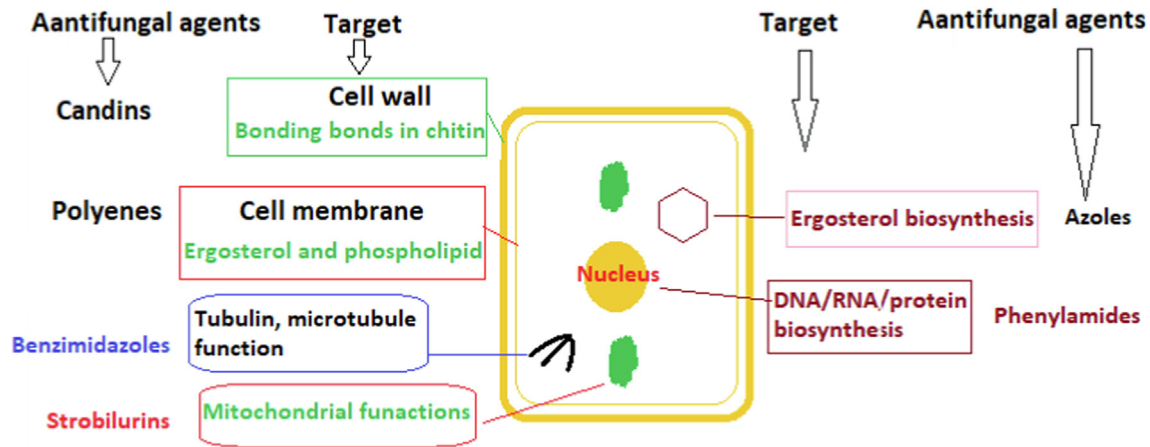


Fig. 1. Mode of biological action of antifungal agents.

There are only three types of drugs (azoles, polyenes, and 5-fluorocytosine) currently used to fight fungal infection, and they use different mechanisms to kill or inhibit pathogenic fungi, including inhibition of ergosterol biosynthesis (azole antifungal drugs), disruption of the cell membrane (polyene antifungal drugs), and inhibition of DNA and RNA biosynthesis (5-fluorocytosine) (Carrillo-Munoz et al., 2006, Whaley et al., 2017, Van Daele et al., 2019). As available options are limited, the emergence of drug-resistant pathogenic fungi is a highly dangerous global risk for public health. Some strains of *Aspergillus* and *Candida* species have the ability to resist numerous standard antifungal drugs (Dismukes 2000, Houšř et al., 2020), and some *Candida* species are able to resist antifungal drugs via several mechanisms. For example, azole resistance mechanisms include inactivation of C-5 sterol desaturase, uptake of the sterols from out-environments, efflux pumps, inhibition of the main transporters to reduce the accumulation of azoles inside the cell, and reducing azole binding to lanosterol 14- $\alpha$ -demethylase (Ryley et al., 1984, Whaley et al., 2017). Polyene-resistant *Candida* strains can defeat polyene antifungals through genetic mutations in genes that encode enzymes for ergosterol biosynthesis or modification of the sterols and fatty acid of the cytoplasmic membrane and of the ratio of sterols to phospholipids (O'Shaughnessy et al., 2009, McCarthy et al., 2017). Mechanisms of resistance to 5-fluorocytosine in *Candida* species need further investigation. A genetic study on 5-fluorocytosine-resistant *Candida glabrata* showed genetic mutations in genes that encode cytosine deaminase, uridine monophosphate, and pyrophosphorylase. Thymidylate synthase or a cytosine permease may be responsible for the resistance to 5-fluorocytosine (Vandeputte et al., 2011). The mode of biological action of antifungal agents against human pathogenic yeast and mold is represented in Fig. 1.

In recent years, monitoring and screening studies have shown a global prevalence of antifungal-resistant strains in hospital-acquired infections caused by *Candida* species. It has been reported that 7% of the *Candida* strains isolated from blood samples collected by the Centers for Disease Control and Prevention (CDC) were pathogenic isolates that have the ability to resist antifungal drugs (<https://www.cdc.gov/fungal/diseases/candidiasis/antifungal-resistant.html>). Globally, it has been reported that more than 2.5% and 9% of human pathogenic *Candida* species can resist fluconazole and itraconazole, respectively (Pfaller et al., 2000). Multidrug-resistant *C. auris* strains have been isolated and diagnosed in ear, blood, and invasive infections in many countries around the world (Chowdhary et al., 2016). In Saudi Arabia, numerous studies have confirmed the presence of strains of resistance in

