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Journal of King Saud University – Science

journal homepage: www.sciencedirect.com

Original article

Seroprevalence of neutralizing antibodies to severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) among healthcare workers in Makkah, Saudi Arabia



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ARTICLE INFO

Article history:

Received 14 December 2020

Revised 12 January 2021

Accepted 5 February 2021

Available online 13 February 2021

Keywords:

COVID-19

SARS-CoV-2

Seroprevalence

Healthcare workers

Saudi Arabia

ABSTRACT

Objective: The new coronavirus disease 2019 (COVID-19) is a major health problem worldwide. The surveillance of seropositive individuals serves as an indicator to the extent of infection spread and provides an estimation of herd immunity status among population. Reports from different countries investigated this issue among healthcare workers (HCWs) who are “at risk” and “sources of risk” for COVID-19. This study aims to investigate the seroprevalence of COVID-19 among HCWs in one of the COVID-19 referral centers in Makkah, Saudi Arabia using three different serological methods.

Methods: In-house developed enzyme-linked immunoassay (ELISA), commercially available electrochemiluminescence immunoassay (ECLIA), and microneutralization (MN) assay were utilized to determine the seroprevalence rate among the study population. 204 HCWs participated in the study. Both physicians and nurses working in the COVID-19 and non COVID-19 areas were included. Twelve out of 204 were confirmed cases of COVID-19 with variable disease severity. Samples from recovered HCWs were collected four weeks post diagnosis.

Results: The overall seroprevalence rate was 6.3% (13 out of 204) using the in-house ELISA and MN assay and it was 5.8% (12 out of 204) using the commercial ECLIA. Among HCWs undiagnosed with COVID-19,

Abbreviations: COVID-19, The new Coronavirus Disease 2019; DMEM-FCS, Dulbecco's Modified Eagle Medium containing fetal calf serum; ELISA, Enzyme-linked immunoassay; ECLIA, Electro-chemiluminescence immunoassay; HCL, 1N hydrochloric acid; HCWs, Healthcare Workers; MN, Microneutralization; OD₄₅₀, Optical density at 450 nm; PBS, Phosphate Buffer Saline; PBST, PBS containing 0.1% Tween 20; SARS-CoV-2, Severe Acute Respiratory Syndrome Coronavirus 2; SFH, Security Forces Hospital; TCID₅₀, Median Tissue Culture Infectious Dose; TMB, 3,3',5,5'-Tetramethylbenzidine; WHO, World Health Organization.

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Peer review under responsibility of King Saud University.



<https://doi.org/10.1016/j.jksus.2021.101366>

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the seroprevalence was 2% (4 out 192). Notably, neutralizing antibodies were not detected in 3 (25%) out 12 confirmed cases of COVID-19.

Conclusions: Our study, similar to the recent national multi-center study, showed a low seroprevalence of SARS-CoV-2 antibodies among HCWs. Concordance of results between the commercial electrochemiluminescence immunoassay (ECLIA), in-house ELISA and MN assay was observed. The in-house ELISA is a promising tool for the serological diagnosis of SARS-CoV-2 infection. However, seroprevalence studies may underestimate the extent of COVID-19 infection as some cases with mild disease did not have detectable antibody responses.

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1. Introduction

In December 2019, a viral-induced pneumonia outbreak caused by a novel betacoronavirus was reported in Wuhan, China. The virus and disease have been named severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and the coronavirus disease of 2019 (COVID-19), respectively (WHO, 2020c, 2020d). The clinical spectrum of COVID-19 ranges widely in term of severity. While some infected individuals experience no or very mild symptoms, others develop life-threatening complications (Hoehl et al., 2020; Huang et al., 2020; Wang et al., 2020). Acute Respiratory Distress Syndrome (ARDS) is a common life-threatening complication of SARS-CoV-2 infection (Huang et al., 2020; Xu et al., 2020; Zhou et al., 2020). Extrapulmonary organ dysfunction is another major complication that may lead to death of COVID-19 patients (Cheng et al., 2020; Xiao et al., 2020). The rapid human-to-human transmission of the virus by respiratory droplets and direct contact with patients led to COVID-19 pandemic (Lam et al., 2020; WHO, 2020; Wu et al., 2020a, 2020b). What intensifies the problem is that both symptomatic and asymptomatic individuals can be sources of infection (Nikolai et al., 2020). By the end of December 2020, Over 80 million COVID-19 cases and 1.7 million deaths have been reported (Who, 2020b).

