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Seasonal pattern of vitamin D hydroxyl metabolite concentrations and their association with cardiac medications – An observational study



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ABSTRACT

Objectives: The sun is the primary source of vitamin D production, and seasonal changes can substantially influence vitamin D levels, which hardly any study has examined in cardiac patients. This research aims to measure 25(OH)D₃ and 25(OH)D₂ concentrations and assess their relation to the seasons and medications given to cardiac patients.

Methods: We collected 116 blood samples from 58 patients in autumn-winter and spring-summer seasons. Ultra-high-performance liquid chromatography-tandem mass spectrometry method was applied to determine 25(OH)D₃ and 25(OH)D₂ concentrations. The statistical assessment was performed using Statistica 13.3 with Plus Kit 3.0.

Results: Only 9% of patients had 25(OH)D₃ concentrations in the recommended range of 30–50 ng/mL. We found significantly higher 25(OH)D₃ average levels in spring-summer period compared with autumn-winter period ($P = 0.001$). Older patients had a higher risk of vitamin D deficiency in autumn-winter (OR = 1.08; $P = 0.011$, OR = 0.32; $P = 0.015$, respectively). Average 25(OH)D₂ concentrations between seasons were insignificant ($Z = 1.04$; $P = 0.3$). Vitamin D deficiency was significantly correlated with administration of angiotensin-II receptor blocker (OR = 7.49; $P = 0.025$), steroidal antiandrogen*age (OR = 1.039; $P = 0.022$). Other medications did not correlate with vitamin D deficiency, 9%NaCl (OR = 0.2; $P = 0.04$) and thiazide (0.076; $P = 0.015$).

Conclusion: The prevalence of vitamin D deficiency and therapeutic drug monitoring are substantial in the observed group of cardiovascular patients, considering those with higher risk factors. Cardiac patients may benefit from vitamin D supplementation or dietary intervention to correct vitamin D levels. Further studies on more patients are required to confirm our results and identify other factors influencing 25(OH)D concentrations in cardiac patients.

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1. Introduction

Vitamin D is a fat-soluble vitamin that functions and chemically resembles steroid hormones. It has two functionally inactive prohormones (vitamin D₂ or ergocalciferol) and (vitamin D₃ or cholecalciferol) (Bikle, 2014). Vitamin D₂ is obtained from the diet after

synthesis in the plants and fungi via ultraviolet-B (UVB) irradiation of ergosterol (Anderson, 1999). Besides, vitamin D₃ can be found in dietary sources; however, it is mainly produced in the skin by 7-dehydrocholesterol stimulation in the keratinocytes by UVB irradiation from sunlight at the wavelength (230–313 nm) (Nussey and Whitehead, 2001).

The hydroxylation of vitamin D₂ and D₃ occurs in the liver by vitamin D 25-hydroxylase and yields the prohormones 25-hydroxyvitamin D₂ (25(OH)D₂) and 25-hydroxyvitamin D₃ (25(OH)D₃ or calcidiol). In the kidney, 1 α -hydroxylase converts the prohormones into active hormones 1,25-dihydroxy vitamin D₂ (1,25(OH)₂D₂) and 1,25-dihydroxy vitamin D₃ (1,25(OH)₂D₃ or calcitriol). 25-hydroxyvitamin D (25(OH)D) conversion to 1,25(OH)₂D (the active form of vitamin D) is being regulated by serum calcium and phosphate, plus parathyroid hormone (PTH). The serum concentrations of 25(OH)D₃ are used to indicate vitamin D stores in the body (Thacher and Clarke, 2011).

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There are many available sources of vitamins D₂ and D₃, including UVB irradiation (vitamin D₃), usual dietary intakes of vitamin D₃ rich foods (mainly oily fish and egg yolks), fortified foods (some dairy products, soy milk, breakfast cereals, and margarine, which generally have vitamin D₂ fortification), and vitamin D₂ and D₃ supplements (Ross et al., 2011). It is advised to achieve 25(OH)D serum concentrations levels above 20 ng/mL (50 nmol/l) (Ross et al., 2011). Thus, to meet the vitamin D requirements of at least 97.5% of the population, Food and Nutrition Board has set Recommended Dietary Allowances for vitamin D to be 600 IU/day for ages 1–70 years and 800 IU/day for ages 71 years and older (Ross et al., 2011).

It is commonly known that the main physiologic function of vitamin D is to enhance calcium absorption to preserve sufficient calcium concentrations, which support proper bone mineralization (Khazai et al., 2008). However, the response of the body to vitamin D relies on two pathways: genomic and non-genomic. The former mechanism involves binding 1,25(OH)₂D to the vitamin D receptor (VDR), which is responsible for regulating gene transcription (Berridge, 2015). VDR is omnipresent in various cells such as cardiomyocytes (Condoleo et al., 2021), vascular smooth muscle (Wu-Wong et al., 2006), and endothelium (Jamali et al., 2018). The interaction between VDR and retinoid X receptor (RXR) creates a heterodimer that attaches to the vitamin D response element (VDRE) responsible for the transcription of vitamin D-sensitive target genes (Carlberg and Molnár, 2015). Also, since VDR was found at the cellular membrane level, it might trigger rapid modifications by epigenetic mechanisms such as DNA methylation or histone modification (Saccone et al., 2015). The non-genomic mechanism can be facilitated by protein disulfide-isomerase A3 (PDIA3), VDR, or both. Vitamin D activates protein kinases such as phosphoinositide 3-kinase, calmodulin-kinase II, and protein-kinase A to mediate calcium influx via L-type voltage-gated calcium channel. The intracellular calcium activates p38 mitogen-activated protein kinases for further downstream signaling without altering gene transcription (Cui et al., 2017).

