



Original article

Gold nanoparticles (AuNPs) and *Rosmarinus officinalis* extract and their potentials to prompt apoptosis and arrest cell cycle in HT-29 colon cancer cells



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ABSTRACT

Background: One of the significant morbidity and mortality causes is the colorectal cancer. *Rosmarinus officinalis* plant is utilized as food and medicine. Currently, nanoparticles are widely employed in medicinal preparations. This work aimed to explore the potentials of *R. officinalis* leaves acetone extract, alone or in combination gold nanoparticles (AuNPs), to kill colon cancer cells.

Methods: Fresh leaves of *R. officinalis* were collected from the Al Soudah, Saudi Arabia and then dried to prepare acetone extract. AuNPs were prepared utilizing the extract and portrayed using UV/Vis spectrophotometry and scanning electron microscopy (SEM). Active ingredients exist in the extract and extract + AuNPs were screened using FT-IR spectroscopy. Biological properties of the extract and extract + AuNPs including anti-cancer activity and apoptotic capacity were studied.

Results: Results of UV/Vis spectrophotometry and SEM demonstrated that AuNPs are of 79 nm in diameter. FTIR analysis revealed the existence of bioactive molecules in the extract. Extract and extract + AuNPs arrested HT-29 colon cancer cells at G2/M phase.

Conclusion: *R. officinalis* acetone extract and extract + AuNPs could arrest the proliferation of HT-29 colon cancer cells. Extract and extract + AuNPs actuated apoptosis in cancer cells as opposed to necrosis.

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1. Introduction

Normal cells divide under well-controlled manner according to host body needs. Cancer cells, the abnormal version of host cells, are characterized by their powerful capacity of proliferation with reduced capacity of apoptosis. The abnormal cell proliferation of cancer is due to the loss of perfect control mechanisms in the production of growth factors leading to imperfect cellular homeostasis and maintenance causing abnormal tissue architecture (Hanahan and Weinberg, 2011). Several pharmaceutical derivatives have

been obtained through the screening of plant natural compounds and showed anticancer properties (Da Rocha et al., 2001).

Rosmarinus officinalis (Rosemary) plant is one of the Lamiaceae (mint) family. It is widely existing in the Mediterranean region and distributed in many other locations through the world. Its leaves are utilized in the treatments of many disorders long time ago as well as food additives. Rosemary has many medicinal activities such as antibacterial (Nieto et al., 2018), the power to treat depressive behavior (MacHado et al., 2012), antitumor (Kontogianni et al., 2013), antioxidant (Nieto et al., 2018), hepato-protective power (Sotelo-Félix et al., 2002; Abdel-Wahhab et al., 2011; Rašković et al., 2014), anti-parasitic, wound-healing agent (Hamidpour, 2017), antispasmodic in renal colic, smooth muscle relaxant (Habtariam, 2016), gastric ulcerative lesions (Corrêa Dias et al., 2000), control of hypercholesterolemia, relief of physical fatigue (Fernández et al., 2014), lipid peroxidation reduction (Posadas et al., 2009), radio-protective-antimutagenic capacities (Del Baño et al., 2006), treatment for cutaneous allergy (Tabassum and Hamdani,

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2014), anti-inflammatory (De Melo et al., 2011), antiviral (Nolkemper et al., 2006), antithrombotic (Yamamoto et al., 2005), and anti-hyperglycemic (Naimi et al., 2017).

Several phytochemicals with biological characteristics were obtained from rosemary oil and extracts. These phytochemicals include acids (rosmarinic, ursolic, oleanolic, caffeic, and carnolic), eucalyptol, camphor, rosmadial, secohinokio, and eugenol/luteolin derivatives (Borges et al., 2019; Einbond et al., 2012; Gonçalves et al., 2019). Rosemary preparations (e.g. carnolic and rosmarinic acids and diterpenes carnosol) are believed to apply strong biological effects (e.g. antitumor, anti-allergic, antibacterial and antioxidant) (González-Vallinas et al., 2015; Moore et al., 2016). Several studies demonstrated that *R. officinalis* exhibit good anti-cell propagation properties against some cancer cell lines (Kontogianni et al., 2013; Petiwala et al., 2013). Many plants, including edible ones, have medicinal impacts against several diseases including colon cancer (Balakrishna and Kumar, 2015; Xu et al., 2015).