Many countries, including Saudi Arabia, have initially implemented control measures to contain the spread of infection but they are now the process of returning to “normal” life (Alandijany et al., 2020b; Ebrahim et al., 2020; MOH-KSA, 2020). Hence, it is expected that SARS-CoV-2 will continue to spread among societies. Healthcare workers (HCWs) and family members of infected individuals are at increased risk of being exposed to SARS-CoV-2 infection (Jin et al., 2020). According to the WHO, 14% of total global cases are related to health workers (Who, 2020a). Systematic screening for the seroprevalence of anti-SARS-CoV-2 antibodies is a valuable indicator for the spread of the infection (Peto, 2020). Several recent studies have investigated the prevalence of antibodies specifically directed against SARS-CoV-2 among HCWs with variable findings (Chen et al., 2020b; Garcia-Basteiro et al., 2020; Gómez-Ochoa et al., 2020; Hunter et al., 2020; Korth et al., 2020). A study conducted on 734 HCWs at Indiana University Health, USA revealed 1.6% prevalence rate of SARS-CoV-2 IgG antibodies (Hunter et al., 2020). Similar rate was reported from a study conducted on 316 individuals working in the University Hospital Essen, Germany (Korth et al., 2020). Another assessment conducted on the Hospital Clinic in Barcelona demonstrated 6.23%, 7.61%, and 8.13% seroprevalence rates of IgM, IgG, and IgA, respectively, among 578 HCWs (Garcia-Basteiro et al., 2020). The prevalence rate was considerably higher (17.14%) in an independent study, which investigated the prevalence of neutralizing antibody to SARS-CoV-2 among 105 HCWs who have been in close contact with COVID-19 patients in Nanjing Drum Tower Hospital, China (Chen et al., 2020b). A systematic review and meta-analysis of 97 reports from different countries estimated

11% and 7% prevalence rates of SARS-CoV-2 infection and antibodies, respectively, among HCWs (Gómez-Ochoa et al., 2020). A recent national multi-center study in Saudi Arabia showed 2.36% seroprevalence of SARS-CoV-2 antibodies among HCWs. However, there was discordant results between the commercial electrochemiluminescence immunoassay (ECLIA) and the pseudotyped viral particles neutralization assay (Alserahi et al., 2020).

We have developed and optimized an in-house indirect ELISA that enable sensitive and specific detection of SARS-CoV-2 IgG antibodies (27). Microneutralization (MN) assay is the gold standard for evaluating the presence of neutralizing antibody for SARS-CoV-2 infection (GeurtsvanKessel et al., 2020; Kohmer et al., 2021; Lisboa Bastos et al., 2020). We have recently applied these two valuable techniques to investigate the seroprevalence rate among healthy blood donors during COVID-19 lockdown in the country (Alandijany et al., 2021). In this study, in-house ELISA in addition to commercially available ECLIA and microneutralization (MN) assay were utilized to determine the seroprevalence rate among HCWs at the Security Forces Hospital (SFH), Makkah, Saudi Arabia. The study site is one of the COVID-19 referral hospitals in Makkah. Our data demonstrate three key findings: (1) a low prevalence rate of anti-SARS-CoV-2 neutralizing antibody among HCWs undiagnosed with COVID-19, (2) neutralizing antibodies were not detected among some HCWs diagnosed with COVID-19 and (3) concordant results between the three serological tests (in house ELISA, commercial ECLIA and MN assay)

2. Materials and methods

2.1. Ethic statement

This study was approved by the research institutional board at SFH in Makkah, Saudi Arabia (0369–24062). Written consent forms were obtained from participants.