Due to the complex nature of the mechanisms mentioned above, studies investigating VDR have been trending in the previous 30 years (Abouziid et al., 2021a), and the mechanisms describing its association with various diseases are not entirely justified yet (Abouziid et al., 2018). For cardiovascular diseases (CVD), VDR can hinder endoplasmic reticulum stress and mitochondrial-dys function-dependent apoptosis, which lead to preventing myocardial reperfusion (Yao et al., 2015). Moreover, administering vitamin D decreases myocardial hypertrophy in streptozotocin-induced diabetic rats (Wei et al., 2017). Vitamin D inhibits β-catenin/TCF4/GSK-3β and mTOR signaling in cardiomyocytes – at the molecular level (Wei et al., 2017). The clinical evidences have shown a relationship between CVD, such as left ventricular hypertrophy (Quyyumi and Patel, 2010) and arterial stiffening (Patel et al., 2011). Additionally, vitamin D deficiency (VitDd) is correlated with increased CVD morbidity and mortality (Al Mheid and Quyyumi, 2017). A recent cohort study involving hospitalized patients with heart failure (N = 10,974) showed that vitamin D supplementation reduced in-hospital mortality rates (Kusunose et al., 2021). VitDd was also common in patients discharged to non-home facilities after admission to the hospital due to coronary heart failure, ischemic heart disease, myocardial infarction, atrial fibrillation, and transient ischemic attack (Patel et al., 2020).

2. Materials and methods

2.1. Hypothesis formulation

Many investigations focus only on measuring plasma levels of 25(OH)D as an indicator for vitamin D status assessment in indi-

viduals and its association with clinical disorders and medications (Lertratanakul et al., 2014; Nguyen et al., 2014; Shah et al., 2017; Tandeter et al., 2009). Thus, it is beneficial to quantify vitamin D metabolites to provide us with profound insights toward understanding the correlation between vitamin D and medications, especially those with the same metabolic pathway, particularly in CVD patients (Al Mheid and Quyyumi, 2017; Danik and Manson, 2012; Verdoia et al., 2015; Wang et al., 2017). Moreover, Robien et al. (2013), van Orten-Luiten et al. (2014), and Sohl et al. (2012) have identified several CVD medications that can interact with vitamin D (i.e., atorvastatin, calcium channel blockers, angiotensin-converting enzyme (ACE) inhibitors, thiazide diuretics, angiotensin-II receptor blocker (ARB), platelet aggregation inhibitors, loop diuretics, beta-blockers). Also, as sunlight at the wavelength (230–313 nm) is considered a principal source to synthesize vitamin D, we expect different seasons to considerably affect the plasma level of 25(OH)D. Consequently, this experiment aims to:

- (1) Identify the impact of the seasonal changes on 25(OH)D₃ and 25(OH)D₂ levels in CVD patients.
- (2) Identify the interactions of 25(OH)D₃ and 25(OH)D₂ with CVD medication.

2.2. Study population

Fifty-eight patients (forty-nine male and nine female, 62.9 ± 8.2 years) participated in the study. We excluded patients with malignant tumors, impaired kidney function, acute myocardial infarction, or administering vitamin D supplements.

Various lab reports were used to identify the health status of the enrolled patients, including complete blood count, lipid panel, liver panel, prothrombin time, and hemoglobin A1C. Patients' clinical conditions and medications are presented in Table 1 and Supportive/Supplementary Material (Table S1).

2.3. Sample collection

The blood samples (N = 116) were collected in two different seasons, autumn-winter or spring-summer. Seasonal analysis of the metabolites was performed with reference to the seasons observed in Poland (Radomski, n.d.). During the morning hours, 7:30 to 8:30, and under standard conditions, we obtained an aliquot of 5 mL of blood into ethylenediaminetetraacetic acid (EDTA) tube. Samples were centrifugated for 10 min at a speed of 1620×g for proper separation of plasma and stored at –176 °Fahrenheit (°F) until the UPLC assessment.

2.4. Determination of 25(OH)D₂ and 25(OH)D₃ concentrations

Vitamin D₂ and vitamin D₃ levels were assessed by measuring the concentration of 25(OH)D₂ and 25(OH)D₃, respectively, using the validated UPLC-MS/MS method developed based on our previous published assay (Abouziid et al., 2020). In brief, we mixed 200 μL of the patients' plasma with 20 μL of methanol, followed by adding 20 μL of D₆-25-hydroxyvitamin D₃ (internal standard) and 200 μL LC/MS grade water. After vortexing for 10 s, we added 400 μL of methanol-isopropanol solution (80:20, v/v). The solution was vortexed for 15 s to allow the complete protein precipitation. Liquid-liquid extraction was performed twice using hexane – each time, 1000 μL of hexane was followed by 3 min of mixing, then centrifugated for 10 min (3000 g, 68 °F). The organic layer was separated into clean glass vials. It was evaporated at 113 °F under vacuum, and the remaining residue was dissolved in 200 μL of methanol–water (80:20, v/v) solution. We injected only 10 μL of this final solution directly into the UPLC Nexera coupled to a triple

Table 1
Characteristics of patients. Values are given as number of patients N (%) or mean ± SD.