The science concerning with study and manufacturing of nano-dimension materials is called nanotechnology (Rajeshkumar, 2016). Gold nanoparticles (AuNPs) production employing plant extract is clean, ecofriendly, and cost effective method comparing to other methods (chemical and physical). Gold nanoparticles are reported to have anti-bacterial (Mohamed et al., 2017), anticancer (Rajeshkumar, 2016b) and immunomodulatory (Dykman and Khlebtsov, 2017) activities.

In the current work, biological proprieties (anticancer, antibacterial, effects on cell cycle/apoptosis) of *R. officinalis* leaf acetone extract and its synthesized AuNPs on HT-29 cells were examined. In addition, the power of the extract to produce AuNPs was evaluated.

2. Experimental methods

2.1. Rosemary leaf extract preparation

Leaves of *R. officinalis* (Fig. 1) were collected in August 2019, from Abha, Aseer, Saudi Arabia. The leaves extract was prepared as described elsewhere (Ibrahim et al., 2021). A stock preparation (1%) was designed in acetone, sterilized (0.45 μm filter, Fisher Scientific) and stocked at $-20\text{ }^{\circ}\text{C}$.



Fig. 1. *R. officinalis* plant collected from Al Soda mountain, Aseer, KSA.

2.2. AuNPs synthesis/characterization and functional group analysis

AuNPs preparation, characterization, size/morphology, and plant extract functional group analysis were done according Ghramh et al. (2019).

2.3. Maintenance, preparation of cells and cytotoxicity tests

HT-29 cancer cell line was maintained utilizing the same methods and reagents described by Ganesan et al. (2020) The investigation of the extract and extract/AuNPs cytotoxicity towards HT-29 cells was done according to Ghramh et al. (2021) at concentration of 0, 0.5, 1.0, 2.0, 4.0, 8.0, 16.0, 32.0, 64.0 132.0, 264, 528.0 and 1056.0 $\mu\text{g}/\text{mL}$.

2.4. Apoptotic effects of extract and extract/AuNPs

The extract and extract containing AuNPs at IC_{50} concentrations were separately added to HT-29 cells and incubated for 48 h. The apoptotic effect was tested using similar reagents and methodology described by Ibrahim et al. (2021).

3. Results

3.1. Gold nanoparticles production

Gold nanoparticles synthesis was observed through the color alteration of the mixture (AuCl_4 extract, Fig. 2A). After color alteration (Fig. 2C), AuNPs assembling was reviewed spectrophotometrically (Fig. 2D). Results uncovered the manufacturing of AuNPs at a specific peak (520 nm).

3.2. Functional groups

The FTIR spectral analysis of the *R. officinalis* extract is displayed in Fig. 3. Strong broad band (characteristic to alcoholic) in the range of $3633\text{--}3300\text{ cm}^{-1}$, concerned to stretching vibration of O—H groups. Two bands (2938 and 2851 cm^{-1}) are due to stretching vibrations of the groups CH_2 and CH_3 . Bands present at 1717 , 1696 and 1661 cm^{-1} are due to stretching vibration of $\text{C=O}/\text{C=C}$ groups of flavonoids and amino acids. A peak ranged from 1457 to 1276 cm^{-1} is attributed to aromatic C=C and due to the presence of series of small peaks in the area $1400\text{--}2000\text{ cm}^{-1}$. Numerous peaks found at $1030\text{--}523\text{ cm}^{-1}$ represent C—O stretch of acid, anhydride, ester, alcohol, ether, monosubstituted alkene and halo compound.