2.2. Study population

The total number of HCWs participated in this study was 204 comprising 192 individuals undiagnosed with COVID-19 and 12 recovered patients. Serum samples were collected during June and July 2020. All samples were collected, transported to the Special Infectious Agents Unit (SIAU), King Fahd Medical Research Center (KFMRC), King Abdulaziz University (KAU), aliquoted, and frozen at -20°C until utilized in the experiments.

2.3. Immunoassays

Sera were screened for the presence of antibody directed against SARS-CoV-2 by (1) a commercially available Elecsys[®] test system from Roche was performed according to the manufacturers' instructions. The assay principle is based on ECLIA utilizing a

modified recombinant nucleocapsid protein. (2) a recently developed in-house ELISA was conducted as previously described (27). Briefly, antigen coating of flat bottom microtiter plates (Immulon® 2 HB, USA) was performed overnight at 4 °C with 100 ng per well of SARS-CoV-2 (2019-nCoV) spike S1 + S2 ECD-His recombinant protein (Sino Biological, China). Subsequently, the plates were subjected to three washes with phosphate buffer saline (PBS) containing 0.1% Tween 20 (PBST) prior to blocking in PBST containing 5% skimmed milk for 1 h at room temperature. This step was followed by three washes with PBST. Sera were diluted at 1:100 dilutions in PBST containing 5% skimmed milk, added at 100 µl volume, and incubated for an hour at 37 °C. Following three washes with PBST, 100 µl of secondary antibody (goat KPL peroxidase-labelled antibodies to human IgG; Seracare, USA) diluted at 1:64,000 in PBST were added and allowed to incubate for an hour at 37°. The plates were subjected to three washes with PBST. Then, 100 µl of substrate (3,3',5,5'-Tetramethylbenzidine (TMB); Seracare, USA) were added for 5 min for color development prior to addition of 100 µl of 1 N hydrochloric acid (HCL) to stop the reaction. Using Elx 800 bioelisa Reader (Biokit, Spain), the optical density was measured at 450 nm (OD₄₅₀). OD₄₅₀ values of > 0.27 were considered positive. Negative and positive controls utilized in this assay were sera of a healthy blood donor and a recovered COVID-19 patient known to have neutralizing IgG antibodies, respectively. The in-house ELISA provides 100% sensitivity, 98.4% specificity, 98.8% agreement, and high overall accuracy (Alandijany et al., 2020a).

2.4. MN assay

MN assay was used as a confirmatory test to determine the presence of SARS-CoV-2-specific neutralizing antibodies. The local SARS-CoV-2 clinical isolate (SARS-CoV-2/human/SAU/85791 C/2020) (Genbank accession number MT630432.1) were utilized in this assay. The virus stock was propagated and titrated by Median Tissue Culture Infectious Dose (TCID₅₀) on African green monkey kidney cells Vero E6 (ATCC® CRL-1586™). Sera were subjected to heat inactivation at 56 °C for 30 min. Then, samples were serially diluted in Dulbecco's modified eagle medium containing 2% fetal calf serum (DMEM-FCS) and added with equal volume of DMEM-FCS containing 100 TCID₅₀ of SARS-CoV-2 on confluent cells. The cells were allowed to incubate at 37 °C in 5% CO₂ until extensive cytopathic effect was observed (typically for 3–4 days). Uninfected cells and SARS-CoV-2 infected cells in the absence of human serum were utilized as controls. MN titers of ≥ 1:20 were considered positive.

2.5. Data curation

Figure drawing, data processing and calculations were performed by GraphPad Prism software.

3. Results

3.1. Distribution of study population

The total number of participants enrolled in this study was 204 (n = 204) which comprises 192 (94.1%) undiagnosed with COVID-19 and 12 (5.9%) diagnosed with COVID-19 by reverse transcriptase polymerase chain reaction (RT-PCR). Serum samples of those diagnosed with COVID-19 were collected four weeks post diagnosis. The categorization of study population according to COVID-19 status (undiagnosed vs diagnosed), gender (male vs female), department (COVID-19 area vs other departments), and profession (physicians vs nurses vs other professions) are shown (Table 1).