Characteristic	N (%)
<i>Sex & demographic data</i>	
Male	49 (84)
Female	9 (16)
Age [years]	62.91 ± 8.21
Weight [kg]	83.82 ± 14.80
Height [m]	1.71 ± 0.08
BMI [kg/m ²]	28.66 ± 3.83
<i>Diagnosis</i>	
Hypertension	41 (71)
Hypercholesterolemia	34 (58)
Diabetes	21 (36)
Heart failure	6 (10)
Hyperlipidemia	4 (9)
Kidney disease	4 (7)
<i>Medication</i>	
Antiplatelet medication	58 (100)
NSAID	58 (100)
Beta-blocker	30 (52)
Atorvastatin	20 (35)
ACE inhibitor	19 (32)
PPI	19 (32)
Rosuvastatin	15 (26)
NaCl	13 (22)
Thiazide diuretic	12 (21)
Ca Channel blocker	11 (19)
Biguanides	7 (12)
ARB	6 (10)
Ezetimibe	5 (8)
Steroidal antiandrogen	4 (7)
Cytoprotective anti-ischemic agent	4 (7)
Insulin	4 (7)

Abbreviations: ARB - angiotensin II receptor blockers; ACE - angiotensin-converting enzyme; NSAID - non-steroidal anti-inflammatory drugs; PPI - proton pump inhibitor.

quadrupole mass spectrometer LCMS-8030. Kinetex 2.6 μm F5 analytical column (50 mm × 2.1 mm) was used to separate the analytes. This method prevents the overestimation of 25(OH)D₂ and 25(OH)D₃ by separating 3-epi-25(OH)D₃ – en epimeric form (Abouزيد et al., 2020).

2.5. Statistical analysis

Statistica 13.3 with Plus Kit 3.0 were used to carry out the statistical assessment (TIBCO Software Inc., Tulsa, OK, USA). Shapiro-Wilk test was used as a plot option to test the normality of vitamin D levels, demographic data, and biochemical profiles. Continuous variables were summarized using descriptive statistics; normal variables were represented as mean and standard deviation (SD) and non-normal variables as the median and interquartile range (IQR). Student’s t-test (paired) or Mann-Whitney U test was applied to compare the differences between two related groups such as patients administered vs. not administered drugs and plasma levels of 25(OH)D₃ and 25(OH)D₂ in autumn-winter vs. spring-summer periods. Kruskal Wallis test supplied with Bonferroni adjustment used for multi-group (inter-seasonal) comparison. Correlation between normally distributed continuous variables was expressed as Pearson’s correlation coefficient. Otherwise, the correlation was presented as a non-parametric Spearman’s rank correlation coefficient. The logistic regression and multi-factorial results were described as odds ratios (ORs) and 95% confidence interval (95% CI). A P-value < 0.05 was considered statistically significant for all tests.

The sample size for analysis of a significant difference in mean plasma 25(OH)D₂ and 25(OH)D₃ levels between the two seasons was calculated based on the formula:

$$Sample\ Size = \frac{2SD^2 (Z_{\alpha/2} + Z_{\beta})^2}{d^2}$$

25(OH)D ₃	25(OH)D ₂
Sample Size = $\frac{2(7.84)^2 (1.96+0.84)^2}{(5)^2}$	Sample Size = $\frac{2(4.03)^2 (1.96+0.84)^2}{(5)^2}$
Sample Size = 38.55	Sample Size = 10.19

With SD values of 7.84 ng/ml and 4.03 for 25(OH)D₃ and 25(OH)D₂, respectively, taken from a previous study (Karaźniewicz-Łada et al., 2018), a sample size of at least 38 and 10 participants per season were required for detecting a difference of 5 ng/mL among the two seasons (confidence level = 95%, α = 0.05; statistical power = 80%, β = 0.20).

3. Results

3.1. Vitamin D₃

Vitamin D status was determined based on vitamin D₃ levels; deficiency (<12 ng/mL), insufficiency (12–20 ng/mL), sufficient (>20 ng/mL) and recommended (30–50 ng/mL) (Ross et al., 2011).

25(OH)D₃ average levels were significantly higher in the spring-summer than autumn-winter (14.96 (10.98–25.74) ng/mL vs. 9.12 (3.22–21.48) ng/mL, Z = 3.21; P = 0.001). Besides, in-depth seasonal analysis shows that vitamin D₃ average levels vary between the four seasons (P = 0.007). The concentrations in summer were significantly higher than in autumn and winter (P = 0.042, P = 0.029, respectively) (Fig. 1).

Moreover, VitDd was three times more to occur in autumn-winter (OR = 3.2; P = 0.01), and the concentrations of vitamin D were 3.5 times increased from deficient to insufficient levels in the spring-summer period (OR = 3.52; P = 0.015) (Table 2).

3.2. Vitamin D₂

25(OH)D₂ average concentrations were numerically lower in spring-summer than autumn-winter, but with no significant difference (1.34 (0.45–1.81) ng/mL vs. 1.887 (1.07–3.05) ng/mL; Z = 1.04; P = 0.3). Besides, 25(OH)D₂ average concentrations were comparable across the four-season analysis (Fig. 1).

3.3. Demographic

Overall, males and females had comparable 25(OH)D₃ and 25(OH)D₂ concentrations. Even though males had numerically higher 25(OH)D₃ and 25(OH)D₂ levels in autumn-winter, but the differences were not significant. Gender seasonal analysis is shown in Supportive/Supplementary Material, Table S2.