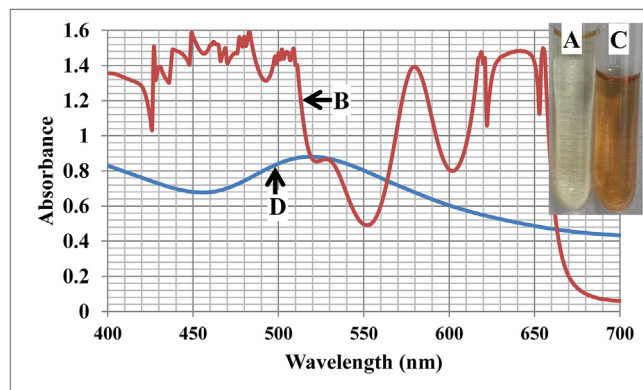


Fig. 2. UV/Vis light monitoring of AuNPs synthesis by *R. officinalis*; A: Extract; B: extract light absorbance; C: extract after synthesis of AuNPs; D: extract + AuNPs light absorbance.

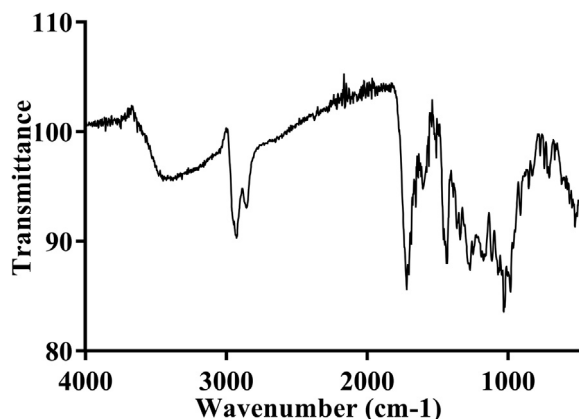


Fig. 3. FTIR spectrum of *R. officinalis* extract.

3.3. Characterization of AuNPs

SEM examination divulged that the manufactured AuNPs are nearly uniform spherical in shape with median size of 79 nm.

3.4. Extract cytotoxicity toward HT-29 cells

R. officinalis extract inhibited HT-29 cell line growth at a significant ($p > 0.0001$) levels at the concentration 32–1000 $\mu\text{g/mL}$. Extract containing AuNPs also inhibited HT-29 cells growth at a significant ($p > 0.0001$) levels but only up to concentrations ranged 32–1000 $\mu\text{g/mL}$ (Fig. 4).

3.5. Apoptotic effects of *Rosmarinus officinalis* acetone extract

Evaluation of the extract and extract containing AuNPs was done using Annexin V staining (Table 1). The extract caused significant ($p < 0.05$) apoptosis (9.44%) in HT-29 cells, while extract + AuNPs treated HT-29 cells showed higher effect (16.04%) over the untreated cells (1.92%).

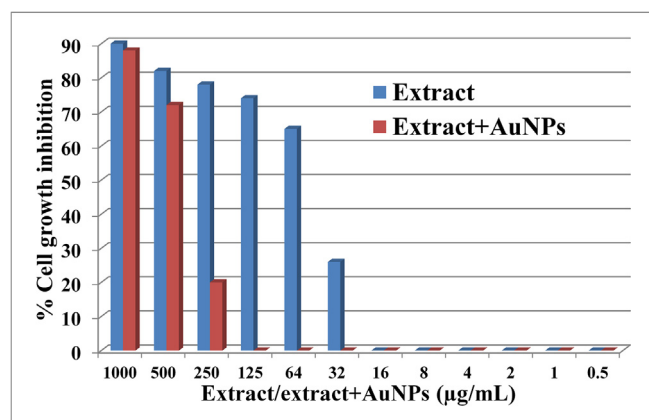


Fig. 4. Effects of *R. officinalis* extract and extract + AuNPs on HT-29 cell growth.

Table 1
Apoptotic effects of *R. officinalis* extract and extract + AuNPs on HT-29 cells.