*C. auris* to several antifungal drug categories (Lone and Ahmad 2019, Aljindan et al., 2021). The importance of the present research is in monitoring this highly dangerous pathogenic yeast, and the ongoing need to update data on its resistance to standard drugs, especially, with the rare studies on synergistic methods in Saudi Arabia as an option that can be applied in the treatment of this pathogenic yeast. This work aimed to evaluate and monitor the prevalence of the human pathogenic *Candida* strains isolated from patients in King Fahd Medical City (KFMD), Riyadh, Saudi Arabia, and to evaluate the synergy of some antifungal drugs against *Candida* species' resistance to antifungal drugs.

## 2. Methodology

### 2.1. Retrospective analysis

From January 1, 2018, to August 30, 2018, and January 1, 2020, to December 30, 2020, the data of antifungal-resistant *Candida* species isolated from patients in KFMC were analyzed to evaluate the biological differences between microbial species and strains

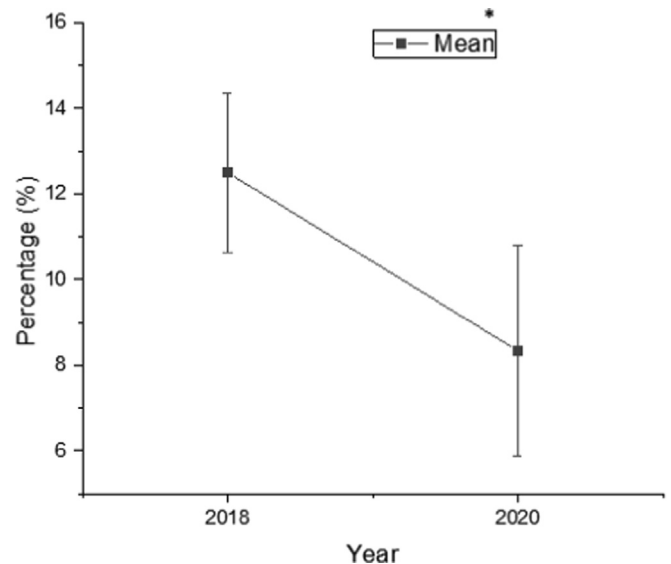


Fig. 2. Means (%) of *Candida* spp. and other pathogenic yeasts isolates identified during 2018 and 2020. \*At the 0.05 level, the means (%) are significantly different ( $P < 0.05$ ).

regarding biochemical characteristics and responses to standard antifungal agents. The data were analyzed using chi-square test, Ward linkage method, and one-way ANOVA (IBM SPSS Statistics 26 and OriginPro 2018). The work was part of an investigation approved by the Ethics Committee, KFMC, Saudi Arabia, IRB: 21-466E.

### 2.2. Testing standard antibiotics against antifungal-resistant *Candida* strains

Minimal inhibitory concentrations (MICs) (Moore et al., 1987) were determined to screen the biological activity of the standard antibacterial drugs against *Candida* strains using E-tests of itraconazole, fluconazole, anidulafungin, amphotericin B, ketoconazole, voriconazole, caspofungin, and flucytosine (Biomerieux, France).

### 2.3. Antifungal-resistant *Candida* strains collection

The *Candida* strains that showed resistance to standard antifungal agents were cultivated on blood agar (Oxoid, UK). The *Candida* strains were preserved in a sterile glycerol solution (30 % V/V) at

–80 °C, and the subcultivation was done at least three times before the further tests.

### 2.4. Synergic test of antifungal and antibacterial agents

The synergy between some standard antibiotics and antifungal drugs against fluconazole- and caspofungin-resistant *C. auris* strains was evaluated using an E-test assay according to (Orhan et al., 2005). The fractional inhibitory concentration (FIC) index ( $\Sigma$ FIC) of the tested drugs was calculated using the following equation (Mehta et al., 2022):

$$\Sigma FIC = FIC_{d1} + FIC_{d2}$$

$$FIC_{FIC} = (MIC \text{ of combination } d1 \text{ and } d2 / MIC \text{ of } d1) FIC_{D2} = (MIC \text{ of combination } d1 \text{ and } d2 / MIC \text{ of } d2)$$