Moreover, older patients had lower 25(OH)D₃ levels and this correlation was significant (R = -0.232; P = 0.03). Multi-factorial logistic regression shows that increasing age predisposes to develop VitDd in autumn-winter [OR = 1.08; 95%CI = 0.01–0.13; P = 0.011] and (OR = 0.32; 95%CI = -2.07–(-0.22); P = 0.015] respectively (Table 2).

3.4. Medications

The relationship between administered medications (Table 1) and 25(OH)D₃ and 25(OH)D₂ concentrations was studied. Several drugs show interaction with 25(OH)D₃ levels: insulin, spironolactone, thiazide diuretics, ARB, 0.9% sodium chloride (saline or NaCl), and cytoprotective anti-ischemic agent (Trimetazidine (TMX)).

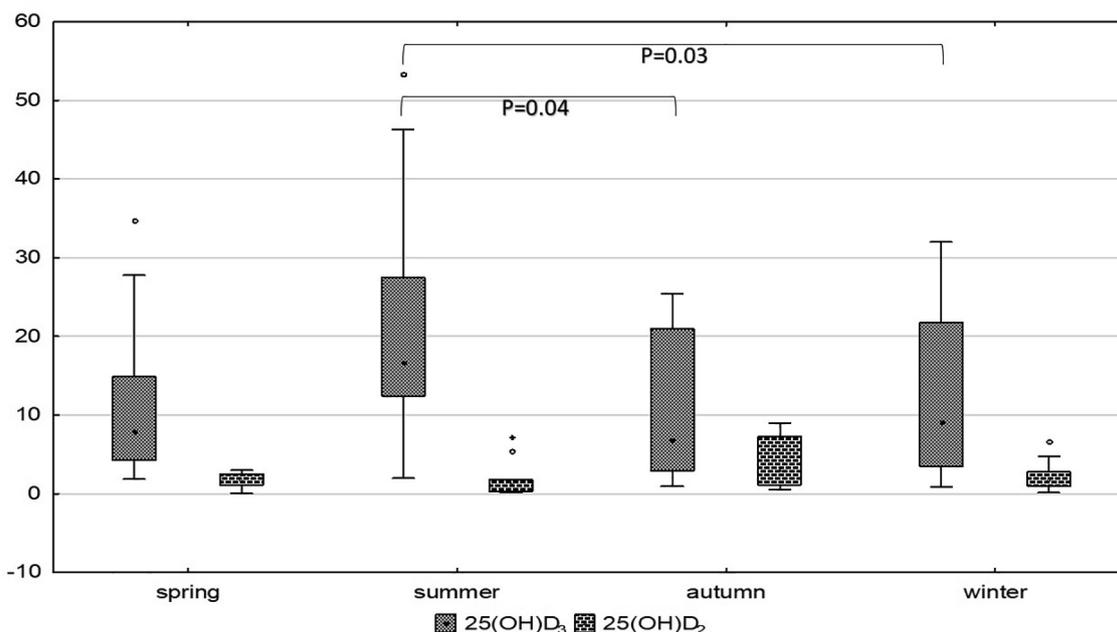


Fig. 1. Vitamin D₃ and vitamin D₂ concentrations - represented by 25(OH)D₃ and 25(OH)D₂, respectively - among different seasons. Kruskal Wallis test supplied with Bonferroni adjustment and showed significantly high 25(OH)D₃ levels in summer compared to winter (P = 0.03) and autumn (P = 0.04). 25(OH)D₂ levels are numerically high in autumn without significance.

Table 2
Multi-factorial logistic regression models.

	Intercept	Standard error	Wald chi-square	Sig.	Exp(B)	95% C.I.for EXP(B)	
						Lower	Upper
Heart Failure							
Age	0.356	0.156	5.180	0.023	1.427	0.049	0.662
Weight*BMI	-0.080	0.038	4.582	0.032	0.923	-0.154	-0.007
Hypercholesterolemia*BMI	1.237	0.557	4.939	0.026	3.445	0.146	2.328
Weight	2.424	1.152	4.427	0.035	11.291	0.166	4.682
Hypertension							
Obesity*Hypercholesterolemia	3.094	1.570	3.881	0.049	22.057	0.016	6.172
Overweight*smoking	4.511	1.324	11.605	0.001	91.004	1.916	7.106
BMI	-1.318	0.409	10.368	0.001	0.268	-2.120	-0.516
Overweight*diabetes	0.014	0.758	0.000	0.985	1.014	-1.471	1.500
Hypercholesterolemia	-2.434	0.921	6.979	0.008	0.088	-4.240	-0.628
Weight	0.138	0.067	4.317	0.038	1.148	0.008	0.269
Vitamin D insufficiency							
TMX*HDL	1.689	0.698	5.865	0.015	5.417	0.322	3.057
Vitamin D deficiency							
Spring-summer	-1.148	0.472	5.916	0.015	0.317	-2.072	-0.223
Age	0.072	0.031	5.271	0.022	1.075	0.011	0.134
Vitamin D deficiency & medication							
NaCl	-1.609	0.778	4.278	0.039	0.2	-3.133	-0.084
Steroidal antiandrogen*Age	0.039	0.017	5.255	0.022	1.039	0.006	0.072
ARB	2.014	0.895	5.057	0.025	7.491	0.259	3.769
Thiazide diuretic	-2.575	1.054	5.972	0.015	0.076	-4.64	-0.51
Vitamin D levels in spring-summer§							
Deficiency [<12 ng/mL]	-1.164	0.457	6.494	0.0108	0.312	-2.059	-0.269
Insufficiency [12–20 ng/mL]	1.257	0.518	5.882	0.0153	3.516	0.241	2.274
Sufficient [>20 ng/mL]	0.138	0.450	0.094	0.7589	1.148	-0.744	1.020
Vitamin D levels in autumn-winter§							
Deficiency [<12 ng/mL]	1.164	0.457	6.494	0.0108	3.202	0.269	2.059
Insufficiency [12–20 ng/mL]	-1.257	0.518	5.882	0.0153	0.284	-2.274	-0.241
Sufficient [>20 ng/mL]	-0.138	0.450	0.094	0.7589	0.871	-1.020	0.744

§- Simple Logistic regression.