Treatment	Apoptosis (%)			Necrosis (%)
	Total	Early	Late	
Extract	9.44	1.52	6.31	1.61
Extract + AuNPs	16.04	6.17	8.21	1.66

4. Discussion

The use of therapeutic medicinal plants to fight cancer is a good approach. *R. officinalis* is an edible and also used in cosmetics. In the current work, we targeted the valuation of the biological effects *R. officinalis* when combined with gold nanoparticles. In addition, we investigated the impact of the extract on HT-29 cells.

In the current study, we utilized *R. officinalis* leaf extract to create AuNPs. The biomolecules contained in the extract, and shown by RTIR analysis, could reduce and cap gold ions forming the AuNPs. Many studies also reported the presence of active biomolecules in *R. officinalis* leaves (Cheung and Tai, 2007; Wang et al., 2012; González-Vallinas et al., 2015; Moore et al., 2016). The created AuNPs appeared as spheres of average size of 79 nm. Many researchers were able to create nanoparticles with diverse sizes utilizing *Rosmarinus officinalis* leaf extracts (Ghaedi et al., 2015; Soltanabad et al., 2018; Hadi Soltanabad et al., 2020).

Typically, the cell cycle of cells passes through four consecutive stages starting from the quiescence stage (G_0) to the propagation (G_1 , S, G_2 , and M) stage, and return back to either the G_0 or G_1 stage (Jingwen et al., 2017). Genes controlling the cell cycle are always mutated in cancers, causing uncontrolled cell division and tumor development (Williams and Stoeber, 2012). Abnormal (cancer) cells start over G_1 straightforwardly after M stage leading to abnormal cell division. The target of anticancer medicines is to induce the cell cycle arrest at the M phase (American Cancer Society, 2015).

Extract and extract + AuNPs prepared in the current work demonstrated growth suppressive impacts on human colon cancer HT-29 cells. Vasanth et al. (2014) stated that nanoparticles included in *R. officinalis* extract had the power to enhance apoptosis in cervical cancer cells through the elevation in ROS levels and its subsequent action. Extract and extract + AuNPs revealed non-toxic potential or cell cycle interference impacts. This distinctly mentions that extract + AuNPs interferes the cell cycle of cancer cells.

Programmed cell death or apoptosis works efficiently in the equilibrium of healthy cells by removing the cancer cells (Levine et al., 2001; Wang et al., 2015). It is documented that the bioactive materials exist in *R. officinalis* have the power to initiate apoptosis in cells of the tumor. Bioactive materials exist in the plant's extract (e.g. carnosol, carnosic & rosmarinic acids) have been shown to induce apoptosis in cells of the tumor, possibly through the synthesis of nitric oxide (Đilas et al., 2012; Tai et al., 2012; Kontogianni et al., 2013; Petiwala et al., 2013). p53, a tumor suppressor, exerts a critical part in the apoptosis inducement and affects the mitochondrial intrinsic apoptosis pathway (Vaseva and Moll 2009; Nieminen et al., 2013).

In the current study, HT-29 cells were arrested by the extract and extract + AuNPs at G_2/M stages. Other *in vitro* investigations utilized several cancer cell lines (DLD-1, CaCo-2, SW480, and SW620 colon cancer cells) concluded that *R. officinalis* has anticancer characteristics (Slameová et al., 2002; Yi and Wetzstein 2011; González-Vallinas et al., 2013).

5. Conclusion

R. officinalis leaf acetone extract could synthesize AuNPs with 79 nm diameter. Extract and extract + AuNPs could arrest HT-29 cancer cell proliferation. Extract and extract + AuNPs actuated apoptosis in cancer cells as opposed to necrosis. The edible plant *R. officinalis* can be utilized in the preparation of anticancer formulas, at least against colon tumors, either alone or in combination with gold nanoparticles.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jksus.2022.102304>.