Table 1
Demographic data of study population (n = 204).

Variable	n (%)
Demographics	
Male	83 (40.7%)
Female	121 (59.3%)
Age: Median (IQR)	33.5 (9)
Occupation	
Physicians	84 (41.2%)
Nurses	119 (58.3%)
Others	1 (0.5%)
Department	
COVID-19 area	93 (45.6%)
Non-COVID-19 area	36 (17.7%)
Emergency room	41 (20.1%)
Radiology	17 (8.3%)
Others	17 (8.3%)
COVID-19 status	
Undiagnosed	192 (94.1%)
Diagnosed	12 (5.9%)

3.2. The overall seroprevalence rate of anti-SARS-CoV-2 IgG antibody

Utilizing in-house and commercially available immunoassays, serum samples from HCWs (n = 204) were assessed for the presence of IgG antibody to SARS-CoV-2 infection. The cut-off values for the in-house ELISA and Elecsys® test system are > 0.27 and ≥ 1, respectively. The in-house assay identified 13 seropositive samples while 12 samples tested “reactive” by Elecsys® test system (Fig. 1). Next, all samples were subjected to MN assay which represents the gold standard method for evaluation of the presence of virus-specific IgG neutralizing antibodies with MN titer of ≥ 1:20 considered positive. There was a 100% match between the results of in-house ELISA and MN assay. The MN titer of positive samples (n = 13) is shown (Fig. 2). Collectively, this data demonstrates an overall seroprevalence rate of 6.3% among the study population.

3.3. Sero-positive samples categorized according to COVID-19 status

Among the 13 seropositive samples, 9 HCWs are confirmed COVID-19 cases and 4 cases are undiagnosed previously with COVID-19. Nine cases of confirmed COVID-19, whether they were asymptomatic or had pneumonia, developed neutralizing antibody titers between 1:160 and 1:1280. Three cases of confirmed COVID-19 who had upper respiratory tract infection did not mount a neutralizing antibody response. Among the 4 undiagnosed COVID-19 cases, 50% of them worked in the COVID-19 area and they had neutralizing antibody titers between 1:320 and 1:640 (Fig. 3 and Table 2).

4. Discussion

COVID-19 is a global crisis caused by SARS-CoV-2. Officials in Saudi Arabia have implemented timely and unprecedented measures to contain the spread of the infection (Alandijany et al., 2020b; Algaissi et al., 2020; Dalia et al., 2020). As of now (January 2021), many COVID-19 restrictions have been relaxed with enforcement of maintaining physical distancing and mask wearing (MOH-KSA, 2020). At this stage and through the post-pandemic phase, it is important to assess the virus circulation and evaluate the seroprevalence of neutralizing antibody among the community (Peto, 2020; Sunjaya and Sunjaya, 2020). Few reports addressed the seroprevalence status among the Saudi population (Alandijany et al., 2021; Alserahi et al., 2020). In this study, we