Diabetic patients (type 1) on insulin treatment seem to have lower 25(OH)D₃ levels (Z = 2.57; P = 0.01). Moreover, patients administered spironolactone had lower 25(OH)D₃ levels (Z = 2.37; P = 0.02). Thiazide diuretics were associated with higher 25(OH)D₃ levels (Z = 2.25; P = 0.02). Finally, patients administered saline had higher 25(OH)D₃ levels (Z = 2.73; P = 0.006) (Table 3).

Besides, simple logistic regression shows that VitDd was associated with the administration of steroidal antiandrogen (OR = 5.63; P = 0.043), ARB (OR = 3.9; P = 0.041), insulin (OR = 11.5; P = 0.027). Thiazide diuretics and saline were not frequently administered in the case of vitDd (OR = 0.22; P = 0.02) and (OR = 0.25; P = 0.023), respectively. Moreover, a multi-factorial model (Table 2) was built

Table 3
The association between 25(OH)D₃ and 25(OH)D₂ plasma concentrations (ng/mL) and administered medications.

Medications	25(OH)D ₃ ^σ			25(OH)D ₂ ^σ		
	N ₁ N ₂	Concentrations (ng/mL)	P-value	N ₁ N ₂	Concentrations (ng/mL)	P-value
Antiplatelet	5	9.43 (6.52–21.49)	0.871	2	3.29 (3.05–3.52)	0.099
	71	13.33 (4.75–21.78)		46	1.37 (0.84–2.51)	
NSAID	5	9.43 (6.52–21.49)	0.871	2	3.29 (3.05–3.52)	0.099
	71	13.33 (4.75–21.78)		46	1.37 (0.84–2.51)	
Steroidal antiandrogen	68	14.75 (6.59–22.22)	0.015	45	1.38 (0.85–2.64)	0.779
	8	3.54 (2.44–9.71)		3	1.67 (0.54–1.78)	
ACE inhibitor	45	14.49 ± 10.87	0.764 ^β	30	1.65 (0.85–2.51)	0.792
	31	15.25 ± 10.57		18	1.29 (0.84–2.79)	
ARB	65	15.33 ± 10.21	0.207 ^β	40	1.35 (0.7–2.63)	0.193
	11	11.7 ± 13.26		8	1.76 (1.52–4.6)	
Beta-blocker	21	15.19 (9.43–21.57)	0.282	12	1.85 (1.04–2.92)	0.533
	55	12.47 (3.46–21.78)		36	1.5 (0.7–2.29)	
Bisphosphonates	65	12.43 (5.08–21.49)	0.521	42	1.65 (0.85–2.62)	0.891
	11	16.59 (2.92–25.03)		6	1.28 (0.84–3.36)	
Ca Channel blocker	58	14.27 ± 9.77	0.437 ^β	36	1.5 (0.7–2.71)	0.897
	18	16.53 ± 13.39		12	1.51 (1.15–2.22)	
Cytoprotective anti-ischemic agent	69	13.33 (4.29–22.67)	0.792	44	1.5 (0.92–2.63)	0.928
	7	12.78 (11.86–17.3)		4	1.15 (0.41–4.51)	
Insulin	69	15.83 ± 10.62	0.007^β	45	1.38 (0.85–2.62)	0.657
	7	4.66 ± 3.91		3	1.78 (0.54–6.69)	
NaCl	54	12 (3.46–21.15)	0.005	37	1.67 (1.07–2.64)	0.106
	22	14.92 (12.15–34.81)		11	1.21 (0.17–1.74)	
PPI	43	15.28 ± 10.7	0.657 ^β	32	1.37 (0.98–2.63)	0.991
	33	14.17 ± 10.79		16	1.71 (0.55–2.94)	
Atorvastatin	26	13.78 ± 9.23	0.335 ^β	18	1.69 (0.85–2.51)	0.819
	35	16.6 ± 12.44		22	1.35 (0.99–2.64)	
Rosuvastatin	32	17.52 ± 12.47	0.12 ^β	19	1.35 (0.99–2.98)	0.936
	29	13.05 ± 9.22		21	1.67 (0.85–2.06)	
Thiazide diuretic	57	11.85 (3.63–20.85)	0.023	35	1.38 (0.85–2.64)	0.693
	19	15.19 (12.44–27.49)		13	1.7 (0.55–1.81)	
Ezetimibe	32	7.87 (3.47–20.45)	0.778	22	1.69 (1.07–2.64)	0.595
	8	14.6 (2.65–21.67)		6	2.2 (0.56–6.69)	

N₁ – Number of patients who did not administer the medication.

N₂ – Number of patients administered the medication.

σ – Differences were calculated using Mann-Whitney U test and data associated are described as median (IQR) and.

β – Differences were calculated using independent t-test, and the associated data are described as mean ± SD.

for VitDd and previously mentioned medications and confirmed such association, especially with increased age (OR = 1.04; 95%CI = 0.006–0.072; P = 0.02). Finally, TMX*HDL (high-density lipoprotein) was (OR = 5.4; 95%CI = 0.332–3.057). Full single factor logistic regression between vitamin D levels and medication is shown in (Fig. 2). We did not observe any significant relationship between 25(OH)D₂ and administered medications.