References

- Abdel-Wahhab, K.G.E.D., El-Shamy, K.A., El-Beih, N.A.E.Z., et al., 2011. Protective effect of a natural herb (*Rosmarinus officinalis*) against hepatotoxicity in male albino rats. *Comun. Sci.* 2, 9–17.
- American Cancer Society (2015) Chemotherapy Drugs: How They Work Understanding the life cycle of a cell. *Am Cancer Soc* 17, 22/10/2015.
- Balakrishna, A., Kumar, M.H., 2015. Evaluation of synergetic anticancer activity of berberine and curcumin on different models of A549, Hep-G2, MCF-7, Jurkat, and K562 cell lines. *Biomed Res. Int.* 2015, 1–7. <https://doi.org/10.1155/2015/354614>.
- Borges, R.S., Ortiz, B.L.S., Pereira, A.C.M., Keita, H., Carvalho, J.C.T., 2019. Rosmarinus officinalis essential oil: A review of its phytochemistry, anti-inflammatory activity, and mechanisms of action involved. *J. Ethnopharmacol.* 229, 29–45.
- Cheung, S., Tai, J., 2007. Anti-proliferative and antioxidant properties of rosemary *Rosmarinus officinalis*. *Oncol. Rep.* 17, 1525–1531. <https://doi.org/10.3892/or.17.6.1525>.
- Corrêa Dias, P., Foglio, M.A., Possenti, A., De Carvalho, J.E., 2000. Antiulcerogenic activity of crude hydroalcoholic extract of *Rosmarinus officinalis* L. *J. Ethnopharmacol.* 69, 57–62. [https://doi.org/10.1016/S0378-8741\(99\)00133-6](https://doi.org/10.1016/S0378-8741(99)00133-6).
- Da Rocha, A.B., Lopes, R.M., Schwartzmann, G., 2001. Natural products in anticancer therapy. *Curr. Opin. Pharmacol.* 1, 364–369. [https://doi.org/10.1016/S1471-4892\(01\)00063-7](https://doi.org/10.1016/S1471-4892(01)00063-7).
- De Melo, G.A.N., Grespan, R., Fonseca, J.P., et al., 2011. *Rosmarinus officinalis* L. essential oil inhibits in vivo and in vitro leukocyte migration. *J. Med. Food* 14, 944–949. <https://doi.org/10.1089/jmf.2010.0159>.
- Del Baño, M.J., Castillo, J., Benavente-García, O., Lorente, J., Martín-Gil, R., Acevedo, C., Alcaraz, M., 2006. Radioprotective-antimutagenic effects of rosemary phenolics against chromosomal damage induced in human lymphocytes by γ -rays. *J. Agric. Food Chem.* 54 (6), 2064–2068.
- Dilas, S., Knez, Ž., Četojević-Simin, D., Tumbas, V., Škerget, M., Čanadanović-Brunet, J., Četković, G., 2012. In vitro antioxidant and antiproliferative activity of three rosemary (*Rosmarinus officinalis* L.) extract formulations. *Int. J. Food Sci. Technol.* 47 (10), 2052–2062.
- Dykman, L.A., Khebtsov, N.G., 2017. Immunological properties of gold nanoparticles. *Chem. Sci.* 8, 1719–1735. <https://doi.org/10.1039/C6SC03631G>.
- Einbond, L.S., Wu, H.-a., Kashiwazaki, R., He, K., Roller, M., Su, T., Wang, X., Goldsberry, S., 2012. Carnosic acid inhibits the growth of ER-negative human breast cancer cells and synergizes with curcumin. *Fitoterapia* 83 (7), 1160–1168.
- Fernández, L.F., Palomino, O.M., Frutos, G., 2014. Effectiveness of *Rosmarinus officinalis* essential oil as antihypertensive agent in primary hypotensive patients and its influence on health-related quality of life. *J. Ethnopharmacol.* 151, 509–516. <https://doi.org/10.1016/j.jep.2013.11.006>.
- Ganesan, T., Sinniah, A., Chik, Z., Alshawsh, M.A., 2020. Punicalagin regulates apoptosis-autophagy switch via modulation of annexin a1 in colorectal cancer. *Nutrients* 12, 1–17. <https://doi.org/10.3390/nu12082430>.
- Ghaedi, M., Yousefinejad, M., Safarpour, M., Khafri, H.Z., Purkait, M.K., 2015. *Rosmarinus officinalis* leaf extract mediated green synthesis of silver nanoparticles and investigation of its antimicrobial properties. *J. Ind. Eng. Chem.* 31, 167–172.