The simulation of 100 experiments on FIC ( $N = 2$ ) used in this investigation was performed using Monte Carlo simulation methods according to (Meletiadis et al., 2010) with some modifications.  $\log_2$  of the FICs were calculated, and then the mean and standard deviation were determined. The normal distribution was tested using the *Norm.dist(FIC, mean, standard division, false)* function in Excel 2016. The first simulation was calculated using the *norm.*

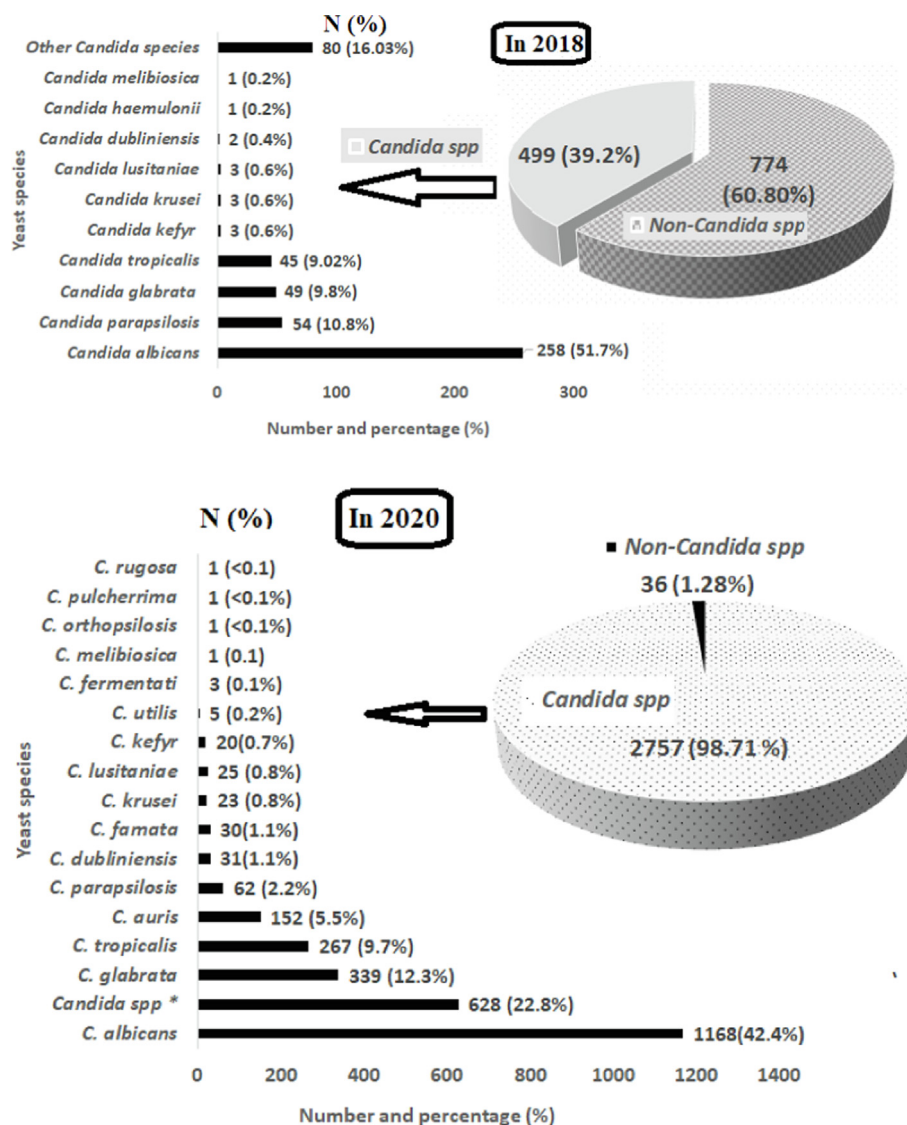


Fig. 3. Number (N) and percentage (%) of *Candida* and non-*Candida* spp. isolated and diagnosed in patients during 2018 and 2020.

inv(rand.)mean, standard division) function in Excel 2016. Two hundred tests of expected FICs were done using a data table and calculating new functions. The percentage of simulated FICs that showed synergy was calculated using the countif(FICs, "<=1"/100) function.

### 3. Results

In 2018, 63.5 % of patients infected by pathogenic yeasts were females, but in 2020, 52 % of the infected patients were male. Pearson chi-square analysis reported a significant (at the 0.05 level) association between males and females in both 2018 and 2020.