4. Discussion

The study aimed to measure 25(OH)D₂ and 25(OH)D₃ concentrations in blood plasma and to assess their relation to the season and applied treatment of CVD patients. Despite the international guidelines suggesting a minimum vitamin D level of 20 ng/mL (Ross et al., 2011), European guidelines advise maintaining vitamin D levels between 30 and 50 ng/mL for adults and children (Płudowski et al., 2013). Thus, in our study, over half of the enrolled patients (66%) had 25(OH)D₃ levels below 20 ng/mL, which indicates vitamin D insufficiency. Almost 25% of patients were in the suboptimal status as 25(OH)D₃ levels were between 20 and 30 ng/mL. Only 9% of patients had optimal 25(OH)D₃ levels between 30 and 50 ng/mL.

4.1. 25(OH)D₃ and 25(OH)D₂ seasonal analysis

Our results are similar to those reported by other authors who investigated vitamin D status in different seasons despite the vari-

ation in the samples (Costanzo et al., 2011; González-Parra et al., 2012; Kashi et al., 2011; Rockell et al., 2008; Shoben et al., 2011). In this study, in the spring-summer period, 25(OH)D₃ average levels were significantly higher than in autumn-winter.

In an assessment performed in Hungary on elderly patients (N = 1307) by Vásárhelyi et al. (2011), seasonal 25(OH)D levels were age-dependent; higher age group was associated with fewer differences. The elderly from 50 to 69 years have 10 ng/mL higher 25(OH)D levels in summer. Those from 70 to 89 years recorded an increase by 5 to 6 ng/mL, suggesting moderate to severe VitDd in the evaluated patients. Płudowski et al. (Płudowski et al., 2016) have investigated vitamin D status in Polish adults (N = 5775). Seasonal analysis revealed that VitDd was common among young adults, men, and participants with higher BMI during winter and spring. Similarly, the British cohort study by Hyppönen and Power (Hyppönen and Power, 2007) (N = 7437) highlighted that for 45-year adults, winter and spring seasons had the highest hypovitaminosis incidents [25(OH)D < 10 ng/mL (15.5%); < 16 ng/mL (46.6%); < 30 ng/mL (87.1%)]. An additional Swedish cohort study by Klingberg et al. (2015) (N = 540) declared that in the spring-summer period, the population with an average age of 41 ± 13 years had the highest 25(OH)D levels. Interestingly, they highlighted the weaker impact of vitamin D supplements on 25(OH)D levels compared to sun exposure.

Even though the above studies declared the prevalence of VitDd in the winter-spring period, results from other studies represented stable vitamin D status throughout different seasons, even in a sunny climate (Costanzo et al., 2011; González-Parra et al., 2012;