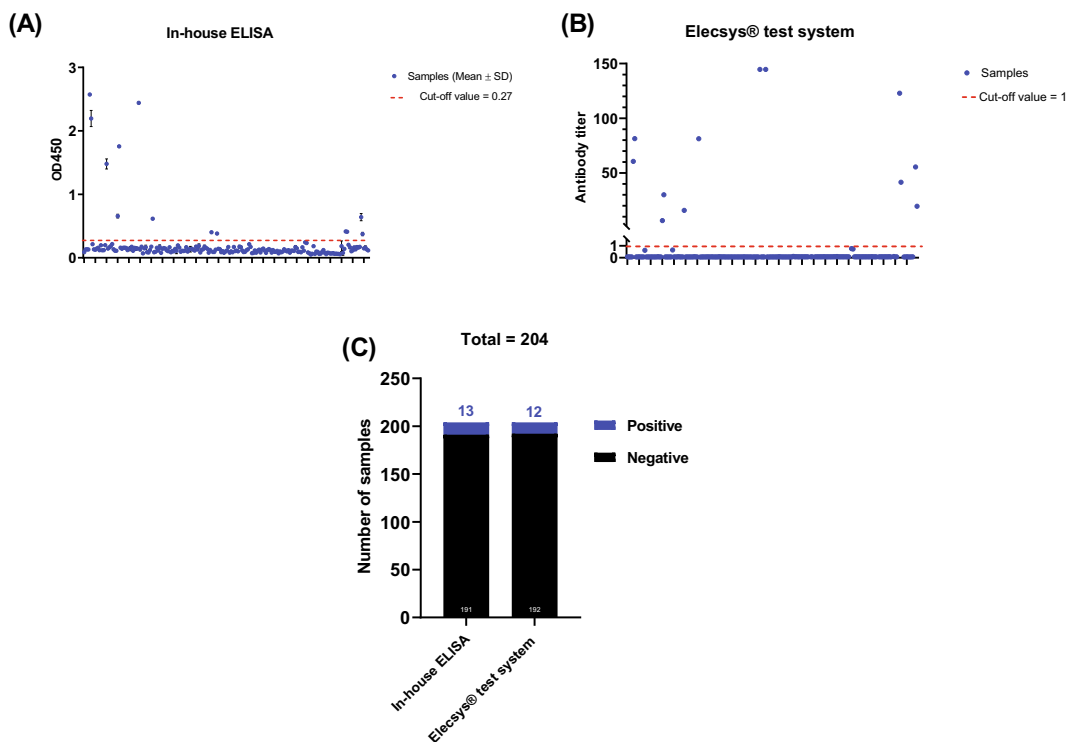


Fig. 1. The overall seroprevalence rate of anti-SARS-CoV-2 IgG antibody among healthcare workers by ELISAs. (A) Results obtained from in-house ELISA. The means and standard deviations of optical density values at 450 nm (OD₄₅₀) are shown. The cut-off value of the assay is 0.27. (B) Results obtained from Elecsys[®] test system. The antibody titer for each sample is shown. The cut-off value of the assay is ≥ 1 . (C) The numbers of positive (blue) and negative (black) samples obtained from in-house ELISA and Elecsys[®] test system.

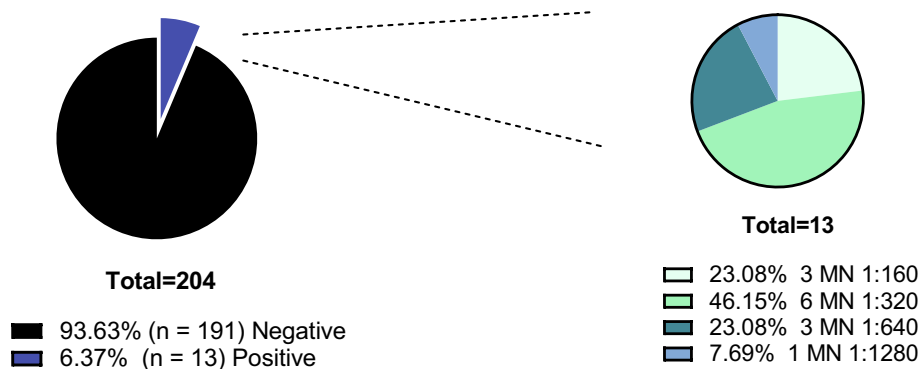


Fig. 2. Assessment of neutralizing antibody titers among healthcare workers by micro-neutralization (MN) assay. (A) The seroprevalence rate of anti-SARS-CoV-2 neutralizing antibody among healthcare workers by MN assay. (B) MN titers of sero-positive samples. The actual numbers and relative percentages (%) are shown.

have evaluated the seroprevalence rate among HCWs at the SFH, one of the COVID-19 referral centers in Makkah, Saudi Arabia.