The results shows that all age groups can be infected by pathogenic yeast. The prevalence of pathogenic yeast infection for the age group 61–80 years was 28.3 % in 2018 and 35.48 % in 2020. The lowest prevalence of infection was in babies younger than one year of age. In both 2018 and 2020, 11.5 % of males and 8.31 % of females older than 80 years of age were infected. In 2020, more than 50 % of the patients infected by pathogenic yeasts were 15–80 years old.

The mean monthly rates of pathogenic yeast isolates (*Candida* and non-*Candida* species) diagnosed were 12.5 % during 2018

and 8.3 % during 2019. The statistical analysis showed a significant difference between 2018 and 2020 (Fig. 2).

In 2018, 39.2 % of the pathogenic yeasts isolated from patients were *Candida* species; *C. albicans* strains exceeded 50 % of the isolates, and ten species of *Candida*, including *C. parapsilosis* (10.8 %), *C. glabrata* (9.8 %), and *C. tropicalis* (9.02 %), were isolated and identified. In 2020, *Candida* species were 98.8 % of isolates; *C. albicans* strains were 42.4 %, *C. parapsilosis* were 2.2 %, *C. glabrata* were 12.8 %, and *C. tropicalis* were 9.7 %. Many species emerged, such as *C. auris* (5.5 %) and *C. famata*, bringing the total number of species to 16 (Fig. 3).

The results showed that the urine, respiratory, and blood samples were the major sources of *Candida* and non-*Candida* isolates. In 2018, more than 50 % of *Candida* and non-*Candida* species were isolated from urine samples, and in 2018 and 2020, respectively, 83.8 % and 67.6 % of the isolates were obtained from urine, respiratory, and blood samples.

The finding shows the number and percentage of *Candida* species that have the ability to resist one or more of the standard antifungal drugs tested in this investigation: 16.6 % of *Candida* isolates were resistant to fluconazole, 13.4 % to itraconazole, 11.9 % to amphotericin B, 11.1 % to voriconazole, 7.1 % to caspofungin, 4.7 % to anidulafungin, 4.0 % to flucytosine, and 1.2 % to ketoconazole.