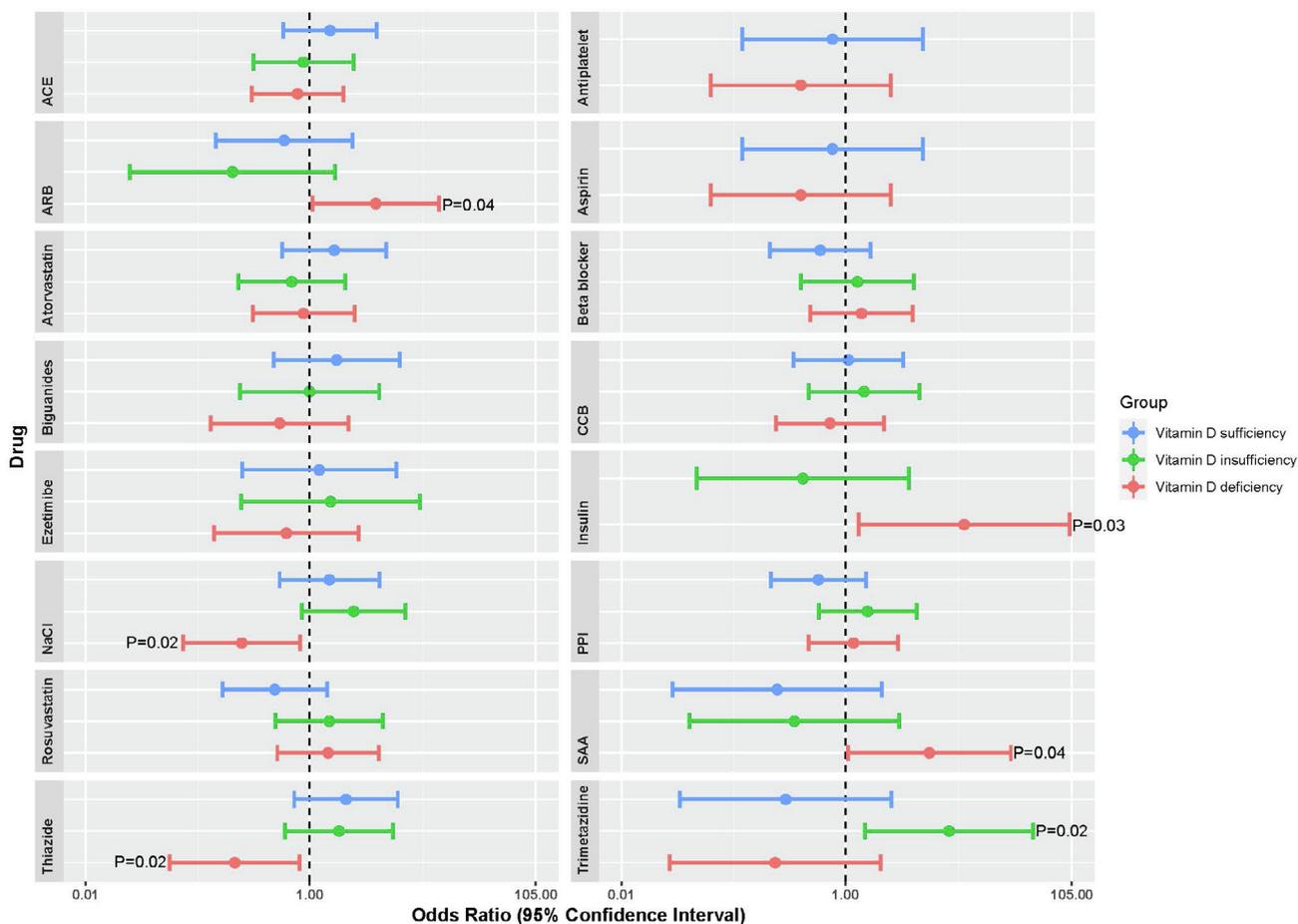


Fig. 2. Single-factor logistic regression between 25(OH)D₃ levels and medication in three groups according to 25(OH)D₃ levels: deficiency (<12 ng/mL), insufficiency (12–20 ng/mL), sufficiency (>20 ng/mL). Data is presented in a forest plot showing the odds ratio (log scale) (circle) and 95% CI (horizontal line). Abbreviations: ACE - angiotensin-converting enzyme inhibitors; ARB - angiotensin II receptor blockers; CCB - calcium channel blocker; PPI -proton pump inhibitor; SAA - steroidal antiandrogen. Raw data is available at [Supportive/Supplementary Material, Table S3](#).

Heidari et al., 2010; Kashi et al., 2011; Rajakumar et al., 2011; Rockell et al., 2008). These observational studies highlight the fact that seasonal changes are not the only factor that alters vitamin D status and extends the research scope to study other reasons contributing to altering vitamin D status, such as demographic data or socioeconomic status (Carnevale et al., 2001; Farrar et al., 2011; Macdonald et al., 2008; Shea et al., 2011). According to Hughes et al. (2011), the concentration of UVB is affected by geographic location and latitude. UVB has inadequate strength for synthesizing sufficient vitamin D in places with maximum latitude during autumn and winter, which may explain vitDd in higher latitudes regions in different seasons (Hughes et al., 2011). Moreover, during winter, people spend more time inside their houses; even if they go outside, they usually wear more clothes which may reduce the ability to synthesize vitamin D₃, hence, influencing 25(OH)D concentrations over the time (Jääskeläinen et al., 2013). Vitamin D seasonal changes could also be associated with single nucleotide polymorphisms of VDR gene: *Apal* (rs7975232), *FokI* (rs2228570) and *BsmI* (rs1544410) (Abouzi et al., 2021b).

In contrast to 25(OH)D₃, 25(OH)D₂ average levels were numerically lower in the spring-summer period than in the autumn-winter period without significant difference (Fig. 1). Thus, these results would imply that the nutritional intake of vitamin D₂ varies by season in a population with measurable levels of 25(OH)D₂. To our knowledge, there are no available literature records on the seasonal change of the plasma 25(OH)D₂ concentrations.

4.2. Demographics

In this study, the patients aged 40–80 years with a mean of 62.91 ± 8.21 years. The higher age was significantly related to low 25(OH)D₃, which may be caused by a low ability of the skin to synthesize 25(OH)D in the elderly population (Heidari et al., 2010; Shea et al., 2011). Moreover, numerically higher 25(OH)D₃ and 25(OH)D₂ levels were noticed in males than in females, but the differences were insignificant. Other studies with a higher number of females have confirmed that females had lower 25(OH)D concentrations which can be linked to higher bone turnover markers in winter than in summer in postmenopausal women (Carnevale et al., 2001; Levis et al., 2005; Macdonald et al., 2008; Wang et al., 2017).

4.3. Medications

Several impacts of the medications used by patients that could alter 25(OH)D levels were observed.

4.3.1. Steroidal antiandrogen

The significant relationship between 25(OH)D₃ levels with steroidal antiandrogen may be explained by the drug's induction of CYP3A4-mediated metabolism of vitamin D. On the other hand, 25(OH)D₃ was reported to blunt the systemic renin-angiotensin-aldosterone system (RAAS) (Carrara et al., 2014). Moreover, calcitriol *in vitro* directly impacts adrenal cortical NCI-H295R cells

via enzymes down-regulating in the steroidogenesis pathway. 25(OH)D₃ with a reduction in parathyroid hormone (PTH) could reduce the serum aldosterone in patients with heart failure regardless of significant clinical benefits (Lundqvist et al., 2010).

4.3.2. ARB

Short-term (i.e. two weeks) ARB co-treatment shows no interaction between ARB and PTH (Bislev et al., 2018). Furthermore, 12 weeks of treatment with telmisartan did not alter 25(OH)D levels in patients with hypertension (N = 31) (Pérez-Castrillón et al., 2012). However, the prolonged use of ARB was shown to significantly reduces PTH levels (Brown et al., 2014; Koiwa et al., 2012; Rossi et al., 1995). Since PTH being a primary stimulator in vitamin D synthesis (Khundmiri et al., 2016), and our patients were on ARB for more than at least 6 months, we noticed a decline in 25(OH)D₃ levels in patients with ARB, and this could be due to ARB inhibitory effect on PTH levels. Current research performed by Turin et al. (Turin et al., 2018) reported that ARB benefit was attenuated in VitDd since both are involved in RAAS inhibition. This can explain such a relationship between co-treatment of ARB with VitDd (Turin et al., 2018).