- Ghranh, H.A., Khan, K.A., Ibrahim, E.H., Setzer, W.N., 2019. Synthesis of gold nanoparticles (AuNPs) using ricinus communis leaf ethanol extract, their characterization, and biological applications. *Nanomaterials* 9 (5), 765.
- Ghranh, H.A., Ibrahim, E.H., Ahmad, Z., 2021. Antimicrobial, immunomodulatory and cytotoxic activities of green synthesized nanoparticles from *Acacia honey* and *Calotropis procera*. *Saudi J. Biol. Sci.* 28, 3367–3373. <https://doi.org/10.1016/j.sjbs.2021.02.085>.
- Gonçalves, G.A., Corrêa, R.C.G., Barros, L., Dias, M.I., Calheta, R.C., Correa, V.G., Bracht, A., Peralta, R.M., Ferreira, I.C.F.R., 2019. Effects of in vitro gastrointestinal digestion and colonic fermentation on a rosemary (*Rosmarinus officinalis* L.) extract rich in rosmarinic acid. *Food Chem.* 271, 393–400.
- González-Vallinas, M., Molina, S., Vicente, G., de la Cueva, A., Vargas, T., Santoyo, S., García-Risco, M.R., Fornari, T., Reglero, G., Ramírez de Molina, A., 2013. Antitumor effect of 5-fluorouracil is enhanced by rosemary extract in both drug sensitive and resistant colon cancer cells. *Pharmacol. Res.* 72, 61–68.
- González-Vallinas, M., Reglero, G., Ramírez De Molina, A., 2015. Rosemary (*Rosmarinus officinalis* L.) extract as a potential complementary agent in anticancer therapy. *Nutr. Cancer* 67, 1223–1231.
- Habtmeriam, S., 2016. The therapeutic potential of rosemary (*Rosmarinus officinalis*) Diterpenes for Alzheimer's disease. *Evid.-based Complement. Altern. Med.* 2016, 1–14. <https://doi.org/10.1155/2016/2680409>.
- Hadi Soltanabad, M., Bagherieh-Najjar, M.B., Mianabadi, M., 2020. Carnosic acid content increased by silver nanoparticle treatment in rosemary (*Rosmarinus officinalis* L.). *Appl. Biochem. Biotechnol.* 191, 482–495. <https://doi.org/10.1007/s12010-019-03193-w>.
- Hamidpour, R., 2017. *Rosmarinus officinalis* (Rosemary): A novel therapeutic agent for antioxidant, antimicrobial, anticancer, antidiabetic, antidepressant, neuroprotective, anti-inflammatory, and anti-obesity treatment. *Biomed. J. Sci. Tech. Res.* 1 (1–6). <https://doi.org/10.26717/bjstr.2017.01.000371>.
- Hanahan, D., Weinberg, R.A., 2011. Hallmarks of cancer: the next generation. *Cell* 144, 646–674. <https://doi.org/10.1016/j.cell.2011.02.013>.
- Ibrahim, E.H., Ghranh, H.A., Alshehri, A., Kilany, M., Khalofah, A., El-Mekawy, H.I., Sayed, M.A., Alotheid, H., Taha, R., 2021. *Lepidium sativum* and its biogenic silver nanoparticles activate immune cells and induce apoptosis and cell cycle arrest in HT-29 colon cancer cells. *Biomater. Tissue Eng* 11 (2), 195–209.
- Jingwen, B., Yaochen, L., Guojun, Z., 2017. Cell cycle regulation and anticancer drug discovery. *Cancer Biol Med* 14 (348). <https://doi.org/10.20892/j.issn.2095-3941.2017.0033>.
- Kontogianni, V.G., Tomic, G., Nikolic, I., Nerantzaki, A.A., Sayyad, N., Stosic-Grujicic, S., Stojanovic, I., Gerothanassis, I.P., Tzakos, A.G., 2013. Phytochemical profile of *Rosmarinus officinalis* and *Salvia officinalis* extracts and correlation to their antioxidant and anti-proliferative activity. *Food Chem.* 136 (1), 120–129.
- Levine, A., Belenghi, B., Damari-Weisler, H., Granot, D., 2001. Vesicle-associated membrane protein of arabisopsis suppresses bax-induced apoptosis in yeast downstream of oxidative burst. *J. Biol. Chem.* 276, 46284–46289. <https://doi.org/10.1074/jbc.M107375200>.