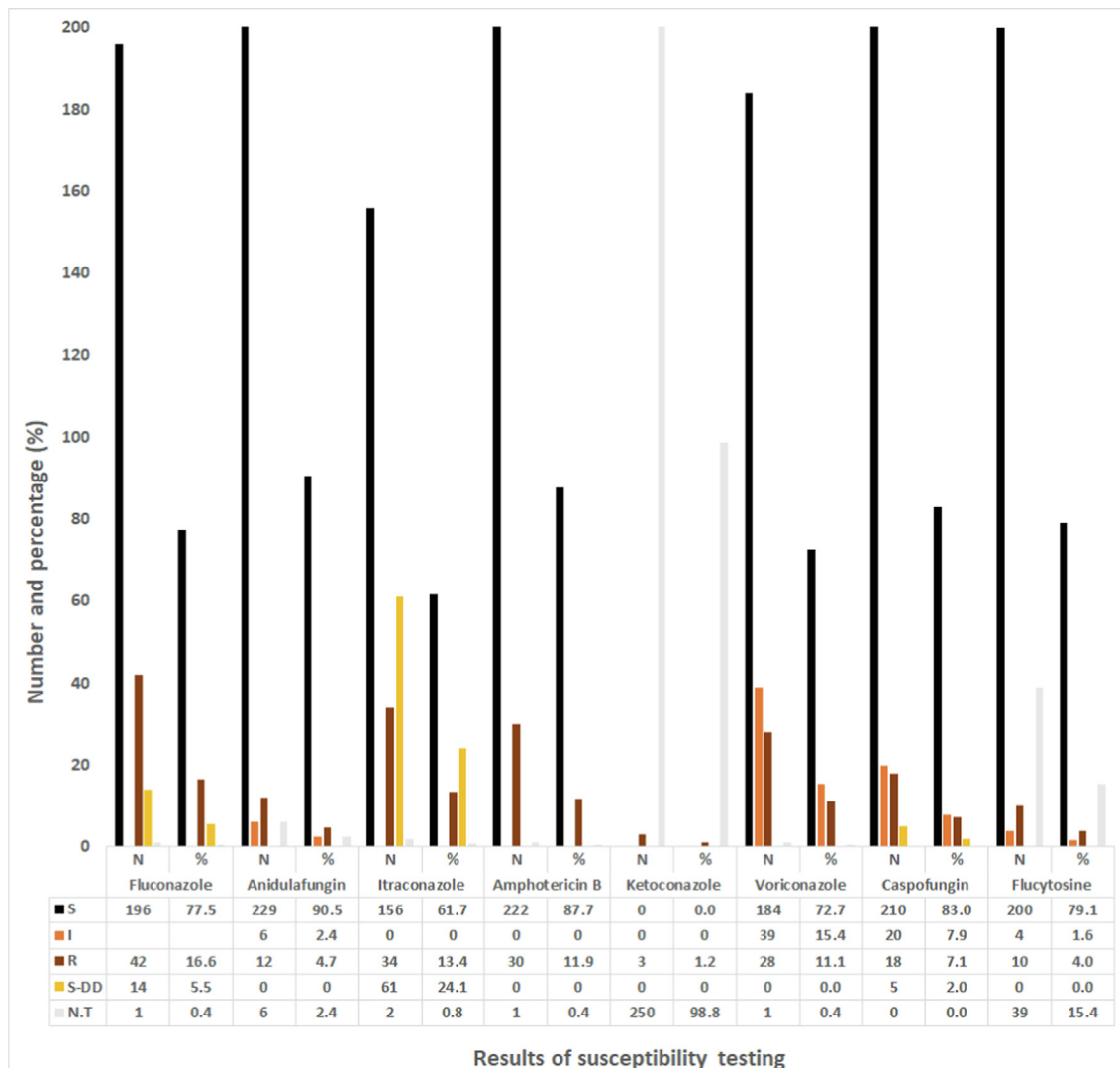


Fig. 4. Number (N) and percentage (%) of *Candida* spp. (N = 253) that have the ability to resist the standard antifungal drugs in 2018. \*All MICs values were interpreted according to CLSI and EUCAST.

zole (Fig. 4). Itraconazole was resisted by *C. glabrata* (79 %), *C. tropicalis* (15 %), and *C. albicans* (5.9 %). No isolate of *C. haemulonii*, *C. parapsilosis*, *C. krusei*, and *C. lusitaniae* resisted itraconazole. More than 30 % of the *C. albicans* isolates were resistant to amphotericin B and ketoconazole. Although most *Candida* strains that resisted itraconazole, fluconazole, anidulafungin, ketoconazole, ketoconazole, and caspofungin were isolates of *C. glabrata*, no *C. glabrata* strain isolated in this work had the ability to resist flucytosine.

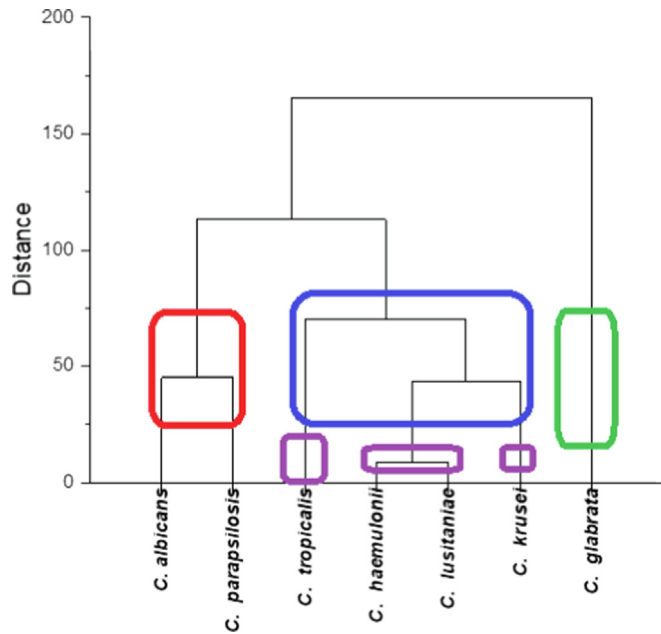


Fig. 5. Hierarchical clusters of *Candida* strains isolated in 2018 from patients that have resistance to itraconazole, fluconazole, anidulafungin, amphotericin B, ketoconazole, voriconazole, caspofungin and flucytosine. Percentage (%) analyzed using Ward method in OriginPro software.