4.3.3. Insulin

Insulin impact on vitamin D is not well defined. However, VitDd could contribute to developing type 1 diabetes and type 2 diabetes (Johnson et al., 1994). Vitamin D directly stimulates insulin secretion through VDR presence in β-cells of the pancreas (Bland et al., 2004) and their expression of the 1-α-hydroxylase enzyme (Johnson et al., 1994). Besides, 1,25(OH)₂D₃ is capable of transcriptional activation of the human insulin receptor gene; hence, it has a vital role in insulin secretion (Maestro et al., 2002). Chiu et al. (2004) and Scragg et al. (2004) noticed a correlation between insulin insensitivity and vitamin D.

4.3.4. Saline

The observed significant association between saline and 25(OH)D₃ was confirmed by Cervellin et al. (2015). They observed lower vitamin D levels in clinical cases of hyponatremia. It is known that saline solution is prescribed for fluid loss treatment and restoration of sodium chloride balance (Li et al., 2016). In our study, all the patients (N = 22) who received saline had normal sodium levels (135 to 145 mmol/L) due to frequent monitoring. Moreover, according to Vaidya and Forman (2010), a loading dose of table salt was linked to increased 1,25(OH)D concentrations.

4.3.5. Thiazides

Thiazides increase calcium absorption from the luminal membrane into the interstitial in exchange for sodium (Akbari and Khorasani-Zadeh, 2020). This results in inhibition of parathyroid hormone, which decreases 1,25(OH)₂D₃ levels (Khundmiri et al., 2016). Thiazide impact on the renal tubules is being followed by a secondary alteration in vitamin D metabolism, hence, there is a rise in both 25(OH)D₃ and 24,25-dihydroxycholecalciferol (24,25(OH)₂D). Therefore, as a result of thiazide administration, overall 25(OH)D remains unchanged despite higher 25(OH)D₃ (Akbari and Khorasani-Zadeh, 2020).

4.3.6. TMX

According to Sentex et al. (1998), TMX could interfere with phospholipids metabolism in the cardiac myocytes to increase phosphatidylinositol turnover and redirect cytidine triphosphate. These changes boost phospholipid levels and shift fatty acids utilization in the heart in exchange for its availability for energy production. Tuunanen et al. (2008) mentioned that treatment with TMX increased circulating lipid profile by increasing HDL cholesterol levels by 11% in CVD patients. In our study, there was a 23%

increase in HDL (P = 0.024) in the group administered TMX (data not presented). Administration of TMX did not show any significant changes in 25(OH)D₃ levels. Besides, no significant correlation was confirmed between HDL levels and 25(OH)D₃ in this group of patients. Besides, in multi-factorial models, only the TMX*HDL relationship was confirmed (Table 2). Therefore, in our results, we cannot relate 25(OH)D₃ levels to TMX co-treatment due to the low sample size.

4.4. Strength and limitations

This is the first study to report seasonal variation of 25(OH)D₃ and 25(OH)D₂ in cardiovascular patients, revealing a higher prevalence of vitDd during autumn-winter and highlighting the importance of vitamin D supplementation in these groups of patients. Still, there are also limitations. First, the majority of the patients were males. Second, even though the number of patients in this study has exceeded the minimum sample size, future studies should have higher numbers. Third, information on the patients' lifestyle, including physical activities, diet, or socioeconomic status, were not gathered.

5. Conclusion

Vitamin D deficiency was common in the studied CVD patients. We statistically highlighted the influence of seasonal patterns and age on 25(OH)D₃ levels. Smoking, overweight, and combined diagnosis of obesity and hypercholesterolemia were primary risk factors for hypertension, whereas weight and hypercholesterolemia alongside higher BMI were the major risk factors for heart failure. Patients treated with steroidal antiandrogen and insulin had 76% and 67% lower 25(OH)D₃ levels than patients who did not receive the medications, whereas those treated with saline and thiazide had higher analyte concentrations by 24% and 58%, respectively. Further experiments should be performed on a larger number of patients to confirm the results of our study and to identify other factors influencing 25(OH)D concentrations in this group of patients.

6. Data availability

The data used to support the findings of this study are included in the article.

Ethical approval

Written consent was obtained from the patients; the study was conducted according to the guidelines of the Declaration of Helsinki. The ethical approval was granted by the Bioethics Committee of Poznan University of Medical Sciences (protocol no. 273/15, date of approval: 5 March 2015 and protocol no. 58/20, date of approval 16 January 2020).

CRediT authorship contribution statement

Mohamed Abouzi: Validation, Investigation, Data curation, Writing – original draft, Writing – review & editing, Visualization, Project administration. **Franciszek Głowska:** Writing – review & editing. **Marta Karaźniewicz-Lada:** Conceptualization, Methodology, Validation, Formal analysis, Investigation, Data curation, Writing – original draft, Writing – review & editing, Supervision, Project administration, Funding acquisition.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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The abstract of this study was presented at the 16th Warsaw International Medical Congress in the Pharmacy session and was awarded second place.

Conflict of interest

The authors declare that they have no competing interests.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jksus.2022.102187>.

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