- Machado, D.G., Cunha, M.P., Neis, V.B., et al., 2012. *Rosmarinus officinalis* L. hydroalcoholic extract, similar to fluoxetine, reverses depressive-like behavior without altering learning deficit in olfactory bulbectomized mice. *J. Ethnopharmacol.* 143, 158–169. <https://doi.org/10.1016/j.jep.2012.06.017>.
- Mohamed, M.M., Fouad, S.A., Elshoky, H.A., Mohammed, G.M., Salaheldin, T.A., 2017. Antibacterial effect of gold nanoparticles against *Corynebacterium pseudotuberculosis*. *Int. J. Vet. Sci. Med.* 5 (1), 23–29.
- Moore, J., Yousef, M., Tsiani, E., 2016. Anticancer effects of rosemary (*Rosmarinus officinalis* L.) extract and rosemary extract polyphenols. *Nutrients* 8 (731). <https://doi.org/10.3390/nu8110731>.
- Naimi, M., Vlavcheski, F., Shamshoum, H., Tsiani, E., 2017. Rosemary extract as a potential anti-hyperglycemic agent: Current evidence and future perspectives. *Nutrients* 9, 1–19.
- Nieminen, A.I., Eskelinen, V.M., Haikala, H.M., Tervonen, T.A., Yan, Y., Partanen, J.I., Klefström, J., 2013. Myc-induced AMPK-phospho p53 pathway activates Bak to sensitize mitochondrial apoptosis. *Proc. Natl. Acad. Sci. U S A* 110 (20). <https://doi.org/10.1073/pnas.1208530110>.
- Nieto, G., Ros, G., Castillo, J., 2018. Antioxidant and antimicrobial properties of rosemary (*Rosmarinus officinalis* L.): A review. *Medicines* 5 (98). <https://doi.org/10.3390/medicines5030098>.
- Nolkemper, S., Reichling, J., Stintzing, F., Carle, R., Schnitzler, P., 2006. Antiviral effect of aqueous extracts from species of the Lamiaceae family against Herpes simplex virus type 1 and type 2 in vitro. *Planta Med.* 72 (15), 1378–1382.
- Petiwal, S.M., Puthenveetil, A.G., Johnson, J.J., 2013. Polyphenols from the Mediterranean herb rosemary (*Rosmarinus officinalis*) for prostate cancer. *Front. Pharmacol.* 4, 1–4. <https://doi.org/10.3389/fphar.2013.00029>.
- Posadas, S.J., Caz, V., Largo, C., De la Gándara, B., Matallanas, B., Reglero, G., De Miguel, E., 2009. Protective effect of supercritical fluid rosemary extract, *Rosmarinus officinalis*, on antioxidants of major organs of aged rats. *Exp. Gerontol.* 44 (6–7), 383–389.
- Rajeshkumar, S., 2016a. Synthesis of silver nanoparticles using fresh bark of *Pongamia pinnata* and characterization of its antibacterial activity against gram positive and gram negative pathogens. *Resour. Technol.* 2, 30–35. <https://doi.org/10.1016/j.refit.2016.06.003>.
- Rajeshkumar, S., 2016b. Anticancer activity of eco-friendly gold nanoparticles against lung and liver cancer cells. *J. Genet. Eng. Biotechnol.* 14, 195–202. <https://doi.org/10.1016/J.JGEB.2016.05.007>.
- Rašković, A., Milanović, I., Pavlović, N., et al., 2014. Antioxidant activity of rosemary (*Rosmarinus officinalis* L.) essential oil and its hepatoprotective potential. *BMC Complement. Altern. Med.* 14 (225). <https://doi.org/10.1186/1472-6882-14-225>.
- Slameová, D., Kubošková, K., Horváthová, E., Robichová, S., 2002. Rosemary-stimulated reduction of DNA strand breaks and FPG-sensitive sites in mammalian cells treated with H2O2 or visible light-excited Methylene Blue. *Cancer Lett.* 177, 145–153. [https://doi.org/10.1016/S0304-3835\(01\)00784-4](https://doi.org/10.1016/S0304-3835(01)00784-4).
- Soltanabad, M.H., Bagherieh-Najjar, M.B., Baghkeirati, E.K., Mianabadi, M., 2018. Ag-conjugated nanoparticle biosynthesis mediated by Rosemary leaf extracts