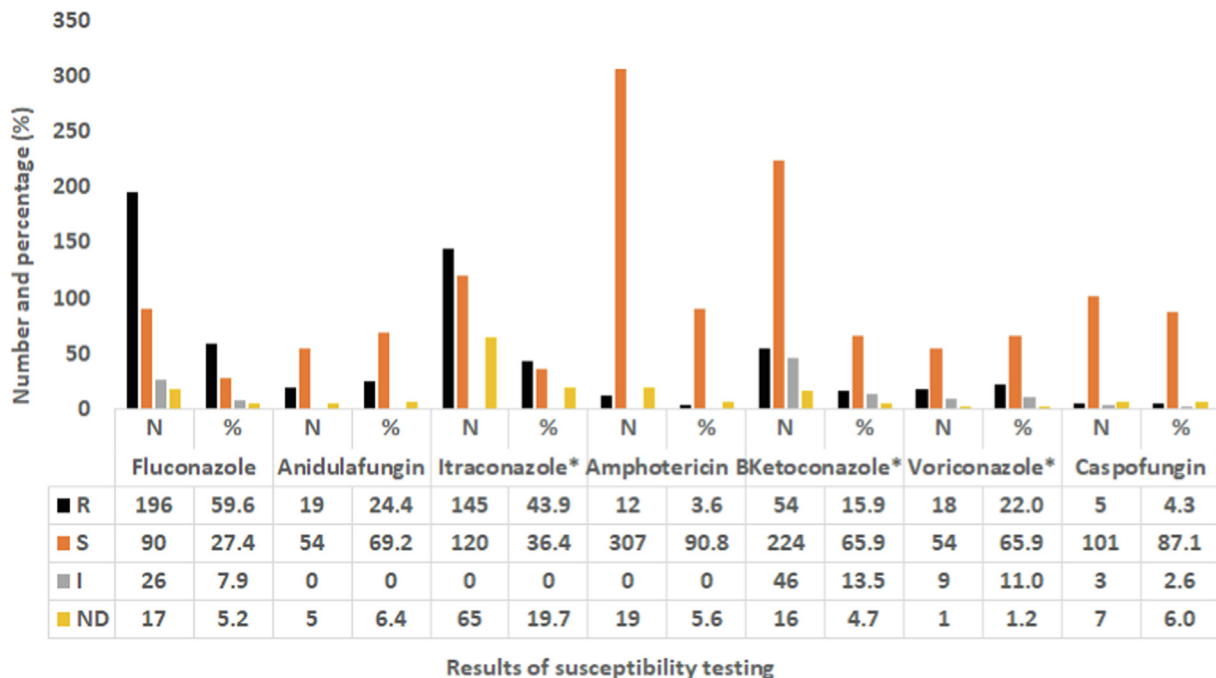
In the hierarchical cluster analysis of *Candida* strains, *C. albicans* and *C. parapsilosis* isolates were grouped in the cluster with the isolates resistant to fluconazole, amphotericin B, voriconazole, caspofungin, and flucytosine (Fig. 5 and Fig. 7). *C. glabrata* isolates were grouped in an independent cluster; more than 30 % of *C. glabrata* isolates were resistant to itraconazole, anidulafungin, ketoconazole, voriconazole, and caspofungin, and all isolates were susceptible to flucytosine.

Fig. 6 shows that more than 50 % of the isolates were resistant to fluconazole and ketoconazole, and some isolates showed resistance to anidulafungin, itraconazole, amphotericin B, voriconazole, and caspofungin. (Only 4.3 % of the isolates were resistant to caspofungin.) All isolates of *C. auris* were resistant to fluconazole. In addition, more than 75 % of isolates of *C. albicans*, *C. tropicalis*, and *C. parapsilosis* were resistant to itraconazole.

The results show no synergy between fluconazole and caspofungin; furthermore, none of the antibiotics tested in this work, which included cefepime, ceftazidime, and moxifloxacin, showed any effect. Additive action was only recorded between cycloheximide and caspofungin, for which the FIC index was  $\geq 0.5$  (Fig. 8). Simulation of 100 tests using Monte Carlo simulation methods showed that 61 % of the experiments would probably show the same results.

#### 4. Discussion

The results of this work confirmed that males and females of all age groups, children, adults, and elders, are at risk of infection. In 2018, we found a female:male infection ratio of 1.7:1, which agrees with the results of (Loster et al., 2016), who found more infections in females than in males (1.9:1 female:male). In 2020, our results were completely different: the infection ratio was 0.9:1 (female:male), different from that in Loster his colleagues but in agreement with (Al Thaqafi et al., 2014) (0.87:1, female:male). The female-to-male infection rate varies notably from year to year according to our results and the results obtained from previous studies, for example, Loster and his colleagues (2016); Al Thaqafi and his col-



Results of susceptibility testing

Fig. 6. Number (N) and percentage (%) of *Candida* spp. (N = 370) that have the ability to resist the standard antifungal drugs in 2020. \*All MICs values were interpreted according to CLSI and EUCAST.

leagues (2014). There are many psychosocial and medical factors that can answer the question whether the *Candida* and non-*Candida* infection is dependent on gender, but the answer will differ by case and country.