The overall seroprevalence of SARS-CoV-2 antibodies among HCWs was 6.3% (13 out 204) and 2% among those undiagnosed previously with disease (4 out 192) (Fig. 1, Fig. 3 and Table 2). Other studies have described varied results depending on the inclusion or exclusion of confirmed COVID-19 cases in the analysis (Gómez-Ochoa et al., 2020). The low SARS-CoV-2 antibodies seroprevalence in our study was comparable to the national multi-center study showing 2.36% seropositivity and it is suggestive of effective infection control measures among HCW (Alserahi et al., 2020). Knowing these results, a significant number of HCWs will be at risk of COVID-19 infection. With the limited supply of vaccination, HCWs in Makkah who are exposed to pilgrims should have a priority in vaccination programs (Abd El Ghany et al., 2016).

The recent national multi-center study utilized a commercially available chemiluminescent microparticle assay to screen sera of HCWs (Alserahi et al., 2020). Only 100 positive samples were subjected to SARS-CoV-2 pseudotyped viral particles neutralization assay. Results showed that 8% of these samples lacked neutralizing activity (Alserahi et al., 2020). In our study, we performed three tests; in house ELISA, commercial ECLIA and MN assay in all 204 cases (Fig. 1 and Fig. 2). There were concordant results in all 13 seropositive cases using the in house ELISA and MN assay which corresponds to the reported sensitivity and specificity of our developed assay (Alandijany et al., 2020a). The in-house ELISA is a promising serological test for the diagnosis of COVID-19 infection. Twelve out of 13 cases were positive using the Elecsys[®] test system (Fig. 1 and Fig. 3). The single false negative result obtained from this assay, in addition to several recent reports, highlight the