- corre-lates with plant antioxidant activity and pro-teín content. *Int. J. Nanosci. Nanotechnol.* 14, 319–325.
- Sotelo-Félix, J.I., Martínez-Fong, D., Muriel, P., Santillán, R.L., Castillo, D., Yahuaca, P., 2002. Evaluation of the effectiveness of *Rosmarinus officinalis* (Lamiaceae) in the alleviation of carbon tetrachloride-induced acute hepatotoxicity in the rat. *J. Ethnopharmacol.* 81 (2), 145–154.
- Tabassum, N., Hamdani, M., 2014. Plants used to treat skin diseases. *Pharmacogn. Rev.* 8, 52–60.
- Tai, J., Cheung, S., Wu, M., Hasman, D., 2012. Antiproliferation effect of Rosemary (*Rosmarinus officinalis*) on human ovarian cancer cells in vitro. *Phytomedicine* 19, 436–443. <https://doi.org/10.1016/j.phymed.2011.12.012>.
- Vasanth, K., Ilango, K., MohanKumar, R., Agrawal, A., Dubey, G.P., 2014. Anticancer activity of *Moringa oleifera* mediated silver nanoparticles on human cervical carcinoma cells by apoptosis induction. *Colloids Surfaces B Biointerfaces* 117, 354–359.
- Vaseva, A.V., Moll, U.M., 2009. The mitochondrial p53 pathway. *Biochim. Biophys. Acta – Bioenerg.* 1787, 414–420. <https://doi.org/10.1016/j.bbabi.2008.10.005>.
- Wang, Y., Liu, C., Luo, M., et al., 2015. Chemotherapy-induced miRNA-29c/catenin- δ signaling suppresses metastasis in gastric cancer. *Cancer Res.* 75, 1332–1344. <https://doi.org/10.1158/0008-5472.CAN-14-0787>.
- Wang, W., Li, N., Luo, M., et al., 2012. Antibacterial activity and anticancer activity of *Rosmarinus officinalis* L. essential oil compared to that of its main components. *Molecules* 17, 2704–2713. <https://doi.org/10.3390/molecules17032704>.
- Williams, G.H., Stoeber, K., 2012. The cell cycle and cancer. *J. Pathol.* 226, 352–364. <https://doi.org/10.1002/path.3022>.
- Xu, G.L., Geng, D., Xie, M., et al., 2015. Chemical composition, antioxidative and anticancer activities of the essential oil: *Curcuma rhizoma*-*sparganii rhizoma*, a traditional herb pair. *Molecules* 20, 15781–15796. <https://doi.org/10.3390/molecules200915781>.
- Yamamoto, J., Yamada, K., Naemura, A., Yamashita, T., Arai, R., 2005. Testing various herbs for antithrombotic effect. *Nutrition* 21 (5), 580–587.
- Yi, W., Wetzstein, H.Y., 2011. Anti-tumorigenic activity of five culinary and medicinal herbs grown under greenhouse conditions and their combination effects. *J. Sci. Food Agric.* 91, 1849–1854. <https://doi.org/10.1002/jsfa.4394>.