The *Candida* infections are caused by many species of opportunistic pathogens (Vázquez-González et al., 2013), so it is a logical impossibility that elderly individuals (more than 60 years) are most susceptible to infections, as confirmed by our results. Lack of correlation between age and average number of isolates is common in many previous studies; for example, as in our study, Kim et al. (Kim et al., 2016) showed that the average number of isolates varies according to many factors.

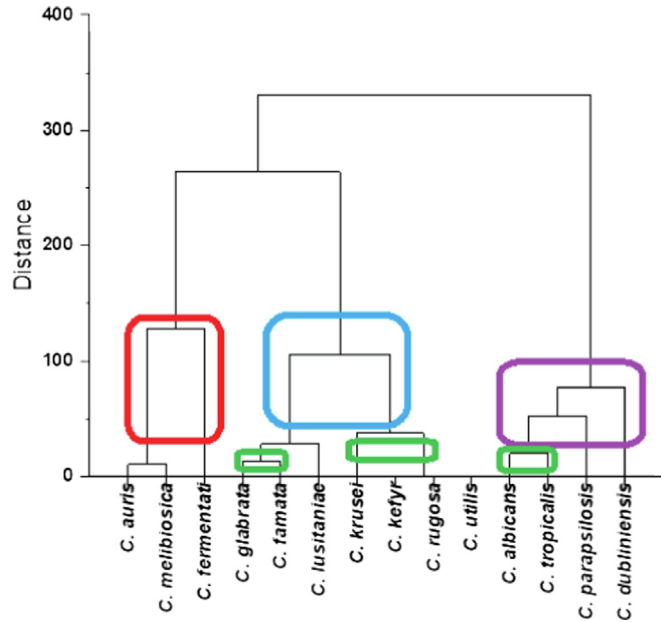


Fig. 7. Hierarchical clusters (Ward method) of *Candida* strains isolated in 2020 from patients that have resistance to fluconazole, itraconazole, amphotericin B and ketoconazole.

We found many antifungal drug-resistant *Candida* strains that emerged in 2020. Emerging drug-resistant *Candida* strains have been reported in several countries. For this reason, improving current strategies and monitoring processes are urgently needed to fight these fungi. Notwithstanding the isolation of three isolates of fluconazole-resistant *C. haemulonii* and voriconazole-resistant *C. haemulonii* in 2018, not one strain of *C. haemulonii* was isolated in 2020. It has been reported that *C. haemulonii* is a rare opportunistic pathogen, and the accuracy of the identification for this yeast is complex due to its high similarity to other *Candida* strains (Coles et al., 2020).

*C. albicans* strains remain major pathogens despite the changes in the species and numbers of *Candida* isolates taken from patients in 2018 and 2020. Reports published in previous years confirmed the predominance of *C. albicans* over other *Candida* species (Talapko et al., 2021). Grouping of the isolates based on susceptibility test patterns showed a difference between 2018 and 2020. For example, in 2018, all *C. glabrata*, *C. tropicalis*, *C. haemulonii*, *C. krusei*, and *C. lusitaniae* isolates were susceptible to flucytosine. In 2018, all tested *Candida* isolates except 10.6 % of the isolates of *C. auris* were susceptible to caspofungin. Caspofungin and anidulafungin (echinocandin drugs) can damage yeast cell walls by inhibiting the enzymatic systems that form glucan, but the present work confirmed emerging isolates of *C. auris* that have the ability to resist caspofungin and anidulafungin. Also, in this work, anidulafungin-resistant *C. albicans*, *C. tropicalis*, *C. glabrata*, and *C. krusei* strains were isolated from the patients.