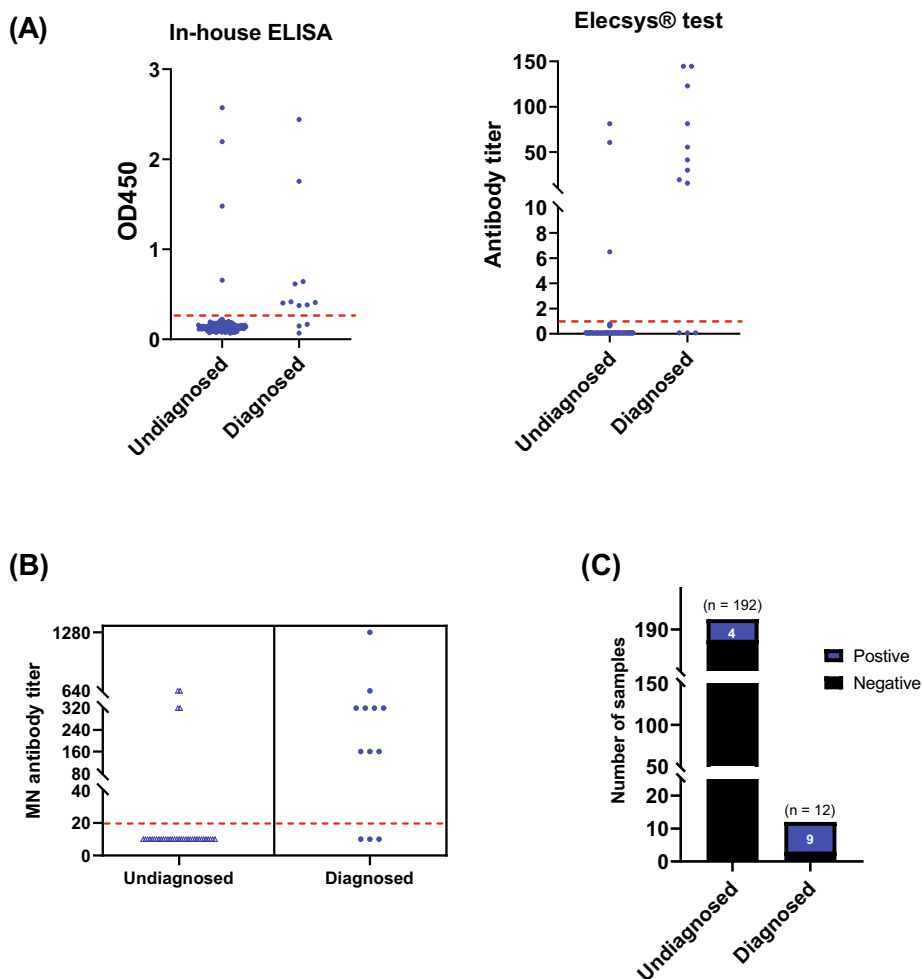


Fig. 3. Categorization of sero-positive healthcare workers according to their COVID-19 status. (A) Results obtained from in-house (left panel) and Elecsys® test system (right panel) ELISAs. Dashed red lines represent the cut-off values for each assay. (B) Results obtained from MN assay. MN titers of each positive sample is shown. (C) The number of positive (blue) and negative (black) samples categorized according to COVID-19 status.

Table 2
Categorization of sero-positive healthcare workers according to their demographic data.

COVID-19 status	Category	Sub-category	n (%)
Undiagnosed(n = 4 out of 192; 2%)	Gender	Male	1 (25%)
		Female	3 (75%)
	Department	COVID-19	2 (50%)
		Non-COVID-19	2 (50%)
	Profession	Physician	1 (25%)
		Nurse	2 (50%)
Diagnosed*(n = 9 out of 12, 75%)	Gender	Male	6 (66.6%)
		Female	3 (33.3%)
	Clinical Manifestations	Asymptomatic	5 (55.5%)
		URTI	0 (0%)
		Pneumonia	4(44.4%)
		Severe Pneumonia	0 (0%)
	Department	COVID-19	3 (33.3%)
		NON COVID-19	6 (66.6%)
	Profession	Physician	5 (55.5%)
		Nurse	1 (11.1%)
		Other professions	3 (33.3%)

importance of evaluating the performance of commercially available kits prior to utilizing them for diagnostic and epidemiological applications (GeurtsvanKessel et al., 2020; Lisboa Bastos et al., 2020).

Among COVID-19 confirmed HCW, 9 out of 12 cases had detectable neutralizing antibodies (Fig. 3 and Table 2). As previously reported, patients with pneumonia tended to have higher neutralizing antibodies compared to cases with asymptomatic infection

(Ko et al., 2020; Kohmer et al., 2021). Three Patients with upper respiratory tract infections did not mount a neutralizing antibody response (Fig. 3 and Table 2). Some reports described lack of humeral response in cases of mild infection (Chen et al., 2020a; Gattinger et al., 2021; Melgaço et al., 2020). In addition, other reports showed waning of neutralizing antibodies over time (Beaudoin-Bussièrès et al., 2020; Robbiani et al., 2020). A real concern is the increasing reports of COVID-19 re-infection (AlFehaidi et al., 2020; Dao et al., 2020). HCWs working in Makkah who are exposed to pilgrims are at risk of respiratory tract infections (Yezli et al., 2019). Hence, further studies are required to assess their vulnerability to recurrent COVID-19 infection.

5. Conclusions

The seroprevalence of neutralizing antibodies to SARS-CoV-2 infection among HCWs in our study was low and it was comparable to the national multi-center study. Our findings are suggestive of effective infection control measures among HCW. However, a significant number of HCWs are still at risk of COVID-19 and should have the priority for future vaccination program, particularly those working in Makkah and frequently exposed to the pilgrims. In addition, our study showed that the performance of our in-house ELISA was comparable to the MN assay validating it as a valuable serological test for the diagnosis of COVID-19. The lack of antibody response among some HCWs with mild infection, raises the need for more studies to evaluate the risk of future recurrent infections.

6. Disclosure of funding

The Author Acknowledgments The Financial Support Provided By King Abdulaziz City for Science and Technology (General Directorate for Research & Innovation Support (GDRIS)) To King Abdulaziz University To Implement This Work Through Fast Track Program For COVID-19 Research Project No. 5–20-01-009-0080.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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