In medical strategies, synergistic interactions are frequently recommended to maximize biological activity to remove infectious agents; however, the relationship between these interactions and the development of multi-drug resistance is not yet clear (Nyilasi et al., 2010). Our results confirm that synergistic interaction strategies using tested standard drugs have limited effectiveness in fighting fluconazole- and caspofungin-resistant *C. auris* strains isolated from patients in KFMC. The synergistic interaction strategies between natural compounds and standard drugs remain good options that need to be investigated further in the future. A good result has been obtained from the formulation of the fungicide cycloheximide (By interfering with the translocation stage, the

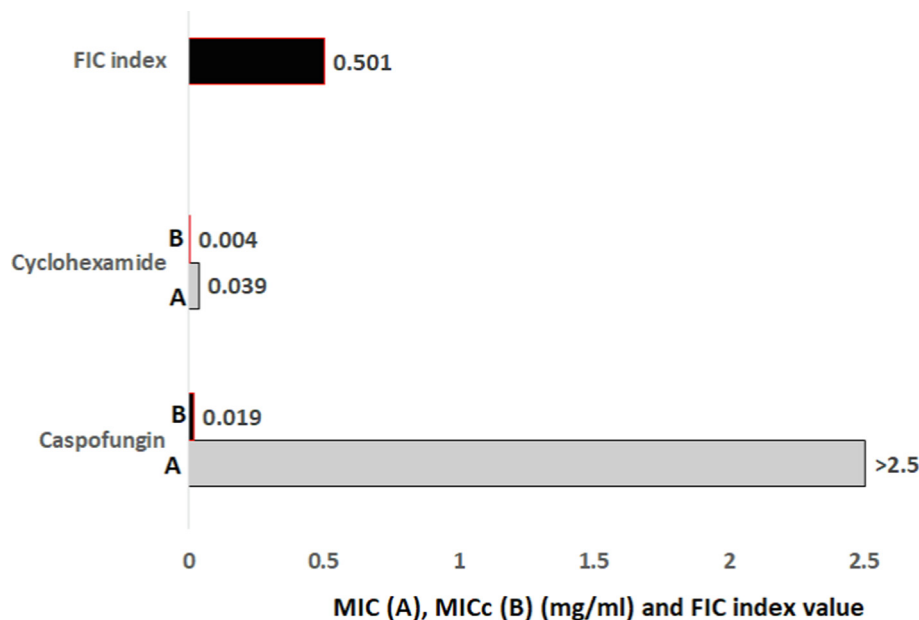


Fig. 8. Minimal inhibitory concentration (MIC) (mg/mL) and fractional inhibitory concentration (FIC) index that showed activity against *C. auris* strain isolated from the patients. \*MIC (A) for each drug alone, MICc (B) for formulation of cycloheximide and caspofungin (20:2.5 mg/ml).

activity of two tRNA molecules and an mRNA relative to the ribosome, it inhibits translation extension and hence blocks the production of proteins (Schneider-Poetsch et al., 2010)), and caspofungin (it prevents the biosynthesis of beta-(1,3)-D-glucan (Letscher-Bru and Herbrecht 2003)), but the toxicity of cycloheximide limits this option. In the present work, the *Candida* species were isolated from many types of samples, including medical materials; for this reason, cycloheximide can be used for some medical media and materials such as transport media. These findings agree with the conclusion of (Dal Pizzol et al., 2021), who reported that cycloheximide led to decreasing fungal growth in an optisol-GS medium without any toxicity to the endothelium of the corneal material transported in it.

## 5. Conclusion

The results of our work show that every age group is susceptible to infection by a human pathogenic *Candida* species at any time of the year. Major infections were identified in male and females aged 61–80 years. *Candida* strains can be isolated from clinical samples from many sources, including the human body (including the respiratory system, the axilla, the groin, superficial wounds, body tissues, eye, ear, under the skin), body fluids (urine, blood, peritoneal drain, abscess, cerebrospinal fluid, aspirated fluid, surgical drain fluid, oropharyngeal samples, oral swabs, vaginal swabs), and medical tools (catheter tools, stent devices, nephrostomy tubes). In 2018 and 2020, more than 65 % of *Candida* species were isolated from urine, respiratory, and blood samples; *C. albicans* strains were the major pathogens, and *C. glabrata* and *C. tropicalis* strains are still the main pathogens among all patients. The *C. auris* strain was not isolated in 2018 but emerged in 2020 as one of the most resistant to antifungal drugs. Drug resistance was shown in several *Candida* species, including the major pathogenic *Candida* species. There is an increasing and constant change in the number and types of yeast strains that can cause disease and develop resistance to drugs. Although there is no evidence of synergy between any of the antifungal drugs tested that would inhibit the fluconazole- and caspofungin-resistant *C. auris* strain isolated in this work, surveillance and synergic strategies remain suitable choices to confront these human pathogenic yeasts.

## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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